Novitates

PUBLISHED BY THE AMERICAN MUSEUM OF NATURAL HISTORY CENTRAL PARK WEST AT 79TH STREET, NEW YORK, N.Y. 10024 Number 2980, 27 pp., 24 figs., 2 tables

August 9, 1990

Notes on Bolivian Mammals 6. The Genus *Ctenomys* (Rodentia, Ctenomyidae) in the Highlands

JOSEPH A. COOK,¹ SYDNEY ANDERSON,² AND TERRY L. YATES³

ABSTRACT

Tuco tucos (genus *Ctenomys*) of the Bolivian highlands are reviewed. Synonymies, references, a key for identification, lists of all known Bolivian specimens and localities, and new data on mor-

phology and karyology are given for the four species (*C. frater, C. opimus, C. lewisi,* and *C. leucodon*) present. Diploid chromosomes for these species number 52, 26, 56, and 36, respectively.

RESUMEN

Se revisaron los tuco tucos (género *Ctenomys*) del altiplano boliviano. Se presentan sinonimos, referencias, listas de todos los especímenes conocidos, un clave por identificación, y nuevos datos sobre morfología y cariología. Se encontró que

existen cuatro especies (C. frater, C. opimus, C. lewisi, y C. leucodon). Sus numeros de cromosomas diploides son 52, 26, 56, and 36, respectivamente.

INTRODUCTION

Ctenomys is a speciose genus of subterranean rodents that is endemic to southern South America. As is true for many rodent taxa of South America, the distribution and biology in general are poorly known for Ctenomys. Few specimens are available for

many of the estimated 30 (Mares and Ojeda, 1982) to 44 (Reig, 1986) species. *Ctenomys* is of considerable biological interest, however, as it is highly convergent ecologically and morphologically with North American pocket gophers (Geomyidae), with Middle

- ¹ Graduate Student, Department of Biology, University of New Mexico.
- ² Curator, Department of Mammalogy, American Museum of Natural History.
- ³ Research Associate, Department of Mammalogy, American Museum of Natural History; Curator of Mammals, Museum of Southwestern Biology, University of New Mexico, Albuquerque, NM 87131.

Eastern and European mole-rats (Spalacidae), and with African mole-rats (Bathyergidae; Nevo, 1979). Karyotypic evolution has been extensive in those subterranean rodents and *Ctenomys* also exhibits extremely high chromosomal variation, with a diploid number ranging from 10 in *C. steinbachi* to 70 in *C. pearsoni* (Anderson et al., 1987). Comparative study of these taxa may lead to productive insights into the processes responsible for these similarities and for karyotypic evolution in general.

Six nominal species of Ctenomys were reported for Bolivia by Cabrera (1961) and the taxonomy of four lowland species was summarized by Anderson et al. (1987). The present paper lists all known literature references to actual specimens for four other species from highland Bolivia and all known specimens. Standard chromosomal and morphological variation within and among these highland species are described herein and the karvotypes of three of the four species are reported for the first time. Notes on C. conoveri, C. steinbachi, and "C. minutus" are added. Further study of the three Bolivian specimens previously and tentatively identified as C. minutus will establish their actual identity.

MATERIALS AND METHODS

TAXONOMIC SYNOPSIS: This synopsis is based on literature and on examination of specimens in a number of major museums (by Anderson). Material from the Bolivian highlands, reported here for the first time, adds to knowledge of distributions and the morphological limits of *Ctenomys*. Museums and their acronyms were published earlier (Anderson et al., 1987).

MORPHOLOGICAL ANALYSIS: Standard external measurements from fresh specimens and cranial measurements (as figured previously, Anderson et al., 1987) taken with calipers are as follows (all measurements are in millimeters, except weight is in grams):

H&B Length of head and body TAI Length of tail vertebrae

HF Length of hind foot, including longest claw

EAR Length of ear from notch

WT Weight in grams
CRL Condylobasilar length
CIL Condyloincisive length

PRL Palatilar length

MXL Alveolar length of maxillary tooth row

DSB Breadth of dental span

PRB Breadth of palate between closest alveolar

margins

ZYB Greatest breadth across zygomatic arches

IOB Breadth at interorbital constriction

LAB Greatest breadth across lambdoidal ridges SKD Depth of skull from plane of occlusal sur-

faces to top of skull.

Using the following series, ratio diagrams (Anderson et al., 1987) were drawn in order to (1) visualize ontogenetic changes in proportions by comparing individuals of different ages in samples of sufficient size and (2) compare patterns for adults of five species.

Ctenomys frater frater from Potosí, 2 specimens; C. f. mordosus from near Tambo in Tarija, 12; C. leucodon, 5; C. lewisi from near Iscayachi in Tarija, 12, range only plotted; C. lewisi from Sama (probably at or near the preceding locality) in Tarija, 5; C. opimus opimus from Carikari in Potosí, 4; from Uyuni in Potosí, 4; from near Laguna Colorado in Potosí, 6; from near Huancaroma in Oruro, 28; from near Río Barros in Oruro, 9; and from near Sajama in Oruro, 11.

Based on comparisons of these diagrams with each other and with similar diagrams prepared for our earlier study of lowland species (Anderson et al., 1987), various differences in size and proportions of both external and cranial dimensions were evaluated and noted in the taxonomic comparisons below.

