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# The Side-Necked Turtle Family Chelidae: A Theory of Relationships Using Shared Derived Characters

EUGENE S. GAFFNEY<sup>1</sup>

#### **ABSTRACT**

The South American and Australian sidenecked turtles of the family Chelidae are analyzed using the shared derived character technique of Hennig. The following hypotheses of monophyly are tested using the characters indicated (see fig. 10):

- Group 1. Family Chelidae (Pseudemydura, Emydura, Elseya, Platemys, Phrynops, Chelus, Chelodina, Hydromedusa)
  - a. Unusually developed lateral cheek emargination
  - b. Loss of quadratojugal
  - c. Loss of mesoplastra
- Group 2. Subfamily Chelinae (Emydura, Elseya, Platemys, Phrynops, Chelus, Chelodina, Hydromedusa)
  - a. Anterior frontal process at least partially separating nasals
- Group 3. Infrafamily Chelodd (Platemys, Phrynops, Chelus, Chelodina, Hydromedusa)
  - a. Symphyseal suture separating lower jaw rami

- b. Dorsal processes of exoccipitals meet medially above foramen magnum
- c. First vertebral scute narrower than second Group 4. Tribe Chelini (Phrynops, Chelus, Chelodina, Hydromedusa)
  - a. Lateral margins of parietals distinctly reduced
- Group 5. Subtribe Chelina (Chelus, Chelodina, Hydromedusa)
  - a. Cervical vertebrae longer than dorsal vertebrae
  - b. Medial portions of jugal and postorbital facing more laterally than posteriorly
- Group 6. Infratribe Hydromedusad (Chelodina, Hydromedusa)
  - a. Posterolateral process of parietal absent
  - b. Extremely reduced horizontal process of parietal
  - c. Quadrate-basisphenoid contact
  - d. Four claws on forefoot

#### INTRODUCTION

The purpose of the present paper is to develop a theory of relationships for the side-necked tur-

tles of the family Chelidae. These turtles are found in South America, with about 12 living

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species usually recognized, and are the dominant continental turtles of Australia, where about 14 living species occur. The chelids are a predominantly carnivorous, fresh-water aquatic family that contains some of the strangest turtles (e.g., Chelus) as well as some of the rarest (e.g., Pseudemydura). There have been few attempts at explicit phylogeny reconstruction for the chelid genera and little osteologic information is available in the literature. My intention here is a phylogeny reconstruction relying primarily on cranial characters.

My method is derived from that of Hennig (1966) and others, and is often termed phylogenetic systematics or cladism. As I see it, this method is the closest approach in systematics to the logical criteria emphasized by Popper (1968) as characterizing science. Popper has argued that the best science is developed in terms of hypothesis formation and test, in which the tests attempt to falsify rather than confirm the hypothesis. In phylogeny reconstruction, hypotheses of relationship are tested by character distributions in which the characters are analyzed in terms of primitive and derived. The analysis consists of a further series of testable hypotheses. Testability, i.e., the potential to criticize and falsify a hypothesis, is the critical feature. The logical aspects of phylogenetic systematics and a summary of the method of using shared derived characters in phylogeny reconstruction are presented elsewhere (Gaffney, In press; Wiley, 1975) and the reader is referred to these works and more general references (Bonde, 1974; Brundin, 1968; Cracraft, 1972, 1974; Eldredge and Tattersall, 1975; Hennig, 1965).

Although fossil chelids are not discussed here, this is due more to the lack of skull material than lack of interest. I am currently engaged in a study of fossil chelids from Australia, including some good skull material of Miocene age. Roger Wood has (in preparation) a study of fossil chelids from South America. Other literature references on fossil chelids may be found in Kuhn (1964) and Mlynarski (1976).

The higher category classification of turtles used here is developed in Gaffney (1975), and the anatomic terminology can be found in Gaffney (1972b).

# ABBREVIATIONS ANATOMICAL

ang, angular pa, parietal art, articular pal, palatine pf, prefrontal bo, basioccipital bs, basisphenoid pm, premaxilla cor, coronoid po, postorbital den, dentary pr, prearticular epi, epipterygoid pt, pterygoid ex, exoccipital qj, quadratojugal fr, frontal qu, quadrate ju, jugal so, supraoccipital mx, maxilla sq, squamosal na, nasal sur, surangular op, opisthotic vo, vomer

#### INSTITUTIONS

AMNH, the American Museum of Natural History, New York

FMNH, Field Museum of Natural History, Chicago

NMNH, National Museum of Natural History, Smithsonian Institution, Washington, D.C.

MCZ, Museum of Comparative Zoology, Harvard University, Cambridge

WAM, Western Australian Museum, Perth

#### ACKNOWLEDGMENTS

Once again I am grateful to Dr. Samuel B. McDowell for allowing me free access to his work on chelids. Dr. Glen Storr of the Western Australian Museum very kindly lent me of Pseudemydura. Dr. Richard specimens Zweifel, the American Museum of Natural History, Dr. George Zug, National Museum of Natural History, Smithsonian Institution, Dr. Hymen Marx, Field Museum of Natural History, and Dr. Ernest Williams, Museum of Comparative Zoology, Harvard University, helped in finding and lending me material in their care. Mr. John Goode of Frankston, Victoria, obtained Australian specimens for me and I appreciate his help.

#### PREVIOUS WORK

In the section on Basic Taxa (below) I include some of the more important literature references to particular chelid genera, and here I discuss some of the more pertinent literature concerning

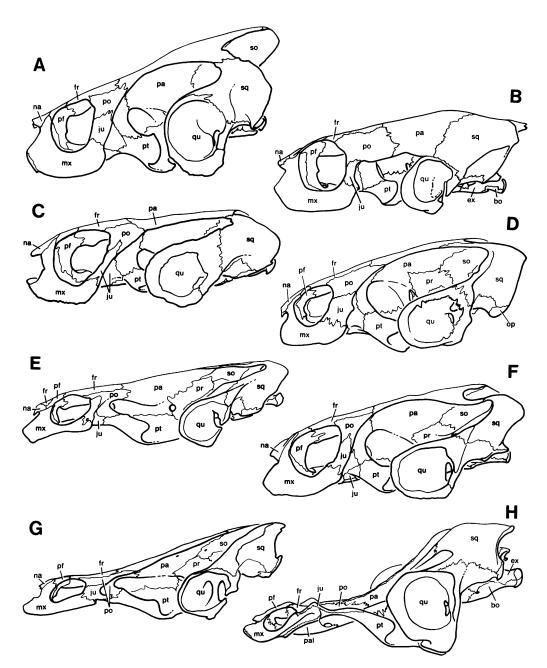


FIG. 1. Lateral views of chelid skulls, measurements are midline condylo-premaxilla length. A. Emydura macquarrii (AMNH 110486; 49 mm.). B. Pseudemydura umbrina (WAM R29341; 35 mm.). C. Platemys platycephala (AMNH 74811; 28 mm.). D. Phrynops geoffroanus (AMNH 79048; 60 mm.). E. Hydromedusa tectifera (FMNH 31032; 51 mm.). F. Phrynops (Mesoclemmys) gibba (FMNH 45669; 37 mm.). G. Chelodina expansa (AMNH 108948; 75 mm.). H. Chelus fimbriata (AMNH 108955; 104 mm.).

phylogeny and reviews of the whole family. Attempts at phylogeny reconstruction involving all or most of the genera in this family are virtually nonexistent. Boulenger (1889) and Grav (1864) presented "natural" keys, which may be interpreted as phylogenies. Burbidge, Kirsch, and Main (1974) come closest to a chelid phylogeny even though their interests were primarily in the Australian forms. On the basis of a phenetic analysis of biochemical data, they conclude that Emydura and Elseya have an ancestor in common not found in common with other three forms. The taxa Pseudemydura. Emydura-Elseya, and Chelodina are equally related; that is, they form a trichotomy in their analysis. Furthermore, some South American taxa were studied serologically leading to a conclusion that the Australian species form one monophyletic group and the South American species form another monophyletic group. In general, phenetic studies using morphological data can be resolved into primitive and derived character states (or convergences, etc.) and a phylogenetic study can then deal with the characters. In the case of serology and related techniques numerical methods have been suggested (e.g., Farris, 1972) to analyze phenetic data in terms of primitive and advanced characters. Nonetheless, I am not convinced of the usefulness of this approach and, in any case, Burbidge, Kirsch, and Main (1974) do not present such an analysis. Although they have produced an interesting phylogeny, albeit distinctly at variance with mine, I cannot point out areas of weakness (or strength) or attempt to resolve it with my character distributions.

There are no detailed reviews of the Chelidae, but Boulenger (1889), Gray (1855, 1870), Siebenrock (1909), and Wermuth and Mertens (1961) are the best sources for lists of species and synonyms. The Australian chelids are reviewed by Goode (1967) who presented keys, figured living specimens and types, and provided a synonymy and bibliography (the last suffers from errors, however). Other important studies of the Australian taxa are: Boulenger (1888), Burbidge, Kirsch, and Main (1974), Cogger (1975, distribution maps), de Rooij (1915), Ogilby (1905), Waite (1929), and Worrell (1963); whereas Blackmore (1969), Strauch (1890), and

Werner (1909) provided more limited information.

The South American chelids are more poorly known than the Australian forms and there are no reviews of them. In addition to the works listed above, useful studies dealing with more than one genus are Froes (1957), Luederwaldt (1926), Siebenrock (1904), and Strauch (1890).