Measurements of series of *Ctenomys opimus opimus* from several localities were analyzed to evaluate differences relating to sex and estimated age. Three of the species were compared by analysis of Canonical Discriminant Functions to assess the amount of variation due to locality.

Measurements of selected specimens are included in table 2.

SPECIMENS EXAMINED: Sample localities for karyotypes are indicated in figure 1, along with all localities for highland *Ctenomys* known from the literature or from specimens examined by us. All specimens are listed in the taxonomic synopsis; those examined karyotypically for this study are indicated by NK numbers; and catalog numbers for those referred to specifically in text, figures, or ta-

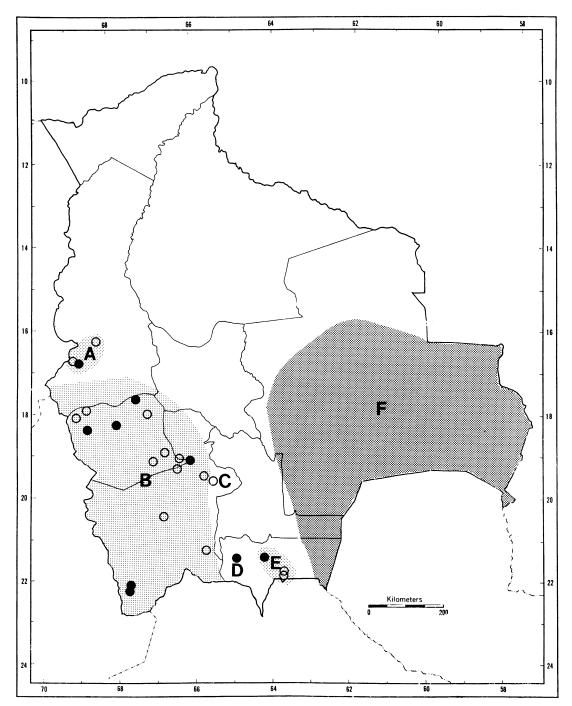


Fig. 1. Map of southwestern Bolivia showing all known localities of specimens of *Ctenomys* from highland Bolivia. Black dots are localities from which karyotypes have been examined. Taxa are: A. C. leucodon, B. C. opimus, C. C. frater frater, D. C. lewisi, and E. C. frater mordosus. F. Stippled area represents the distribution of lowland species of *Ctenomys* (Anderson et al., 1987).

bles also are given. Measurements in table 2 are of the largest specimen of a male and of a female for each species (in condylobasilar measurement).

KARYOTYPIC ANALYSIS: Nondifferentially stained karyotypes were prepared from 43 specimens representing four highland species of Bolivian Ctenomys. Specimens were collected using Macabee gopher traps or Victor steel traps. Traps were checked frequently to reduce mortality prior to karyotyping. Karyologic techniques were described in Anderson et al. (1987). Specimens of Ctenomys opimus were collected during expeditions in 1984 and 1986, while C. frater, C. lewisi, and C. leucodon were collected on the latter trip. Diploid number was verified by counting at least 10 cells per individual. Voucher specimens were deposited in the Mammal Division of the Museum of Southwestern Biology at the University of New Mexico, the Bolivian Museo Nacional de Historia Natural in La Paz, or in the Department of Mammalogy at the American Museum of Natural History.

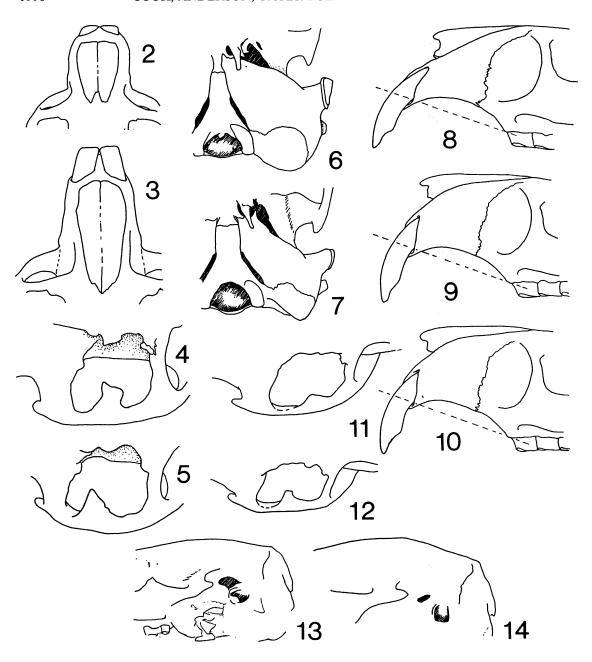
ACKNOWLEDGMENTS

Most of the specimens reported here were obtained on expeditions partly supported by grants from the National Science Foundation (BSR 83-16740 and BSR 84-08923), National Institutes of Health (DRR-RR08139-14), American Society of Mammalogists Grantin-Aid, Sigma Xi, and a Tinker Foundation Field Research Grant. We are grateful to Nancy Olds for measuring and photographing skulls in the laboratory and for participating in the fieldwork. Others who helped in the fieldwork included Jorge Salazar Bravo, Jackaline Miralles Goytia, Teresa Tarifa Suárez, Freddy Caceres Vega, Carmina Miranda Morales, Otto Carlos Jordán Camacho, Carl Schuster, and especially Scott L. Gardner (BSR 86-12329). We are grateful to the curators of the museums where we studied specimens, to Philippe Desjeux of the Instituto Boliviano de Biología de Altura for the many courtesies he extended, and to Gerhard Storch for verifying the identification of specimens in the Forschungsinstitut Senckenberg. We are grateful also to our reviewers, Forrest Davis, Jennifer Frey, Milton H. Gallardo, Scott L. Gardner, Enrique P. Lessa, and James L. Patton for their helpful comments. Nella Sánchez Cook assisted in the chromosomal analyses. The local people at the Estancia Huancaroma, the ENDE (Empresa Nacional de Electrificación) camp at Laguna Colorado, and at other places were helpful in many ways. William Franklin kindly did the statistical analyses for us.

KEY TO BOLIVIAN SPECIES OF CTENOMYS

inhabiting eastern Santa Cruz

- Pelage blackish; postorbital processes farther forward (fig. 4); inhabiting western Santa Cruz; 2N = 10 Ctenomys steinbachi
 Pelage brown; postorbital processes farther back (fig. 5); widely ranging in Santa Cruz; 2N variable, 36 to 46 ... Ctenomys boliviensis
- - Pelage varying from tan to blackish; bullae not expanded (fig. 7) 6
- - Pelage dark brown; width of an upper incisor less than 3.3; incisors less procumbent (fig.



Figures 2-14

TAXONOMIC SYNOPSIS

Genus Ctenomys Blainville, 1826

CONTEXT: Order Rodentia, superfamily Octodontoidea, family Ctenomyidae, in-

cludes five fossil genera but only one living genus, *Ctenomys* (Woods, 1984: 427).

CONTENT: Ctenomys includes about 33 recognized living species (Honacki et al., 1982) and some additional subspecies, and reaches its northern limits in Peru, Bolivia, and southern Brazil.

Data on three species of highland Bolivian Ctenomys (C. opimus, C. lewisi, and C. frater)

were analyzed separately for males and for females, with weights (in grams) and without weights, by Canonical Discriminant Function Analysis. In all four analyses, 100 percent were correctly classified as to group membership. This merely reinforces our earlier conclusion that every relatively complete specimen from the Bolivian highlands can be readily identified as to species. We have thus far discovered no complications such as we found earlier in lowland populations of *Ctenomys boliviensis* (Anderson et al., 1987).

Ctenomys frater Thomas, 1902 Ctenomys frater frater Thomas, 1902

Ctenomys frater Thomas, 1902: 228 (two from the type locality of Potosí, Bolivia). Neveu-Lemaire and Grandidier, 1911: 14 (Potosí).

Ctenomys frater frater: Cabrera, 1961: 548 (Potosí). Anderson, 1985: 14 (name only).

CHARACTERISTICS: Because species of Ctenomys tend to occupy mutually exclusive geographic ranges, the locality of origin of a specimen when compared with a map of distributions will usually provide an identification. However, many details of distribution remain to be discovered and all identifications should be checked on the basis of the visible morphological characters of the specimen, as outlined in the following notes.

COMPARISONS: In comparison with Ctenomys boliviensis (of both Bolivian subspecies), C. frater (of both Bolivian subspecies) having longer, softer, and darker, blackish, rather than brownish, pelage; throat collar indistinct; tail and hind foot both slightly shorter (relative to head and body length, HBL); skulls not noticeably different in dimensions used in the ratio diagrams (figs. 15 and 16). Based on direct comparison of skulls (of the subspecies C. f. mordosus, there being only three skulls of C. f. frater and none being available at the moment), C. frater smaller than C. boliviensis; incisors smaller and perhaps slightly less procumbent; anterior part of nasal seemingly less flattened and less shortened; zygomatic arch with lateral profile, in dorsal view, flattened in middle third rather than evenly, widely, and outwardly bowed; bullae much smaller, less inflated, less proiecting, and less exposed posterior to lambdoidal crest; supraorbital process of frontal small or absent rather than conspicuous, elevated, and continuing posteriorly onto the parietal as a distinct ridge; dorsal profile with concavity in frontal area rather than with concavity absent or slight.

In comparison with C. conoveri, C. frater having shorter, finer, and blackish, rather than brownish, pelage (figs. 17 and 18); tail, hind foot, and ears longer, relative to the HBL; and palatilar length (PRL) lesser, breadth of dental span (DSB) greater, breadth of palate between closest alveolar margins (PRB) greater, and depth of skull (SKD) lesser, all relative to the condylobasilar length (CRL). Based on direct comparison of skulls, C. frater smaller; upper incisors smoothly curved rather than with shallow longitudinal grooving (seen in no Bolivian Ctenomys except C. conoveri); anterior root of zygomatic arch projecting laterally rather than posterolaterally: supraorbital process small or absent: dorsal process of jugal relatively low and triangular rather than high and with a secondary point; postorbital breadth more than interorbital breadth rather than the reverse; lateral profile of arch in dorsal view slightly curved in medial third rather than sharply curved; mesopterygoid fossa an open V rather than more acuminate; bullae relatively less inflated, especially posteriorly.