# **BASIC TAXA**

The theory of chelid relationships developed here, uses seven generic level taxa as the fundamental units in the hypothesis. The logical nature of this hypothesis requires that these basic taxa be strictly monophyletic, but rigorous tests of monophyly, including studies of species distribution, etc., are beyond the scope of this paper. I do find it necessary, however, to make a partial examination of this problem.

My principle difficulty is the small sample of specimens available for the named genera, and the absence of any cranial material objectively identifiable for many of the species. I have done my best with the material at hand, but I suspect that some of the characters will be subject to more variation than I have indicated. Nonetheless. I doubt that a more extensive examination of specimens will seriously alter the character distributions as I have reported them. In my opinion, a more important source of further tests is in other organ systems, such as jaw musculature, hyoid apparatus, limb musculature, and limb osteology. Only by increasing the number of areas examined can phylogenetic hypotheses be discarded or substantiated.

# A. Pseudemydura Figures 1B, 2B, 4B, 6B

Specimens Examined. Pseudemydura umbrina, WAM R29341, Twin Swamp Reserve, Western Australia, Australia; Pseudemydura umbrina, WAM R21859, Bullsbrook Reserve, Western Australia, Australia; Pseudemydura umbrina, WAM R29338, Ellenbrook Reserve, Western Australia, Australia.

Discussion. Pseudemydura umbrina has had an interesting taxonomic history, being first named by Siebenrock (1901; figured and described in 1907; these figures are repeated in Williams,

1958) on the basis of one preserved specimen. Subsequently, Glauert (1954) described a form he called *Emydura inspectata*, which Williams (1958) identified as *Pseudemydura* thereby "rediscovering" what is possibly the rarest Recent turtle. Goode (1967) has photographs of living specimens and the Siebenrock type specimen as well as sketches of the skull. Burbidge, Kirsch, and Main (1974) gave good shell and skull figures (including the lower jaw).

I interpret the following features of *Pseude-mydura* as autapomorphies; that is, derived features found only in this species:

- 1. Quadrate-parietal contact (fig. 1B)
- Supraoccipital laterally expanded in contrast to other Chelidae (fig. 2B)
- 3. Parietal laterally expanded (fig. 2B)
- 4. Postorbital ventrolaterally expanded (fig. 2B)
- 5. Anterior extension of squamosal (figs. 1B, 2B)
- Prearticular separating coronoid and splenial (Burbidge, Kirsch, and Main, 1974)
- Medial approximation of maxillae along labial ridge separating or nearly separating premaxillae into anterior and posterior portions (fig. 4B)

The argument that characters 1 through 6 are autapomorphies is developed below in the section on Group 2.

# B. Emydura-Elseya Figures 1A, 2A, 4A, 6A, 7

Specimens Examined. Elseya latisternum, AMNH 103700, Bulimba Creek, Brisbane, Queensland; Emydura macquarrii, AMNH 77637, no data; Emydura macquarrii, AMNH 77648, no data; Emydura macquarrii, AMNH 11487, South Australia, Australia; Emydura macquarrii, AMNH 110486, South Australia, Australia; Emydura macquarrii, AMNH 110488, 40 mi. SE Mildura, Victoria, Australia; Emydura macquarrii, AMNH 108962, Patho, Victoria, Australia; Emydura macquarrii, AMNH 103702, Victoria, Australia; Emydura kreffti, AMNH 72406, no data; Emydura kreffti, AMNH 108958, Queensland, Australia; Emydura australis, AMNH 108957, Darwin area, Northern Territory, Australia.

Discussion. I am treating these two genera as one basic taxon because I have been unable to

differentiate them consistently using cranial characters. However, this may be due to my extremely small sample of adequately identified Elseya skulls. Burbidge, Kirsch, and Main (1974) and Goode (1967) used features of the intergular scute, cervical scute, skull cap, snout, and postorbital skin tuberculation, as well as serology in the case of Burbidge, Kirsch, and Main (*ibid.*) to separate the named species into two genera.

Morphologic information substantiates Emydura-Elseya monophyly. Emydura and Elseya have heavier lower jaws with wider triturating areas and slightly developed symphyseal "hooks" in contrast to all other chelids, although there is some variation in this feature. This would appear to be derived within the Chelidae but the common possession of heavy lower jaws among pelomedusids weakens the use of this feature. Also, megacephaly seems to be common in Emydura and Elseya (Goode, 1967) and may occur in other chelids such as Phrynops. Nonetheless, at present it seems best to treat Emydura and Elseya as a strictly monophyletic assemblage.

Other References. Boulenger (1888, 1889; skull figures); Burbidge, Kirsch, and Main (1974, skull figures); Gaffney (1975, skull figures); Goode (1967, skull figures); Gray (1863a, 1863b, 1872); Hoffmann (1890, skull figures); Krefft (1876); Loveridge (1934); Ogilby (1905); Ouwens (1914); Peters and Doria (1878); Siebenrock (1906, 1907, 1912); Vogt (1911).

# C. Platemys Figures 1C, 2C, 4C, 6C

Specimens Examined. Platemys platycephala, AMNH 74811, no data; Platemys platycephala, AMNH 75101, no data; Platemys platycephala, FMNH 45659, Loreto, Peru.

Discussion. My sample of Platemys skulls consists of three specimens of P. platycephala, which show the following autapomorphous features: truncated and reduced crista supraoccipitalis, lateral edges of parietal parallel and orbits relatively large (presumably correlated with small size of adult animals). Again, the absence of skulls identifiable as P. pallidipectoris, P. spixi, and P. radiolata hamper the usefulness of these criteria.

One of the principle shell features used to

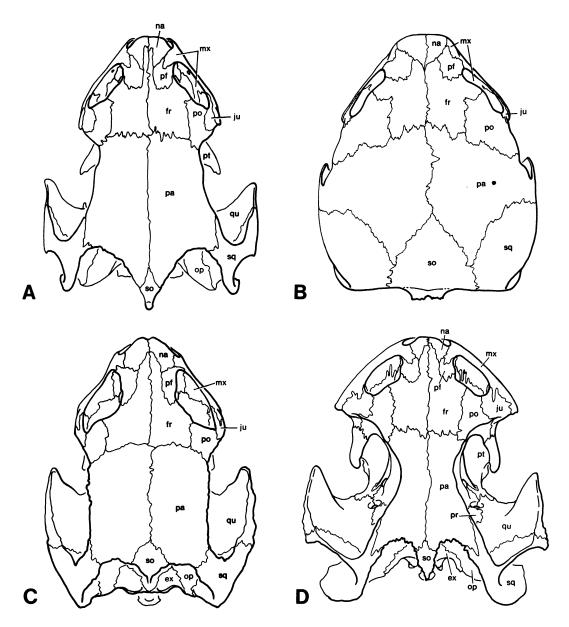


FIG. 2. Dorsal views of chelid skulls. Skull measurements in figure 1. A. Emydura macquarrii (AMNH 110486). B. Pseudemydura umbrina (WAM R29341). C. Platemys platycephala (AMNH 74811). D. Phrynops geoffroanus (AMNH 79048).

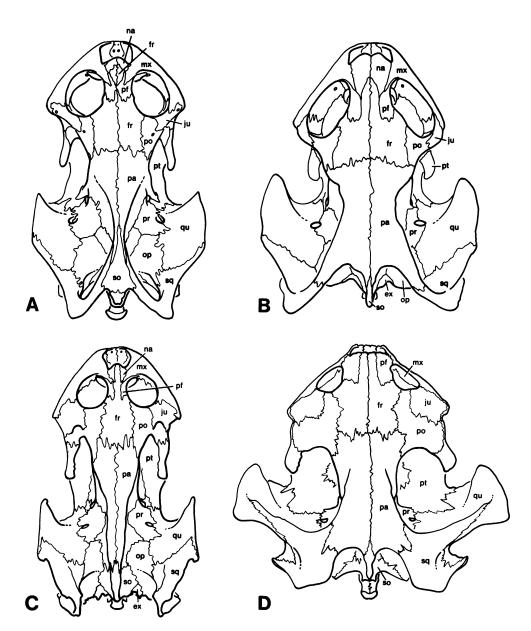


FIG. 3. Dorsal views of chelid skulls. Skull measurements in figure 1. A. Hydromedusa tectifera (FMNH 31032). B. Phynops (Mesoclemmys) gibba (FMNH 45669). C. Chelodina expansa (AMNH 108948). Chelus fimbriata (AMNH 108955).

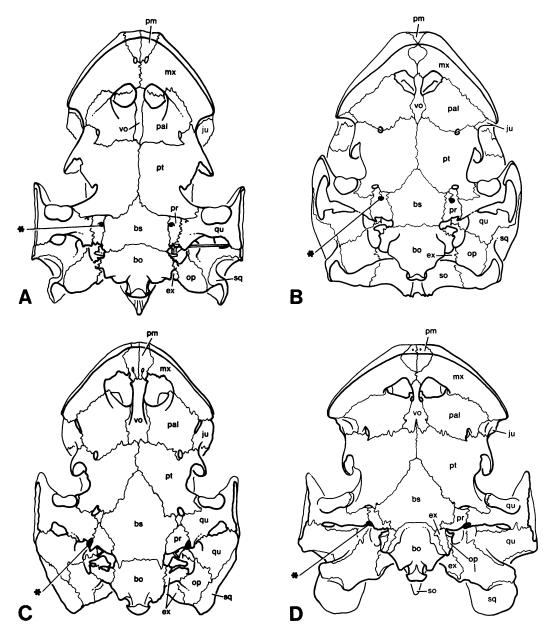


FIG. 4. Palatal views of chelid skulls. Skull measurements in figure 1. Asterisks (\*) show position of foramen posterius canalis carotici interni. A. Emydura macquarrii (AMNH 110486). B. Pseudemydura umbrina (WAM R29341). C. Platemys platycephala (AMNH 74811). D. Phrynops geoffroanus (AMNH 79048).