In comparison with C. leucodon, C. frater (both subspecies) having darker, seemingly stiffer, less lax pelage; hind feet more slender; size slightly smaller; tail shorter, relative to HBL; and, relative to the CRL, condyloincisive length (CIL) shorter, tooth row (MXL) longer, and SKD shallower; and DSB/MXL less. Based on direct comparison of skulls, C. frater smaller; upper incisors less procumbent, anterior surface orange rather than pale orange or yellowish; anterior margin of zygomatic bridge in dorsal view sloping posterolaterally rather than nearly at right angle to longitudinal axis; dorsal process of jugal not so far posterior, orbit smaller; rostrum tapering but slightly anteriorly, rostral breadth near incisive alveoli broad, relative to size of skull; dorsal profile of skull less vaulted, slightly concave in frontal area, skull not so high; bullar tubes directed laterally rather than a bit more dorsolaterally (as a specialization for life in open places both eyes and ears may be placed high on the head as seen in altiplano

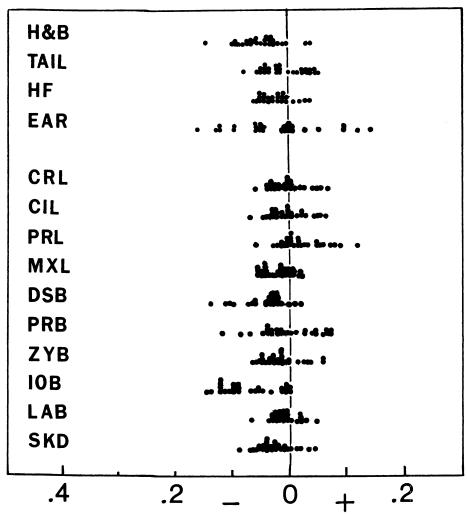


Fig. 15. Ratio diagram showing log differences for measurements of 28 specimens of *Ctenomys opimus* from near Huancaroma in Oruro, the largest series available for any highland tuco tucos in Bolivia. The standard is a specimen of *C. steinbachi* used in our earlier study of lowland *Ctenomys* (Anderson et al., 1987). No very juvenile animals are included in the series, but adults of various ages and of both sexes are included.

dwellers such as Microcavia niata and Galenomys garleppi).

In comparison with *C. lewisi*, *C. frater* is similar in darkness and color of dorsal pelage, venter less brightly ochraceous; smaller; hind foot smaller, relative to HBL; and (relative to the CRL) DSB, PRB, zygomatic breadth (ZYB), interorbital breadth (IOB), lambdoidal breadth (LAB), SKD, and MXL all greater. Based on direct comparison of skulls, *C. frater* smaller; upper incisors less procumbent, smaller; zygomatic bridge sloping posterolaterally (more than in *C. lewisi* but the

contrast less than between C. frater and C. leucodon); dorsal jugal process more posterior; bullae small relative to size of skull in both C. frater and C. lewisi.

In comparison with Bolivian C. minutus (which, as noted earlier, may eventually prove to be of some other species), C. frater having brownish pelage darker, especially middorsally where almost black; stiff hairs along lateral edge of manus whitish and contrasting with blackish dorsal pelage rather than reddish brown in both places; larger; ears larger relative to HBL; and, relative to CRL, PRL

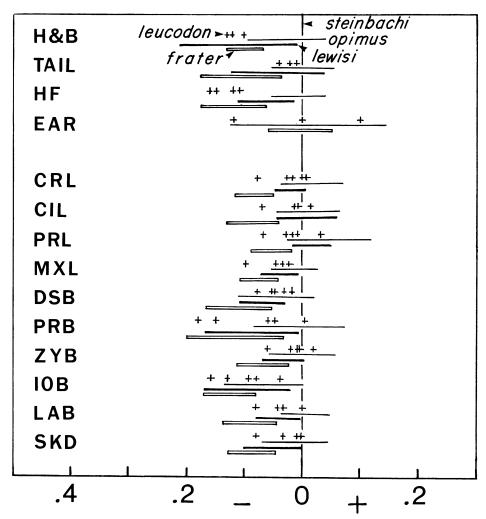


FIG. 16. Ratio diagram showing ranges of values for adults of four Bolivian highland species of *Ctenomys* compared to the same standard used in figure 15. The smallest individual of *Ctenomys opimus* in figure 15 has been omitted. Differences in proportions are discussed in text.

and PRB greater. Based on direct comparison of skulls, *C. frater* larger; rostrum relatively longer; bullae absolutely and relatively smaller.

In comparison with *C. opimus, C. frater* having dark brown dorsal pelage, relatively little contrast between the tips, and dark gray rather than tan bases of hairs, and with much contrast between pale tips and darker bases; pelage stiffer and less lax to the touch; size generally smaller; hind foot shorter, relative to HBL. Cranial proportions not very different. Based on direct comparison of skulls, *C. frater* smaller; orbit smaller, jugal process and

supraorbital process less conspicuous, smaller; auditory tube projecting laterally from a lower position on the bulla; bullae and some associated parts of skull smaller, volume estimated as one-fifth that of *C. opimus*, gap between anterior projections of bullae about 3 mm rather than 2 as in *C. opimus*; margin of mesopterygoid fossa a more open, less narrow, V-shape.

In comparison with *C. steinbachi*, *C. frater* having dark brownish rather than black glossy dorsal pelage; venter paler than dorsum but less markedly contrasting; less distinct pale throat collar; tail longer and ears larger, both

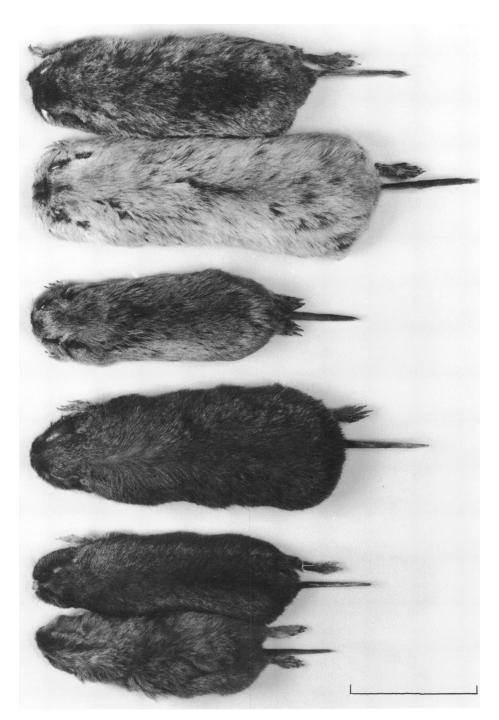


FIG. 17. Photographs of study skins in dorsal view of five taxa of *Ctenomys*. From top down, these are: *C. opimus opimus* (AMNH 263031 and 263035, the former with worn pelage), *C. leucodon* (AMNH 263028), *C. lewisi* (AMNH 263015), *C. frater mordosus* (AMNH 263014), and *C. frater frater* (BMNH 2.2.2.112). The scale represents 100 mm.



Fig. 18. Photographs of study skins in ventral view of five taxa of *Ctenomys*. Same specimens and scale shown in figure 17.

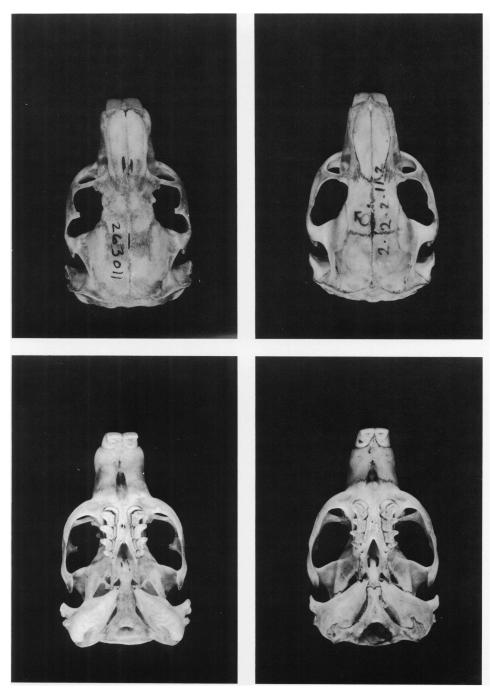


Fig. 19. Dorsal and ventral views of skulls of *Ctenomys frater mordosus* (left, AMNH 263011) and *C. frater frater* (right, BMNH 2.2.2.112). Same scale as figure 20.

relative to HBL; and, relative to CRL, greater PRL, greater PRB, and lesser IOB. Based on direct comparison of skulls, *C. frater* smaller; incisors less procumbent, smaller; nasals nar-

rower, less truncate anteriorly; rostrum not so broad; dorsal process of jugal more posterior, triangular rather than with secondary point; bullae less inflated (fig. 19). DISTRIBUTION: Ctenomys frater frater is known only from the department of Potosí (see fig. 1). Cabrera, 1961: 547, recognized five subspecies, three in Argentina and two in Bolivia.

SPECIMENS, 3: dept. of Potosí: 1935/6545, Potosí, 2 BM (2.2.2.112 and 113), 1 ANSP (19352).

REMARKS: Thomas, 1902, distinguished C. frater from C. opimus and assigned specimens from [at or near the city of] "Potosi" to both species. In the paper with the original description, Thomas quoted the collector as having caught the specimens of C. frater "in ground in moist sand" whereas the specimens of C. opimus were taken "in sandy places on ridge" and "in decomposed trachyte soil on ridge." The exact distributions and habitats occupied by the two species near the city need to be studied further. We hypothesize that their precise local distributions will not overlap or be ecologically sympatric and that C. opimus will be found at higher elevations and C. frater at lower elevations. The identification of one young individual (ANSP 19352) is tentative. The back of the skull with the most diagnostic features of the bullae is missing.

Ctenomys frater mordosus Thomas, 1926

Ctenomys sylvanus: Thomas, 1925: 580 (Carapari), not of Thomas, 1919.

Ctenomys sylvanus mordosus Thomas, 1926: 325 (type locality Tambo, department of Tarija, Bolivia; also cited Melocoton). Rusconi, 1928: 247 (Tambo).

Ctenomys frater mordosus: Cabrera, 1961: 548 (Tambo). Anderson, 1985: 14 (name only).

Ctenomys frater: Lessa and Cook, 1989: 857 (8 km W of Rancho Tambo).

CHARACTERISTICS: Thomas (1926) characterized C. s. mordosus as having "size and general external appearance quite as in true sylvanus" [from Salta in Argentina, now considered to be a subspecies of C. frater] but the incisors were said to be much broader and heavier, "their combined breadth attaining 7 mm. in old specimens as compared with 5.5 mm. in equally aged specimens of sylvanus." The rostrum of mordosus is thickened, in a manner seen "to a still greater extent in C. boliviensis" and in C. steinbachi. "An allied species, with equally broad inci-

sors, is the *Ctenomys frater* of Potosi, but that has peculiarly narrowed bullae and is of a decidedly lighter colour." This summarizes relevant information in the literature.