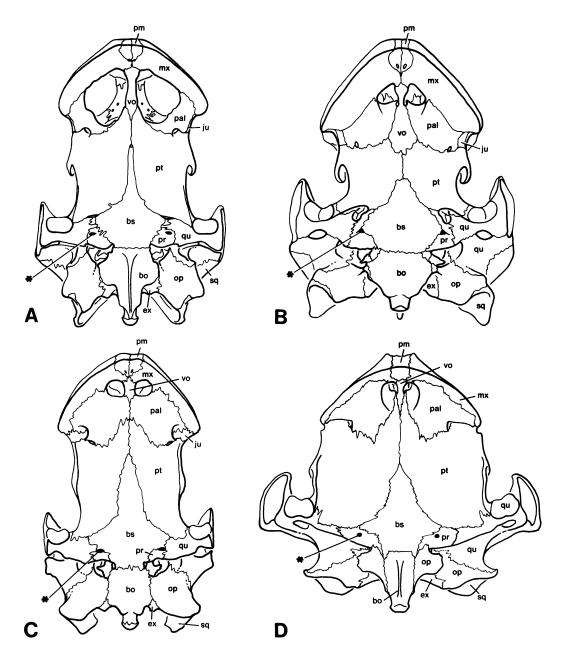


FIG. 5. Palatal views of chelid skulls. Skull measurements in figure 1. Asterisks (\*) show position of foramen posterius canalis carotici interni. A. *Hydromedusa tectifera* (FMNH 31032). B. *Phrynops* (Mesoclemmys) gibba (FMNH 45669). C. Chelodina expansa (AMNH 108948). D. Chelus fimbriata (AMNH 108955).

characterize *Platemys* is the presence of a trough along the carapace midline, but this feature is apparently absent in *Platemys radiolata*, (Freiberg, 1945).

Other References. Dunn (1945); Freiberg (1945, 1947); Froes (1957); Luederwaldt (1926); Medem (1960a); Mertens (1967); Müller (1939); Siebenrock (1897, 1904, skull figures); Wagler (1830).

# D. *Phrynops*Figures 1D, 1F, 2D, 3B, 4D, 5B, 6D, 6F

Specimens Examined. Phrynops geoffroanus, AMNH 79048, no data; Phrynops geoffroanus, AMNH 58201, no data; Phrynops geoffroanus, AMNH 58110, 'Peru; Phrynops (Batrachemys) nasuta, AMNH 108908, no data; Phrynops (Batrachemys) sp., AMNH 58123, Iquitos, Peru; Phrynops (Batrachemys) nasuta, MCZ 58099; Orinoco River, Venezuela; Phrynops (Batrachemys) nasuta, MCZ 1456, Pernambuco, Brazil; Phrynops (Mesoclemmys) gibba, FMNH 45669, Yarinacocha, Loreto, Peru; Phrynops (Mesoclemmys) gibba, FMNH 45671, Yarinacocha, Loreto, Peru.

Discussion. The species here included in this genus have had a particularly confusing history of generic assignment. Some of the earlier aspects may be obtained from Gray (1855, 1864, 1870). In 1909, the taxa involved were classified by Siebenrock (closely following Boulenger, 1889, and Siebenrock's own work of 1904) as follow:

Rhinemys nasuta Mesoclemmys gibba Hydraspis hilarii Hydraspis geoffroyana Hydraspis tuberosa Hydraspis rufipes Hydraspis wagleri

Stejneger (1909) showed that the type species of *Hydraspis* Bell is *Testudo longicollis* Bell, 1828, and as this species was earlier made the type species of *Chelodina* Fitzinger, 1826, the genus *Hydraspis* is a junior synonym of *Chelodina*. He also argued that *Phrynops* Wagler, 1830, is the correct name for the taxon previously

called *Hydraspis*, and, finding *Rhinemys* Wagler, 1830, to be a synonym of *Phrynops*, erected *Batrachemys* to replace it. Therefore, we find later works, such as Wermuth and Mertens (1961) with the following classification (give or take a few species):

Batrachemys nasuta

Batrachemys dahli (erected by Zangerl and Medem, 1958)

Mesoclemmys gibba

Phrynops geoffroanus (with three subspecies: geoffroanus, hilarii, tuberosus)

Phrynops rufipes

Zangerl and Medem (1958), however, in allusion to a study in progress at that time by Williams and Vanzolini (unpublished), stated that the three genera Batrachemys, Mesoclemmys, and Phrynops were closely related and should be placed in one genus, Phrynops, with the three former genera recognized as subgenera. Bour (1973) has argued that even these taxa are not objectively recognizable, but neither he nor Zangerl and Medem (ibid.) gave a diagnosis of Phrynops in the larger sense. As Bour (ibid.) has suggested, the Zangerl and Medem concept of Phrynops is essentially the same as that of Gray's (1864) Hydraspis, and we appear to have come full circle.

For the purposes of this study I use *Phrynops* in Bour's sense, without subgenera, even though I cannot rigorously support its strict monophyly. I have been unable to find unique derived characters in the skull of *Phrynops* but I have also been unable to find derived characters in common between some of the species in *Phrynops* and *Chelodina*, *Hydromedusa*, or *Chelus*. Therefore, at present I can falsify neither the hypothesis that *Phrynops* is monophyletic nor the hypothesis that it is paraphyletic. Even if *Phrynops* is paraphyletic it will not drastically alter the phylogenetic hypothesis advanced here.

Other References. Albrecht (1976, cranial arteries); Boulenger (1889, skull figures); Dunn (1945); Froes (1957); Gray (1873); Kanberg (1926); Luederwaldt (1926); Medem (1960a, 1960b, 1960c, 1966, 1973); Mertens (1967, 1969, 1970); Müller (1939); Siebenrock (1904, 1905); Zangerl and Medem (1958, skull figures).

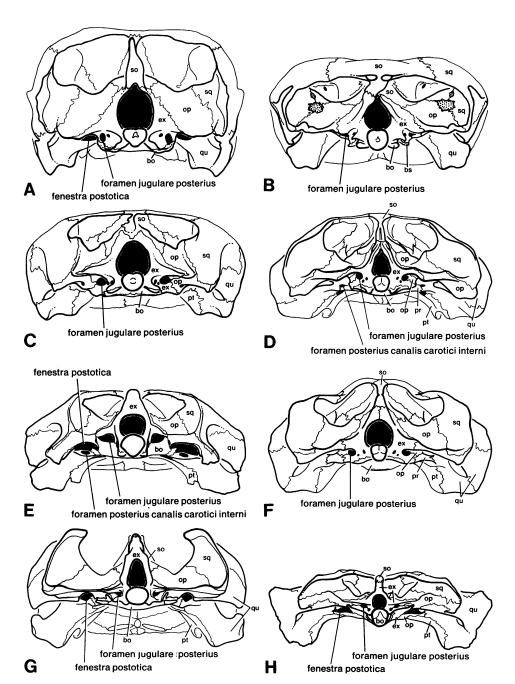


FIG. 6. Occipital views of chelid skulls. A. Emydura macquarrii (AMNH 76199). B. Pseudemydura umbrina (WAM R29341). C. Platemys platycephala (AMNH 74811). D. Phrynops geoffroanus (AMNH 79048). E. Hydromedusa tectifera (NMNH 15189). F. Phrynops (Mesoclemmys) gibba (FMNH 45669). G. Chelodina expansa (AMNH 108948). H. Chelus fimbriata (AMNH 108955).

# E. Chelus Figures 1H, 3D, 5D, 6H

Specimens Examined. Chelus fimbriata, AMNH 108955, no data; Chelus fimbriata, AMNH 111962, no data; Chelus fimbriata, AMNH 6596, no data; Chelus fimbriata, AMNH 43298, no data.

Discussion. Only one Recent species of this genus is usually recognized, but in any case, Chelus is riddled with autapomorphies and looks as if it had been run over by a truck. The following cranial features I interpret as unique derived characters for this taxon:

- 1. Nasals absent (fig. 3D)
- 2. Prefrontal broadly exposed along dorsal margin of apertura narium externa [narrowly exposed in *Chelodina*] (fig. 3D)
- 3. Pterygoids extend anteriorly into apertura narium interna and often separate vomer from palatines (fig. 5D)
- 4. Extreme flattening of skull, particularly in center (fig. 1H)
- 5. Cavum tympani extended laterally to considerable degree (fig. 3D)
- 6. Medial processes of jugal and postorbital lie entirely on external surface of skull [see discussion under Group 5] (fig. 3D)
- 7. Maxilla relatively reduced in exposure on triturating surface so that palatine bears lingual ridge (fig. 5D)

Other References. Boulenger (1889, skull figures); Dunn (1945); Froes (1957); Fuchs (1931, lower jaw figures); Gregory (1946, skull figures); Hoffmann (1890); Luederwaldt (1926); Medem (1960a); Muller (1939); Siebenrock (1897, skull figures); Wagler (1830, skull figures).