COMPARISONS: Ctenomys frater mordosus is darker than C. f. frater, known from Potosí in a generally higher and drier area to the west. Additional specimens of the latter are needed in order to evaluate possible cranial differences.

DISTRIBUTION: Ctenomys frater mordosus occurs in Tarija east of the ranges of Ctenomys frater frater and of C. lewisi, as shown in figure 1.

SPECIMENS, 26 (New Mexico karyotype, or NK, numbers and sexes are in parentheses): Tarija; 2127/6419, Rancho Tambo, 61 km by road E of Tarija, 2 AMNH (including NK 14610 F, 14613 and 263014 F); 2127/6423, Tambo, 8 BM, 1 FMNH; 2127/6424, 8 km W of Rancho Tambo, 2 AMNH (263011, 263012), 1 MNLP, 2 MSB (NK 14621 F, 14622 M, 14623 M); 2149/6346, Carapari, 2 BM (Thomas, 1925); 2202/6345, Yacuiba, 2 BM; coordinates uncertain, Melocoton, 6 BM (including 26.1.1.126).

Ctenomys leucodon Waterhouse, 1848

Ctenomys leucodon Waterhouse, 1848: 281 (type locality San Andres de Machaca, south of Lake Titicaca, department of La Paz, Bolivia). Thomas, 1927: 552 (lectoparatypes). Rusconi, 1928: 239 (La Paz); Anderson, 1985: 14 (name only).

CHARACTERISTICS: Extremely proodont, relatively narrow, and pale upper incisors are noteworthy. *Ctenomys leucodon* exhibited a high-pitched whistle vocalization on several occasions. Individuals of this species seemed to spend more time at burrow openings than did other Bolivian tuco tucos. See other comparisons below.

COMPARISONS: In comparison with *C. boliviensis*, *C. leucodon* having longer softer pelage; no paler throat collar; plumbeous bases of ventral hairs longer and darker; hind foot shorter and condylobasilar (CRL) greater, relative to the head and body length (HBL); and, relative to CRL, breadth of palate between closest alveolar margins (PRB) narrower. Based on direct comparison of skulls, *C. leucodon* having more procumbent, small-

er, and paler upper incisors; rostrum narrower and more attenuate anteriorly; zygomatic arch and bridge more slender; arch less evenly, widely, and outwardly bowed; supraorbital process present, but process and ridge posterior to it are less conspicuous; lateral tubes of bullae perhaps more dorsolaterally directed; bullae smaller, less inflated.

In comparison with C. conoveri, C. leucodon having finer pale yellowish brown rather than coarse reddish brown pelage; tail longer, hind foot longer, and CRL greater, all relative to HBL; and, relative to the CRL, palatilar length (PRL) lesser, breadth of dental span (DSB) greater, interorbital breadth (IOB) less, and depth of skull (SKD) less. Based on direct comparison of skulls, C. leucodon much smaller; skull generally less massive; incisors more procumbent, narrower, and paler; anterior root of zygomatic arch projecting laterally rather than posterolaterally; outer margin of arch nearly parallel in middle third rather than conspicuously curved there; supraorbital process less conspicuous: bullae less inflated.

In comparison with C. lewisi, C. leucodon having paler, more yellowish, pelage; tail longer and hind foot shorter relative to HBL and relative to CRL; DSB greater relative to the length of the upper tooth rows (MXL). Based on direct comparison of skulls, C. leucodon smaller; incisors more slender and paler, although procumbent in both species; rostrum attenuate anteriorly, narrower; dorsal profile convex rather than concave in frontal area; dorsal process of jugal and supraorbital process more posterior; bullar tubes directed a bit more dorsolaterally rather than laterally; bullae roughly comparable in size, perhaps slightly smaller in relation to size of skull and more attenuate anteriorly in lewisi.

In comparison with Bolivian "C. minutus," C. leucodon having longer, more lax, paler, and more yellowish pelage; size larger; tail longer, hind foot shorter, CRL greater, all relative to HBL; and, relative to CRL, MXL slightly less. Based on direct comparison of skulls, C. leucodon much larger; incisors more procumbent and paler; anterior base of zygomatic arch in relation to position of zygomatic bridge more anterior; orbit seems relatively larger, dorsal jugal process more posterior; bullae relatively less inflated.

In comparison with C. opimus, C. leucodon is similar in color and texture of pelage; smaller; tail longer and hind foot shorter, both relative to HBL; and cranial proportions similar. Based on direct comparison of skulls, C. leucodon smaller; skull less massive; rostrum more attenuate, narrower; incisors more proodont, narrower, paler; bullar tubes directed dorsolaterally rather than laterally, but from a lower position on the bulla so that the external ears of both species are probably relatively high on the head; eyes of both seem comparatively large, supraorbital and dorsal jugal processes relatively posterior in position in both species: bullae much less inflated: gap between anterior processes of bullae nearer 3 than 2 mm (fig. 20).