# F. Chelodina Figures 1G, 3C, 5C, 6G, 8

Specimens Examined. Chelodina steindachneri, AMNH 101978, Woodstock, Western Australia, Australia; Chelodina longicollis, AMNH 108953, no data; Chelodina longicollis, AMNH 108947, no data; Chelodina longicollis, AMNH 108947, no data; Chelodina longicollis, AMNH

108951, Patho, Victoria, Australia; Chelodina longicollis, AMNH 76569, no data; Chelodina longicollis, AMNH 108952, Patho, Victoria, Australia; Chelodina novaeguineae, AMNH 57589, Mabaduane, Papua, New Guinea; Chelodina novaeguineae, AMNH 86547, Armraynald, Queensland, Australia; Chelodina expansa (?), AMNH 103699, Bulimba Creek, Brisbane, Queensland. Australia; Chelodina expansa, AMNH 108948, Patho, Victoria, Australia; Chelodina expansa, AMNH 108949, Patho, Victoria, Australia; Chelodina rugosa (?), AMNH 104338, Mt. Burnett, Western Australia, Australia; Chelodina rugosa, AMNH 82532, Cape York Peninsula, Queensland, Australia; Chelodina rugosa, AMNH 108954, Darwin area, Northern Territory, Australia.

Discussion. Chelodina has a series of characters that I am hypothesizing as autapomorphies or unique derived characters.

- Nasals usually separated by anterior processes of frontals (fig. 3C)
- 2. Frontals fused along midline (fig. 3C)
- 3. Temporal bar absent (fig. 3c)
- Extensive quadrate-basisphenoid contact (fig. 5C)

Characters 1 and 2 are unique in turtles, although the frontal in *Chelus* reaches the margin of the apertura narium externa the nasals are absent in that form. The temporal bar is absent in some cryptodires but the remaining roofing bones are of distinctly different morphology. An extensive quadrate-basisphenoid contact occurs in pelomedusids (e.g., *Podocnemis*) but again, the morphology of the bones involved is inconsistent with the hypothesis that the contacts are homologous. A limited quadrate-basisphenoid contact occurs in *Hydromedusa* and I hypothesize that the limited condition is primitive for *Hydromedusa* and *Chelodina*.

These characters are consistent with strict monophyly of *Chelodina*. Burbidge, Kirsch, and Main (1974) presented serologic data which they conclude also argues for *Chelodina* monophyly.

Rhodin and Mittermeier (1976) gave good descriptions and figures of the skull in *Chelodina siebenrocki* and their new species, *C. parkeri*. They provided a glossary of anatomical terms

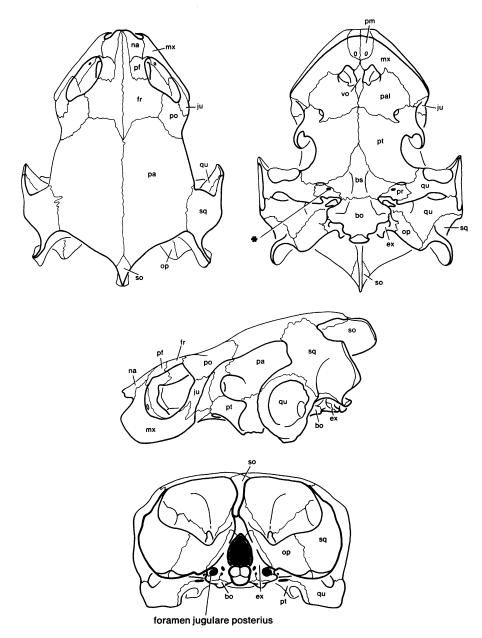


FIG. 7. Elseya latisternum (AMNH 103700; 53 mm.).

used to describe systematically important features.

Other References. Boulenger (1888); Burbidge, Kirsch, and Main (1974, skull figures); Fry (1915); Gaffney (1975, skull figures); Glauert

(1922); Goode (1967, skull figures; 1968); Goode and Russell (1968); Gray (1856, 1869, skull figures); Hoffmann (1890, skull figures); Loveridge (1934); Ogilby (1890, 1905); Schnee (1899); Siebenrock (1897, 1905, 1914, skull figures); Stejneger (1909); Vestjens (1969); Vogt (1911); Waite (1929, skull figures); Werner (1901); Worrell (1961).

# G. Hydromedusa Figures 1E, 3A, 5A, 6E

Specimens Examined. Hydromedusa maximiliani, MCZ 2856, Brazil; Hydromedusa tectifera, NMNH 15189; Hydromedusa tectifera, FMNH 31032.

Discussion. Hydromedusa has the following unique derived features:

- 1. Relatively large bony apertura narium interna formed by reduced ossification of palatine [the fleshy internal narial openings may not be enlarged] (fig. 5A)
- 2. Prefrontals meet in midline and may overlap anterior processes of frontals so that frontals are exposed anterior and posterior to the prefrontal contact (fig. 3A)
- 3. The cervical (nuchal) scute is relatively large and separated from the anterior edge of the shell by medial contact of the first pair of marginal scutes (Boulenger, 1889)

The large bony internal nares and the median prefrontal contact occur in no other chelids (although they do occur in many other turtles), and it seems most parsimonious to consider them independently derived in *Hydromedusa* rather than a primitive retention. The shell feature is unique among turtles.

Wood and Moody (1976) provide figures for the shells and descriptions of the shells in *Hydro*medusa maximiliani and *H. tectifera*, as well as shell characters allowing recognition of these two forms.

Other References: Froes (1957); Hay (1908, skull figures); Luederwaldt (1926); Mertens (1967); Müller (1939); Peters (1839, skull figures); Wagler (1830, skull figures).

## PHYLOGENETIC HYPOTHESIS

The following sections should be read with the cladogram (fig. 10) in mind. The group numbers refer to numbers on the cladogram. A sum-

<sup>1</sup> Note that the recessed nuchal bone with peripheral bones meeting medially found in *Hydromedusa maximiliani* also occurs in the extinct baenoid *Compsemys* (Gaffney, 1972a) and is not unique to *Hydromedusa* as stated by Wood and Moody.

mary of the shared derived characters may be found in the Abstract. See also table 2 for cranial characters discussed here and in the Basic Taxa section.

#### GROUP 1-FAMILY CHELIDAE

Table 1 is a comparison of Pelomedusidae and Chelidae using a series of characters that test monophyly for both families. I have elsewhere (Gaffney, 1975) argued that pleurodires are strictly monophyletic, and that discussion should be consulted. I am including Recent and fossil taxa to the extent that they are available.

The chelids have the following synapomorphies or shared derived characters: (1) Unusually developed lateral cheek emargination; (2) loss of quadratojugal; (3) loss of mesoplastra.

Although *Pseudemydura* has relatively less cheek emargination than the other chelids it still differs strongly from pelomedusids in the shape and number of bones bordering the emargination. Also, there is reason to think that the extensive skull roof in *Pseudemydura* is derived for chelids and that the condition in *Emydura* is the primitive one for chelids. The parietals, squamosals, postorbitals, and supraoccipitals of *Pseudemydura* are different in their extension and shape not only from other chelids but also from pelomedusids. If the *Pseudemydura* pattern were primitive, one would expect fo find it in pelomedusids or cryptodires.

A further aspect of the emphasis in chelids on cheek emargination is the loss of the quadratojugal in all members of this family. Even Pseudemydura, a form possessing a well-developed temporal roof, lacks a quadratojugal, further suggesting that it evolved from a more emarginate ancestor. Some species of Cuora, Hieremys, Geoemyda, and Terrapene also lack a quadratojugal, and in these forms it appears to be associated with well developed cheek emargination.

Chelids lack mesoplastra, the presence of which is presumably primitive for pleurodires. Although mesoplastra were lost independently within the cryptodires, there is no evidence that this has happened more than once within the pleurodires.

#### **GROUP 2-SUBFAMILY CHELINAE**

Pseudemydura has a number of unique features but only one of them would appear to be

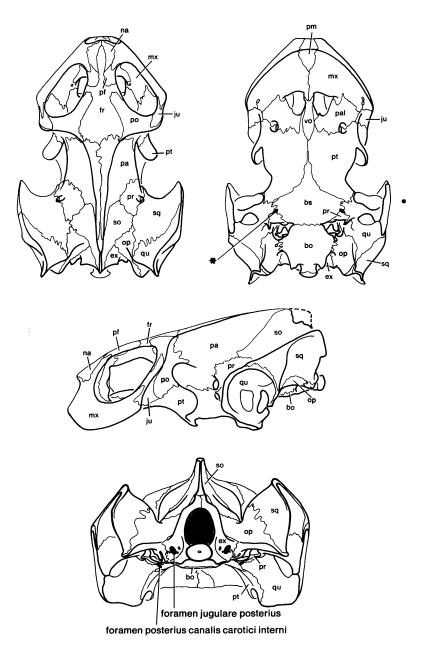


FIG. 8. Chelodina novaeguineae (AMNH 57589; 41 mm.).

primitive for the Chelidae. In all other chelids the frontal has a process that extends anteriorly (with the other frontal) along the median suture to partially separate the nasals. In *Pseudemydura* (figs. 2, 3) the nasals slightly separate the frontals and there is only a short anterior process on the

frontal. No other living turtles possess nasals but in fossil turtles (baenoids, toxochelyids, plesiochelyids, chelosphargine protostegids, *Solnhofia*, and *Proganochelys*) that do have nasals, the frontals do not separate the nasals. Therefore, the condition in all chelids except *Pseudemydura* 

TABLE 1

A Comparison of the Pelomedusidae and Chelidae
(Including information on Recent and fossil forms.)

Character	Pelomedusidae	Chelidae
1. Nasals	absent	present (except in Chelus)
2. Prefrontals	meet in midline	do not meet in midline (except in Hydromedusa)
3. Posterior temporal emargination	varies from virtually absent (e.g., <i>Dacquemys</i> ) to extensive (e.g., <i>Pelomedusa</i> )	usually poorly developed with persistent squamosal-parietal contact (except in Chelodina)
4. Cheek (lateral temporal emargination)	variable, but never developed to the extent seen in Chelidae	developed to an unusual degree with only a parietal- squamosal bar remaining
5. Quadratojugal	present	absent
6. Triturating surface	usually broader	usually narrower
7. Vomer	usually absent	present
8. Splenial	absent	present
9. Mesoplastra	present	absent
10. Cervical (nuchal) scute	absent	present (except in most  Elseya and as an infrequent  variation)
11. Cervical vertebrae <sup>a</sup>	second biconvex	fifth and eighth biconvex

a(Williams, 1950)

would appear to be derived for pleurodires and a useful test for monophyly of the non-Pseudemydura chelids.

I am interpreting the other unique features of *Pseudemydura* as autapomorphies, that is, derived characters found only in this species and, therefore, not useful in phylogeny reconstruction.

This particular hypothesis, namely that Pseudemydura is the sister taxon to the other chelids is perhaps the weakest aspect of my chelid phylogeny, and I would like to discuss some alternatives at this point. One alternative (fig. 9A) would have Emydura and Pseudemydura as sister taxa, that is, with an ancestor in common not in common with any other turtle. However, Pseudemydura and Emydura have few unique characters in common. The cervical vertebrae (particularly the anterior ones) of these two genera do have zygapophyses that are more widely separated than in other chelid genera but this feature is presumably primitive for pleurodires and quite unsatisfactory for corroborating monophyly.

Similarly the second alternative (fig. 9B) also fails from the lack of a derived character in common between *Pseudemydura* and all chelids except *Emydura*.

My contention that Pseudemydura is the sister taxon to all other chelids should not be thought of as an argument that all of its morphology is primitive. The nasal-frontal morphology does seem primitive, but other distinctive features of the skull roof seem to be autapomorphies or advanced features unique to this species. The extensive temporal roof of *Pseudemydura* is best interpreted as a unique derived condition for chelids rather than a primitive one. In fact, comparison with pelomedusids and cryptodires suggests to me that the Emydura type of temporal roof with an extensive lateral or cheek emargination and a shallow posterior emargination is probably primitive for chelids. The absence of a quadratojugal in Pseudemydura is consistent with a hypothesis of expansion of the parietals, postorbitals, and supraoccipital into the emarginated areas. In other words, my hypothesis of relationships

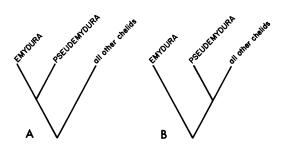


FIG. 9. Alternate hypotheses relating *Emydura*, *Pseudemydura*, and remaining chelids. A third, but preferred alternative, is presented in figure 10. See text for discussion.

would have the primitive chelid possessing most of the features of *Emydura* except the nasal-frontal morphology in which it would resemble *Pseudemydura*.

In the lower jaw of *Pseudemydura* the prearticular extends anteriorly to separate or nearly separate the coronoid and splenial (Burbidge, Kirsch, and Main, 1974, p. 389). In all other chelids the coronoid and splenial have an extensive contact. Few turtles retain a splenial, but in those that do (i.e., *Solnhofia*, baenids, *Plesiochelys*), the coronoid and splenial have a well-developed contact. Therefore, the prearticular extension in *Pseudemydura* is best interpreted as a unique derived character.

#### **GROUP 3-INFRAFAMILY CHELODD**

Platemys, Phrynops, Chelus, Chelodina, and Hydromedusa all have a symphyseal suture separating the two lower jaw rami. Pseudemydura and Emydura have the rami fused as do all other turtles except for Hesperotestudo, fide Bramble, [MS], whereas most reptiles have the rami separated. The appropriate out-group comparison must be with cryptodires rather than other reptiles and I conclude that separate rami are a derived feature corroborating common ancestry of Platemys, Phrynops, Chelus, Chelodina, and Hydromedusa.

Another feature with this distribution involves the dorsal processes of the exoccipitals (fig. 6). In the five genera indicated these dorsal processes extend dorsomedially to meet each other in a sagittal suture above the foramen magnum and prevent the usual exposure of the supraoccipital

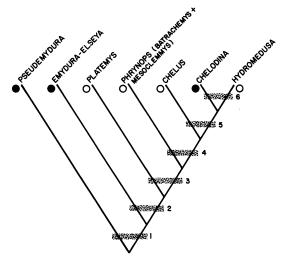


FIG. 10. A theory of relationships of the chelid turtles. The numbers and associated bars refer to groups and character distributions discussed in the text (see Abstract for list). The solid black circles are Australian genera, the open circles South American.

on the dorsal margin of the foramen. In *Emydura* and *Pseudemydura* the exoccipitals do not meet, although they very nearly do so in *Pseudemydura*.

As noted by Boulenger (1889), Emydura and Elseva have the first vertebral scute narrower than the second, whereas the other chelids have the first vertebral wider than the second. Pseudemydura (unknown to Boulenger) also has the first vertebral scute narrower than the second, giving this character the same distribution as the two characters discussed above. A comparison of this condition with pelomedusids unfortunately does not allow a useful test of polarity (primitive vs. derived). Pelomedusids have both conditions although Pelusios and Pelomedusa, forms that I would consider as generally plesiomorphic pelomedusids, have the narrower first vertebral scute. In any case, the character does not falsify the hypothesis advanced here.

## **GROUP 4-TRIBE CHELINI**

In *Phrynops*, *Chelus*, *Chelodina*, and *Hydromedusa* the lateral margins of the parietals are distinctly reduced in comparison to the degree of

emargination seen in *Emydura* and *Platemys* (figs. 2, 3). I interpret this more emarginate condition as derived for the four taxa indicated and suggest that this is consistent with the hypothesis that they are strictly monophyletic. A reduced squamosal-parietal arch also characterizes this group, except *Chelus*.

The above four genera also differ from other chelids in generally having neurals; the other genera usually lack them (table 3; see also Boulenger, 1889 for figures). *Chelodina* has neurals in only one species, *C. oblonga* (Burbidge, Kirsch, and Main, 1974), however. The presence of neurals would presumably be primitive and their absence derived, making this character distribution an important contradiction to my hypothesis. Nonetheless, I am concluding that it is more par-

simonious to accept my hypothesis and reject the neural bone test because the latter is only one character, whereas my hypothesis suggests several. The acceptance of neural absence as being consistent with strict monophyly of Pseudemydura, Emydura-Elseya, and Platemys requires a number of ad hoc hypotheses to invoke character convergence. The rejection of the neural features requires the acceptance of one of the following ad hoc hypotheses: (1) the absence of neurals is a primitive feature for chelids and the presence of neurals in the Tribe Chelini (Group 4) is an example of convergence, or (2) the three generic level taxa lacking neurals lost them independently. Neither hypothesis is supported by other tests and this remains an important problem area.

TABLE 2
Cranial Features of Chelid Genera

	Pseudemydura	Emydura & Elseya	Platemys	Phrynops	Chelus	Chelodina	Hydromedusa
1. Nasals	present	present	present	present	absent	present	present
2. Anterior process of frontal	absent	present	present	present	present	present	present
3. Nasals completely separated by anterior frontal pro- cess	no .	no	no	no	nasals absent	yes	no
4. Prefront- als exposed along dorsal margin of apertura narium ex- terna	no	no	no	no	broadly	narrowly	no
5. Prefrontals meet in midline so that frontals are exposed anterior and posterior to prefrontal contact	no	no	no	no	no	no	yes

# GAFFNEY: CHELIDAE

TABLE 2 – (Continued)

	Pseudemydura	Emydura & Elseya	Platemys	Phrynops	Chelus	Chelodina	Hydromedusa
6. Frontals fused	no	no	no	no	no	yes	no
7. Dorsal portion of postorbital	large	small	small	small	small	small	small
8. Temporal arch	very exten- sive; formed by parietal, squamosal and supra- occipital	moderate; formed by squamosal and parie- tal	moderate; formed by squamosal and parie- tal	moderate; formed by squamosal and parie- tal	moderate; formed by squamosal and parie- tal	absent	very slender; formed by squamosal and supra- occipital
9. Skull flattened	no	no	no	no	extreme	moderate	moderate
10. Dorsal (hor- izontal) por- tion of parietal	•	covers cen- tral area of adductor fossa but not lateral area	covers cen- tral area of adductor fossa but not lateral area	covers little of adductor fossa but still present (although greatly re- duced in some)	of adductor fossa		absent; covers none of ad- ductor fossa
11. Lateral edges of parietals	sub- parallel but laterally ex- tensive	tapering anteriorly	parallel	wasp- waisted	wasp- waisted	tapering posterior- ly, greatly reduced	tapering posteriorly, greatly re- duced
12. Supra- occipital- parietal contact	present	absent	absent	absent	absent	absent (temporal bar absent)	present
13. Quadrate- parietal contact	present	absent	absent	absent	absent	absent	absent
14. Dorsal hor- izontal por- tion of supraocci- pital	broadly expanded	not ex- panded	slightly ex- panded	not ex- panded	not ex- panded	not ex- panded	slightly ex- panded
15. Crista supra occipitalis	- does not ex- tend be- yond fora- men magnum	extends be- yond for- amen mag- n num and condylus occipitalis	other chelid does not ex	amen mag-	tend be-	does not ex- tend be- yond fora- men mag- num	does not ex- tend beyond foramen mag- num

TABLE 2 – (Continued)