In comparison with C. steinbachi, C. leucodon having paler reddish brown rather than glossy blackish dorsal pelage, and ochraceous, rather than grizzled, venter; venter not sharply contrasting with sides; no paler collar marking; tail longer, hind feet shorter, and CRL greater, all relative to HBL; and, relative to CRL, MXL less, breadth of palate between closest alveolar margins (PRB) less, and IOB less. Based on direct comparison of skulls, C. leucodon smaller, skull less massive, narrower; incisors more proodont, narrower, and paler: rostrum narrower, attenuate; orbit larger, supraorbital and dorsal jugal processes relatively posterior; bullae smaller, less inflated; interorbital area much narrower.

DISTRIBUTION: Ctenomys leucodon occurs in a relatively small area to the southeast of Lake Titicaca. See figure 1.

SPECIMENS, 9: dept. of La Paz: 1613/6827, Chilalaya, 1 BM (1901.6.7.56); 1644/6901, San Andres de Machaca, 4 BM; 1648/6853, 14 km by road SW of San Andres de Machaca, 2 AMNH (including 263028), 1 MSB (NK 14789 M, 14791 F, 14793 F). Department unknown: coordinates unknown, Comauchi, 1 BM.

Ctenomys lewisi Thomas, 1926

Ctenomys lewisi Thomas, 1926: 323 (type locality Sama, 4000 m, department of Tarija, Bolivia). Rusconi, 1928: 247 (Sama). Anderson, 1985: 14 (name only). Lessa and Cook, 1989: 857 (1 km E of Iscayachi).

CHARACTERISTICS: Thomas (1926) characterized *C. lewisi* as a large reddish-brown

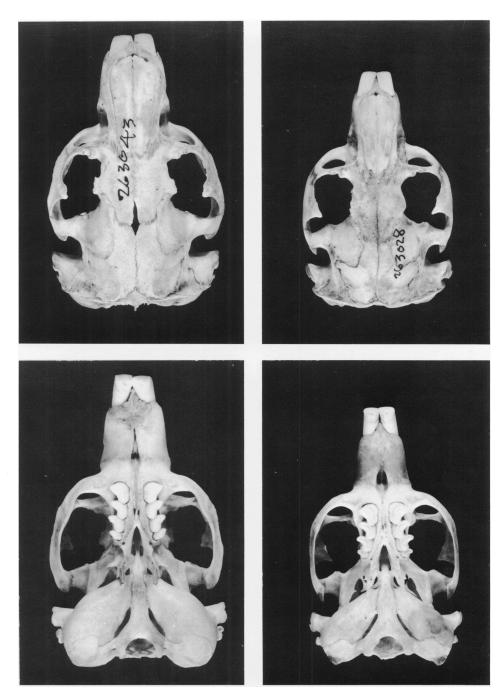


Fig. 20. Dorsal and ventral views of skulls of *Ctenomys opimus* (left, AMNH 263043) and *C. leucodon* (right, AMNH 263028). For scale, the zygomatic breadth of 263043 is 36.5 mm.

species with unusually proodont incisors (see figs. 17 and 18 for illustrations of skins). Ctenomys leucodon "has similarly proodont incisors, but these are comparatively slender,

are not so strongly flattened in front, and are almost unpigmented."

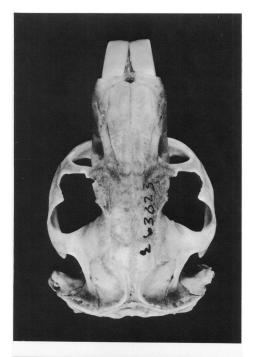
Some measurements of the holotype (a young adult male) and of an old female were:

head and body 219, 204; tail 68, 71; hind foot 37, 37; greatest length of skull 52, 50; condyloincisive length 54, 51.5; zygomatic breadth 33, 31; nasals 17×9.5 , 17×8.8 ; interorbital breadth 13.5, 12.3; mastoid breadth 29.5, 29.2; breadth across muzzle 14, 12.8; palatilar length 25.8, 25; and combined breadth of incisors 9, 8.

COMPARISONS: In comparison with C. boliviensis, C. lewisi having longer, finer, darker and less reddish dorsal pelage; collar indistinct; plumbeous bases more distinct on ventral pelage; size smaller; tail shorter, relative to head and body length (HBL); and, relative to condylobasilar length (CRL), breadth of dental span (DSB), breadth of palate between closest alveolar margins (PRB), zygomatic breadth (ZYB), and interorbital breadth (IOB) all narrower. Based on direct comparison of skulls, C. lewisi having incisors more procumbent and wider; dorsal profile with concavity in frontal area; supraorbital process and ridge extending posteriorly from it less conspicuous; zygomatic arch less massive, less widely bowed; bullae smaller, less inflated (fig. 21).

In comparison with C. conoveri, C. lewisi having finer, longer, darker dorsal pelage: greater contrast between ochraceous tips and dark gray bases in ventral pelage rather than hairs more or less unicolored and with inconspicuous grayish bases; smaller; tail and hind foot both longer relative to HBL; and, relative to CRL, DSB greater, and palatilar length (PRL), PRB, ZYB, IOB, and depth of skull (SKD) all less. Based on direct comparison of skulls, C. lewisi smaller; incisors more procumbent, wider, on the average slightly darker orange; nasals slightly narrower and their anterior margins slightly nearer alveolar margins of incisors; dorsal profile of skull concave in frontal area rather than convex throughout; ratio of interorbital breadth to postorbital breadth at constrictions seems greater; lateral profile of zygomatic arch in dorsal view with greatest curvature in posterior third of arch rather than in middle third; anterior root of arch projecting laterally rather than posterolaterally; supraorbital process small and rounded rather than longer and pointed; bullae smaller, less inflated, especially posteriorly.