			TABLE 2 -	(Continued)			
	Pseudemydura	Emydura & Elseya	Platemys	Phrynops	Chelus	Chelodina	Hydromedusa
16. Medial portions of jugal and postorbital facing more laterally than posteriorly	no	no	no	no	yes, entire- ly on exter- nal surface of skull	yes	yes
17. Dorsal processes of exoccipitals meet above foramen magnum	no	no	yes	yes	yes	yes	yes
18. Cavum tympani extended laterally	no	no	no	no	yes	no	no
19. Maxilla reduced in exposure on triturating surface so that palatine bears lingual ridge	no	no	no	no	yes	no	no
20. Medial max- illary con- tact divid- ing premax- illae longi- tudinally	yes	no	no	no	no	no	no
21. Vomer- palatine contact	yes	yes	yes	yes	no, due to anterior pterygoid processes reaching vomer	yes	no, due to en- larged apertura narium interna reducing palatine
22. Large bony apertura narium interna formed by reduction of palatines		no	no	no	no	no	yes

TABLE 2 - (Continued)

	Pseudemydura	Emydura & Elseya	Platemys	Phrynops	Chelus	Chelodina	Hydromedusa
23. Quadrate- basisphe- noid contact	absent	absent	absent	absent	absent	extensive	limited
24. Symphy- seal suture separates lower jaw rami	no	no	yes	yes	yes	yes	yes
25. Relatively massive mandibles with symphy seal "hooks"	absent	present	absent	absent	absent	absent	absent
26. Prearticular separates or nearly sepa- rates coro- noid splenial	yes	no	no	no	no	no	no

#### GROUP 5-SUBTRIBE CHELINA

Boulenger (1889, p. 207) divided the "Chelydidae" into two sections: "I. Neck longer than the dorsal vertebral column..." and "II. Neck shorter than the dorsal vertebral column..." and this distinction, as emphasized by Goode (1967) and Burbidge, Kirsch, and Main (1974) still has usefulness for systematists. The extremely long cervical vertebrae of Chelus, Chelodina, and Hydromedusa may be hypothesized as a shared derived character testing the monophyletic nature of this group. In all other chelids the cervicals are shorter than the length of the vertebrae attached to the carapace and, as this latter condition is found in nearly all other turtles (as far as I know), it is presumably primitive.

Another feature found only in *Chelus*, *Chelodina*, and *Hydromedusa* involves the jugal and postorbital bones (figs. 1, 2, 3). In pleurodires

<sup>1</sup>Rhodin and Mittermeier (1976) described the jugal and postorbital areas in *Chelodina* and developed a useful terminology for it. Their figures 4 and 6 should be consulted as an aid to identification.

the jugal along with the postorbital make up the characteristic postorbital wall found in this group (Gaffney, 1975). The jugal usually forms the more lateral portion of the wall, whereas the postorbital forms the more medial area. As in cryptodires the jugal of pleurodires may reach the palatine but there is always a strong contact with the anterior edge of the pterygoid, presumably to aid in support of the uniquely pleurodiran processus trochlearis pterygoidei. In the primitive condition both bones consist of two portions, a medial section exposed behind the fossa orbitalis and forming the front of the fossa temporalis inferior, and a lateral portion exposed on the external surface of the skull. This situation occurs in pelomedusids, which is the basis for considering it primitive. It also occurs in Emydura, Pseudemydura, Platemys, and Phrynops. In Chelus, Chelodina, and Hydromedusa, however, the medial portion, which is covered by musculature (except in Chelus, see below), is turned outward to face somewhat more laterally rather than more posteriorly as in other chelids (Chelodina novaeguineae, fig. 8, is less like the other forms of Chelodina and more like

TABLE 3
Postcranial Features of Chelid Genera

	Pseudemydura	Emydura	Elseya	Platemys	Phrynops	Chelus	Chelodina Hydro	Hydromedusa
<ol> <li>Cervical vertebrae longer or shorter than dorsal vertebrae</li> </ol>	shorter	shorter	shorter	shorter	shorter	longer	longer	longer
2. Cervical (nuchal) scute	small and marginal	small and marginal	usually absent	small and marginal	small and marginal	small and marginal	small and marginal	large and separated from shell margin
3. Neural bones	absent	absent	absent	absent	variable, four to six	seven	variable, zero to eight	seven
4. Intergular scute entirely separating gular scutes	yes	yes	yes	yes	yes	no, gulars meet pos- teriorly <sup>A</sup>	no, gulars meet an- teriorly	yes
5. Intergular scute entirely separating humeral scutes and anterior portion of pectoral scutes	yes	no	О	ou	ou Ou	OI OI	yes	no
6. Claws on forefoot	five	five	five	five	five	five	four	four
7. First vertebral scute wider or narrower than second vertebral scute	narrower	narrower	патгоwег	wider	wider	wider	wider	wider

 $^d$ Except in fossil Chelus reported by Wood (1976).

Emydura, but I interpret this as a secondary condition). Chelus, on the other hand, is extreme in its degree of rotation of the medial portions of the jugal and postorbital. The medial areas in Chelus lie entirely on the surface of the skull, are not covered by musculature, and have no demarcation between medial and external surfaces. This condition of Chelus I interpret as uniquely derived or autapomorphic, as it does not occur in any other turtle.

It is interesting to note that Wood (1976) has recently described a Miocene Chelus, C. colombianus, which differs from C. fimbriatus in having an intergular enclosed by gulars (one individual apparently has an extra set of gulars) much as in Chelodina. This suggests the hypothesis that an enclosed intergular may be primitive for Chelodina, Hydromedusa, and Chelus. In any case, either the enclosed intergular evolved twice, or the open intergular evolved twice.

#### GROUP 6-INFRATRIBE HYDROMEDUSAD

The genera Chelodina and Hydromedusa are very similar in skull morphology. They both have relatively long, thin, and flat skulls, although Chelodina novaeguineae (fig. 8) is deeper than other Chelodina. Both taxa have extremely reduced temporal roof coverings and a markedly narrow parietal area between the temporal fossae (figs. 2, 3). The interorbital distance is narrower than in other chelids and the orbits face dorsally to a greater extent than a flattened form such as Chelus. The posterolateral process of the parietal, seen in other chelids (except Pseudemydura, apparently in coincidence with other unique features of the temporal roof) are absent in Hydromedusa and Chelodina. Chelodina is the only chelid to entirely lack a temporal bar of some sort, whereas in Hydromedusa an anteromedial process of the squamosal reaches the parietal and the latter bone has no lateral component in this region.

Chelodina and Hydromedusa also usually have a quadrate-basisphenoid contact, usually ventral to the prootic, but always leaving some of that bone exposed (figs. 4, 5). The contact is well developed in Chelodina but limited in Hydromedusa, barely taking place in some instances. My sample of Hydromedusa (three skulls) is too limited to determine variation of this feature, but I would not be surprised to see the contact absent in some specimens. Nonetheless, there are a number of unique features in common between Hydromedusa and Chelodina and I hypothesize that they are a strictly monophyletic group with respect to other chelids.

As noted by Boulenger (1889) Chelodina and Hydromedusa have four claws on their forefeet rather than five as in all other chelids and pelomedusids. I regard this as a derived feature also.

# **CLASSIFICATION**

In my opinion, a classification should be a redundant reflection of a phylogenetic hypothesis. Further discussion of this point of view may be found in Gaffney (1975, In press) and McKenna (1975). Although stability is often considered an important quality of classifications, I believe that it is often a spurious and misleading indication of the attainment of phylogenetic "truth." All of our notions about phylogeny are hypotheses that could be wrong; they can never be proved correct. If a classification is to have wide-ranging biologic usefulness, it must be susceptible to change. The classification presented here is as unstable as the phylogenetic hypothesis presented here.

#### CLASSIFICATION OF THE CHELID TURTLES

Infraorder Pleurodira (Cope, 1868b) Family Pelomedusidae Cope, 1868a Family Chelidae<sup>1</sup> Gray, 1825 Subfamily Pseudemydurinae, new Pseudemydura Subfamily Chelinae Gray, 1825, new rank Infrafamily Emydurodd,<sup>2</sup> new Emydura Elseya Infrafamily Chelodd Gray, 1825, new rank Tribe Platemini, new Platemys Tribe Chelini Gray, 1825, new rank Subtribe Hydraspina<sup>3</sup> Bonaparte, 1838, new rank Phrynops Subtribe Chelina Gray, 1825, new rank Infratribe Chelad Gray, 1825, new rank Infratribe Hydromedusad, new Chelodina Hvdromedusa

<sup>1</sup>The family name of this group of turtles has been spelled in a number of ways but most commonly Chelyidae. However, I am here following Williams (1950) and Wermuth and Mertens (1961), among others, in the use of Chelidae. As far as I can see, the oldest valid name for this family is Chelidina Gray, 1825, which becomes Chelidae with the addition of the appropriate ending. There could be some question as to whether or not Chelides Cuvier, 1817, might not be valid, but I am taking the view that it was not explicitly stated as a family level taxon.

The spelling variations are primarily due to the fact that although *Chelus* Dumeril, 1806, was the first spelling of this genus, it was later "corrected" to *Chelys* and many family level taxa were based on the latter spelling, which was popular during the 19th century. Kuhn

#### LITERATURE CITED

Albrecht, P. W.

1976. The cranial arteries of turtles and their evolutionary significance. Jour. Morph., vol. 149, no. 2, pp. 159-182.

Bell, T.

1828. On *Hydraspis*, a new genus of freshwater tortoises, of the family Emydida. Zool. Jour., vol. 3, pp. 511-513.

Blackmore, E. H.

1969. On the Australian Chelidae (Chelonia). Victorian Nat., vol. 86, no. 10, pp. 280-283.

(1967) gave a number of these spellings and a very useful guide to the higher categories.

<sup>2</sup>Elsewhere (Gaffney, 1972a) I use the arbitrarily chosen ending -odd for infrafamily and continue the practice here.

<sup>3</sup>Bonaparte (1838) originally named a family level taxon, Hydraspidina, based on the genus *Hydraspis*, a name now considered a synonym of *Chelodina* (see text and Stejneger, 1909). However, when Bonaparte used *Hydraspis* it referred to what is now *Phrynops* and, as I understand the rules on family level taxa, the name Hydraspidina must go with *Phrynops*.

<sup>4</sup>As there are no rules or suggestions dealing with tribal level endings, I arbitrarily choose -ad as the ending for infratribe.

Bonaparte, C. E.

1838. Amphibiorum Tabula Analytica. Nuovi Annali delle Scienze Naturali, vol. 1, pp. 391-393.

Bonde, Niels

1974. [Review of] Interrelationships of fishes. Greenwood, P. H., R. S. Miles and C. Patterson (eds.), Syst. Zool., vol. 23, pp. 562-569.

Boulenger, G. A.

1888. On the Chelyoid chelonians of New Guinea. Annali de Museo Civico de Storia Naturale Giacomo Doria, second ser., vol. 6, pp. 449-452.

1889. Catalogue of the Chelonians, Rhynchocephalians and Crocodiles in the British Museum (Nat. Hist.). New Edition, London, Printed by Order of the Trustees, 311 pp.

#### Bour, Roger

1973. Contribution a la connaissance de *Phrynops nasutus* (Schweigger: 1812) et *Phrynops tuberculatus* (Luederwaldt: 1926). Description d'une nouvelle sous-espece originaire du Paraguay, *Phrynops tuberculatus vanderhaegei* (Testudinata-Pleurodira-Chelidae). Bull. Soc. Zool. France, tome 98, no. 1, pp. 175-190, pl. 1.

# Bramble, Dennis Marley

[MS.] Functional morphology, evolution, and paleoecology of gopher tortoises. Ph.D. Diss., 1971, Univ. Calif., Berkeley, 341 pp.

#### Brundin, Lars

1968. Application of phylogenetic principles in systematics and evolutionary theory. In Φrvig, T. (ed.), Current problems of lower vertebrate phylogeny. Nobel Symposium 4, New York, Interscience Publishers, John Wiley and Sons, pp. 473-495.

Burbidge, Andrew A., John A. W. Kirsch, and A. R. Main

1974. Relationships within the Chelidae (Testudines: Pleurodira) of Australia and New Guinea. Copeia, no. 2, pp. 392-409.

#### Cogger, H.

1975. The reptiles and amphibians of Australia, A. H. and A. W. Reed, Sydney, pp. 1-584.

#### Cope, Edward D.

1868a. An examination of the Reptilia and Batrachia obtained by the Orton Expedition to Equador and the Upper Amazon, with notes on other species. Proc. Acad. Nat. Sci. Philadelphia, vol. 20, pp. 96-140.

1868b. On the origin of genera. *Ibid.*, vol. 20, pp. 242-300.

## Cracraft, Joel

1972. The relationships of the higher taxa of birds: problems in phylogenetic reasoning. Condor, vol. 74, no. 4, pp. 379-392.

1974. Phylogenetic models and classification. Syst. Zool., vol. 23, no. 1, pp. 71-90.

#### Cuvier, Georges

1817. Le regne animal distribue d'apres son organisation. tome 2, 1st ed., Paris, Deterville, Libraire, pp. xviii-532.

# Dumeril, A. M. Constant

1806. Zoologie analytique, ou methode naturelle de classification des animaux. Paris, Allais, Libraire, 1806, pp. xxxii-336.

#### Dunn, E. R.

1945. Los Generos de Anfibios y Reptiles de Colombia, IV. Cuarta y Ultima Parte: Reptiles, Ordens Testudineos y Crocodilinos, Caldasia, vol. III, no. 13, pp. 239-269.

#### Eldredge, Niles, and Ian Tattersall

1975. Evolutionary models, phylogenetic reconstruction, and another look at hominid phylogeny. *In* Szalay, F. (ed.), Approaches to Primate Paleobiology, Contrib. Primatol., vol. 5, pp. 218-242.

#### Farris, James S.

1972. Estimating phylogenetic trees from distance matrices. American Nat., vol. 106, no. 951, pp. 645-668.

#### Fitzinger, L.

1826. Neue Classification der Reptilien. Wien, J. G. Heubner Verlag, 66 pp.

# Freiberg, Marcos A.

1945. Una nueva especie de tortuga del genero "Platemys" Wagler. Physis, Rev. Soc. Argentina Cien. Nat., tomo 20, pp. 19-23.

1947. El alotipo de la tortuga "Platemys pallidipectoris" Freiberg. Ibid., tomo 20, pp. 112-115.

# Froes, Oscar Miranda

1957. Notas quelonologicas. I - Atualizacao da nomenclatura dos quelonios brasileiros. Iheringia Zool., no. 2, pp. 1-24.

# Fry, D. B.

1915. Herpetological Notes, Proceedings of the Royal Society of Queensland, vol. 27, pp. 61-95.

# Fuchs, Hugo

1931. Über den Unterkiefer und die Unterkiefernerven (Ramus tertius nervi trigemini et Chorda tympani) der Arrauschildkröte (*Podocnemis expansa*). Nebst Bemerkungen zur Kiefergelenksfrage. Zeitschr. Anat. Entwicklungsgeschichte, vol. 94, pp. 206-274.

#### Gaffney, Eugene S.

1972a. The systematics of the North American family Baenidae (Reptilia, Cryptodira).

- Bull. Amer. Mus. Nat. Hist., vol. 147, pp. 241-320.
- 1972b. An illustrated glossary of turtle skull nomenclature. Amer. Mus. Novitates, no. 2486, pp. 1-33.
- 1975. A phylogeny and classification of the higher categories of turtles. Bull. Amer. Mus. Nat. Hist., vol. 155, pp. 387-436.
- [In press.] A logical introduction to phylogenetic reconstruction. Carnegie Museum Publication.

#### Glauert, L.

- 1922. Contributions to the fauna of Western Australia. A new freshwater tortoise from the Murchison River. Jour. and Proc. of the Royal Society of Western Australia, vol. IX, part I, pp. 53-56.
- 1954. New swamp tortoise from the Swan River district, *Emydura inspectata* sp. nov. Western Australia Nat., vol. 4, no. 6, pp. 125-127.

#### Goode, John

- 1967. Freshwater tortoises of Australia and New Guinea (in the family Chelidae). Melbourne, Lansdowne Press, x plus 154 pp.
- 1968. Morphological variations in Victorian tortoises with special reference to the intermediate form of *Chelodina expansa*. Victorian Nat., vol. 85, pp. 263-267.

## Gray, John Edward

- 1825. A synopsis of the genera of reptiles and Amphibia, with a description of some new species. Ann. Philos., vol. 10, pp. 193-217.
- 1855. Catalogue of shield reptiles in the collection of the British Museum. Part I. Testudinata (tortoises). London, British Mus., pp. 1-82.
- 1863a. On the species of *Chelymys* from Australia, with the description of a new species. Ann. and Mag. Nat. Hist., ser. 3, vol. 12, pp. 98-100.
- 1863b. Additional observations on *Chelymys* dentata. Ibid., ser. 3, vol. 12, p. 246.
- 1864. On the genera of Chelydidae and the characters furnished by the study of their skulls. Proc. Zool. Soc. London, pp. 128-135.
- 1870. Supplement to the catalogue of shield reptiles in the collection of the British Museum. Part I. Testudinata (tortoises). London, British Mus., pp. ix-120.
- 1872. On the genus *Chelymys* and its allies from Australia. Proc. Zool. Soc. London, pp. 504-514, pls. 27-29.

1873. Additional notes on Spatulemys lasalae. Ann. and Mag. Nat. Hist., vol. 11, pp. 73-75, pl. 2.

#### Gregory, William King

1946. Pareiasaurs versus placodonts as near ancestors to the turtles. Bull. Amer. Mus. Nat. Hist., vol. 86, pp. 275-326, pls. 34-35.

#### Hay, O. P.

1908. The fossil turtles of North America. Carnegie Inst. Washington Publ., no. 75, pp. 1-568.

#### Hennig, Willi

- 1965. Phylogenetic systematics. Ann. Rev. Entomol., vol. 10, pp. 97-116.
- 1966. Phylogenetic systematics. Urbana, Univ. Ill. Press, 263 pp.

#### Hoffmann, C. K.

1890. Reptilien. I. Schildkröten. In Klassen und Ordnungen des Thier-Reichs, H. G. Bronn (ed.), Winter'sche Verlagshandlung, Leipzig, vol. 6, no. 3, pp. 1-442.

# Kanberg, Hans

1926. Mesoclemmys gibba Schweigger, eine seltene brasilianische Schlangenhalsschildkröte. Mitteil. Zool. Mus. Berlin, vol. 12, pp. 336-338.

# Krefft, G.

1876. Notes on Australian animals in New Guinea with description of a new species of fresh water tortoise belonging to the genus *Euchelymys* (Gray). Annali del Museo Civico de Storia Naturale de Genova, vol. 8, pp. 390-394.

#### Kuhn, O.

- 1964. Fossilium Catalogus. I: Animalia, Pars 107, Testudines. The Hague, Junk, 299 pp.
- 1967. Amphibien und Reptilien. Katalog der Subfamilien und höheren Taxa mit Nachweis des ersten Auftretens. Stuttgart, Gustav Fischer Verlag, pp. vii-124.

#### Loveridge, A.

1934. Australian reptiles in the Museum of Comparative Zoology, Cambridge, Massachusetts. Bull. Mus. Comp. Zool. Harvard, vol. 77, no. 6, pp. 243-383.

# Luederwaldt, Hermann

1926. Os Chelonios brasileiros com a lista das especies do Museu Paulista. Rev. Mus. Paulista, vol. 14, pp. 403-470.

# McKenna, Malcolm C.

1975. Toward a phylogenetic classification of the Mammalia. *In* Phylogeny of the Primates, Luckett, W. P., and F. S.

Szalay (eds.), New York and London, Plenum Press, pp. 21-46.

#### Medem, Fred

- 1960a. Datos zoo-geograficos y ecologicos sobre los Crocodylia y Testudinata de los rios Amazonas, Putumayo y Caqueta. Caldasia, vol. 8, pp. 341-351.
- 1960b. Informe sobre reptiles colombianos (IV). El primer hallazgo de la tortuga *Phrynops* (*Batrachemys*) nasuta (Schweigger) en Colombia. Novedades Colombianas, vol. I, no. 5, pp. 284-290.
- 1960c. Informe sobre reptiles colombianos (V). Observaciones sobre la distribucion geografica y ecologia de la tortuga *Phrynops geoffroana* ssp. en Colombia. *Ibid.*, vol. 1, no. 5, pp. 291-300.
- 1966. Contribuciones al conocimiento sobre la ecologia y distribucion geografica de *Phrynops (Batrachemys) dahli*; (Testudinata, Pleurodira, Chelidae). Caldasia, vol. 9, no. 45, pp. 467-489.
- 1973. El primer hallazgo de la tortuga Phrynops rufipes (Spix), en Colombia. Rev. Acad. Colombiana, vol. 14, no. 54, pp. 49-66.

## Mertens, Robert

- 1967. Bemerkenswerte Süsswasserschildkröten aus Brasilien. Senck. Biol., vol. 48, no. 1, pp. 71-82.
- 1969. Eine neue Halswender-Schildkröte aus Peru. *Ibid.*, vol. 50, p. 132.
- 1970. Zur Kenntnis von Phrynops nasutus. Ibid., vol. 51, pp. 17-20.

#### Mlvnarski, Marian

1976. Testudines. In Handbuch der Palaoherpetologie, Oskar Kuhn, (ed.), Stuttgart and New York, Gustav Fischer Verlag, pp. 1-130.

#### Müller, Lorenz

1939. Über die Verbreitung der Chelonier auf dem südamerikanischen Kontinent. Physis, Secc. Zool. Antropol., vol. 16, pp. 89-102.

#### Ogilby, J. D.

- 1890. Description of a new Australian tortoise. Rec. Australian Mus., vol. 1, pp. 56-59.
- 1905. Catalogue of the emydosaurian and testudinian reptiles of New Guinea. Proc. Roy. Soc. Queensland, vol. 19, pp. 1-32.

#### Ouwens, P. A.

1914. List of Dutch East Indian Chelonians in the Buitenzorg Zoological Museum. Contrib. a la Faune des Indes Neerlandaises, vol. 1, no. 1, pp. 29-32.

#### Peters, Wilhelm

1839. Zur Osteologie der *Hydromedusa maximiliani*. Arch. Anat., Physiol. und Wissenschaft. Medicine, pp. 280-295.

#### Peters, W., and G. Doria

1878. Catalogo dei Rettili e dei Batraci Raccoliti da O. Beccari, L. M. Albertis E-A. A. Bruijn Nella Sotto-Regione Austro-Malese. Annali del Museo Civico de Storia Naturale de Genova, vol. 13, pp. 323-327.

#### Popper, Karl

- 1968. The logic of scientific discovery. 2nd ed., New York and Evanston, Harper Torchbooks, Harper and Row Publishers, pp. 1-480.
- Rhodin, Anders G. J., and Russell A. Mittermeier 1976. Chelodina parkeri, a new species of chelid turtle from New Guinea, with a discussion of Chelodina siebenrocki Werner, 1901. Bull. Mus. Comp. Zool., Harvard Univ., vol. 147, no. 11, pp. 465-488.

#### Rooij, Nelly de

1915. The reptiles of the Indo-Australian Archipelago. Leiden, E. J. Brill Ltd., vol. 1, pp. xiv-384.

#### Schnee, P.

1899. Vorläufige Mitteilungen über die von mir in Australien zusammengebrachten Kriechtiere und Lurchen Frankfurt Zool, Garten., vol. 40, pp. 381-386.

#### Siebenrock, F.

- 1897. Das Kopfskelett der Schildkröten. Sitzungsberichte der Math.-Nat. Kl. K. Akad. Wissensch., vol. 106, pp. 245-328.
- 1901. Beschreibung einer neuen Schildkrötengattung aus der Familie Chelydidae von Australien: *Pseudemydura*, Anz. K. Akad. Wissensch., Math.-Nat. Kl., Wien, vol. 38, pp. 248-250.
- 1904. Schildkröten von Brasilien. Denkschr. Akad. Wissensch., Math.-Nat. Kl., Wien, vol. 76, pp. 1-28.
- 1905. Chelonologische Notizen. Zoolog. Anz., vol. 28, pp. 460-468.
- 1906. Bemerkungen zu zwei seltenen Schildkröten, *Ibid.*, vol. 30, pp. 127-129.
- 1907. Beschreibung und Abbildung von Pseudemydura umbrina Siebenrock und über ihre systematische Stellung in der Familie Chelydidae. Sitzber. Math.-Nat. Kl. Akad. Wissensch., vol. 116, pp. 1205-1212.
- 1909. Synopsis der rezenten Schildkröten, mit Berücksichtigung der in historischer

- Zeit ausgestorbenen Arten. Zool. Jahrb., suppl. 10, pp. 427-618.
- 1912. Über den Dimorphismus bei *Emydura* novae guineae Meyer. Zool. Anz., Band XL, pp. 301-304.
- 1914. Eine neue *Chelodina*-Art. aus Westaustralien. Anz. K. Akad. Wissensch. Math.-Nat. Kl., vol. 51, pp. 386-387.

Stejneger, Leonhard

1909. Generic names of some chelyid turtles. Proc. Biol. Soc. Washington, vol. 22, pp. 125-128.

Strauch, Alexander

1890. Bemerkungen über die Schildkrötensammlung im zoologischen Museum der Kaiserlichen Akademie der Wissenschaften zu St. Petersburg. Mém. Acad. Impér. Sci. St. Pétersburg, Ser. 7, vol. 38, no. 2, pp. 1-128.

Vestjens, W. J. M.

1969. Nesting, egg-laying and hatching of the snake-necked tortoise at Canberra, A.C.T. Australian Zool., vol. 15 no. 2, pp. 141-149, pls. 4-5.

Vogt, T.

1911. Reptilien und Amphibien aus Neu-Guinea. Sitzungsbericht Gesell. Naturforsch. Freunde zu Berlin, no. 9, pp. 410-432.

Wagler, J.

1830. Natürliches System der Amphibien, mit vorangehender Classification der Säugethiere und Vögel-ein Beitrag zur vergleichenden Zoologie. München, Stuttgart und Tübingen, J. G. Cotta'schen Buchhandlung, pp. vi-354.

Waite, E. R.

1929. The Reptiles and Amphibians of South Australia. Harrison Weir, Government Printer, North Terrace, Adelaide, 267 pp.

Wermuth, H., and R. Mertens

1961. Schildkröten, Krokodile, Brückenechsen. Jena, Gustav Fischer Verlag, pp. xxvi-422.

Werner, F.

1901. Ueber Reptilien und Batrachier aus Ecuador und Neu-Guinea. Verhandlungen der kaiserlich-königlichen zoologisch-botanischen Gesellschaft in Wien, vol. 51, pp. 593-614.

1909. Reptilia exkl. Geckonidae und Scinscidae. In Die Fauna Südwest-Australiens, Michaelsen, W. and R. Hartmeyer (eds.), Jena, Gustav Fischer Verlag, vol. 2, no. 16, pp. 251-278.

Wiley, E. O.

1975. Karl Popper, systematics, and classification: A reply to Walter Bock and other evolutionary taxonomists. Syst. Zool., vol. 24, no. 2, pp. 233-243.

Williams, Ernest E.

1950. Variation and selection in the cervical central articulations of living turtles. Bull. Amer. Mus. Nat. Hist., vol. 94, pp. 505-562.

1958. Rediscovery of the Australian chelid genus *Pseudemydura* Siebenrock (Chelidae, Testudines). Breviora, Mus. Comp. Zool., no. 84, pp. 1-8, 4 pls.

Wood, Roger Conant

1976. Two new species of *Chelus* (Testudine: Pleurodira) from the Late Tertiary of northern South America. Breviora, Mus. Comp. Zool., no. 435, pp. 1-26.

Wood, Roger C., and Richard T. J. Moody

1976. Unique arrangement of carapace bones in the South American chelid turtle *Hydromedusa maximiliani* (Mikan). Zool. Jour. Linnean Soc., vol. 59, no. 1, pp. 69-78.

Worrell, E.

1963. Reptiles of Australia. Angus and Robertson, Sydney; London; Melbourne; Wellington, pp. 1-207.

Zangerl, Rainer, and Fred Medem

1958. A new species of chelid turtle, *Phrynops (Batrachemys) dahli*, from Colombia. Bull. Mus. Comp. Zool., vol. 119, no. 5, pp. 375-390, 2 pls.