In comparison with Bolivian "C. minu-



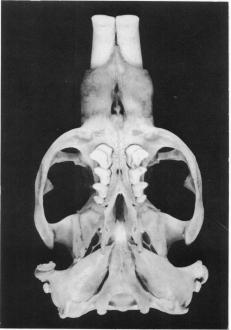


FIG. 21. Dorsal and ventral views of skull of *Ctenomys lewisii* (AMNH 263023). Same scale as figure 20.

tus," C. lewisi having darker and less reddish dorsal pelage and more ochraceous ventral pelage; larger in general; CRL greater relative to HBL; and, relative to CRL, having greater PRL, and lesser MXL, DSB, and PRB. Based on direct comparison of skulls, *C. lewisi* much larger; incisors more procumbent, wider, orange in both species; anterior roots of zygomatic arch are, relative to position of bridge, more anterior; bullae relatively less inflated; skull more massive.

In comparison with *C. opimus*, *C. lewisi* much darker dorsally, more ochraceous ventrally; slightly smaller, but cranial proportions not very different.

In comparison with C. steinbachi, C. lewisi having reddish brown rather than glossy black dorsal pelage; venter not sharply contrasting with sides, venter ochraceous rather than paler and grizzled; indistinct collar on throat; tail shorter (in the non-Iscavachi series), and, relative to the CRL, greater PRL and lesser DSB, PRB, IOB, and SKD. Based on direct comparison of skulls, C. lewisi having incisors more procumbent, darker orange, wider; orbits smaller; dorsal profile with concavity in frontal area; anterior margin of mesopterygoid fossa less pointed, fossa wider; bullar tubes lower on bullae; bullae smaller; gap between anterior processes of bullae nearer 3 than 2 mm.

DISTRIBUTION: Ctenomys lewisi is known only from Tarija, see figure 1.

SPECIMENS, 35: Tarija; 2129/6457, 1 km E of Iscayachi, Rio Tomayapo, 13 AMNH (including 263015 and 263023), 1 MNLP, 5 MSB (NK 14650 F, 14651 F, 14652 F, 14653 and 57186 F, 14670 F, 14709 F); 2129/6502, Sama, 13 BM (including 26.1.1.116, the holotype, and 26.1.1.122; Thomas, 1926), 1 KU, 2 FMNH.

REMARKS: Semiaquatic habits of *C. lewisi* were suggested by captures in holes in wet stream banks (Thomas, 1926). No such habit has been suggested for any other *Ctenomys*. Some, but not all, of the specimens captured in 1986 near Iscayachi were in damp soil near the river, but there was no indication of a semiaquatic existence.

Ctenomys opimus opimus Wagner, 1848

Ctenomys Brasiliensis: Waterhouse, 1848: 274, pl. 8, fig. 6 (Potosí), not of Blainville, 1826.

Ctenomys opimus Wagner, 1848: 75 (type locality Bolivia, restricted by Thomas, 1900, to Mount Sahama, department of Oruro). Thomas, 1902:

227 (Oruro, Challapata, Pampa Aullaga, Livichuco, and 5 from Potosí). Osgood, 1916: 210 (Oruro). Neveu-Lemaire and Grandidier, 1911: 14 (Sahama). Anderson, 1985: 14 (name only). Lessa and Cook, 1989: 857 (four localities in department of Oruro).

Ctenomys opimus opimus: Yepes, 1930: 325 (Potosí, Pampa Aullaga, Livicucho).

CHARACTERISTICS: Large tuco tucos; pale vellowish, tending to become darker as tips of hairs wear, sometimes in patches, as on the top of the head; pelage long, soft, lax. Ctenomys opimus inhabited burrow systems with Phyllotis xanthopygus and Galea musteloides at sites near Cruce Ventilla (observations by Cook); and likewise with a murid mouse and a guinea pig, Auliscomys boliviensis and Microcavia niata respectively, at a site 45 miles distant and on a compass bearing of 123° from Challapata (observations by Anderson). See specific comparisons below, especially with C. leucodon and C. frater, the two Bolivian species occurring nearest to the large range of C. opimus.

The length-to-width ratio tends to be slightly greater in larger skulls, within either sex, and males differ from females (fig. 22) in being larger at a given estimated "age" or stage of development based on a composite length-to-width index (= CIL + CRL/2/ ((ZYB + LAB)/2)); abbreviations under Materials and Methods). Figure 22 compares CRL with the index in the Huancaroma sample of C. opimus. Most dimensions show this sexual difference to some degree. Considering external dimensions, based on visual comparisons of graphs, tail length and weight are seen to be clearly different between the sexes whereas head and body length, length of hind foot, and length of ear are not. Among the cranial dimensions, CIL, CRL, palatilar length (PRL), ZYB, LAB, interorbital breadth (IOB), and depth of skull (SKD) are noticeably different, whereas alveolar length of maxillary toothrow (MXL), palatal breadth (PRB), and dental span (DSB) are not.

COMPARISONS: In comparison with C. boliviensis, C. opimus with longer, softer, laxer, paler, more yellowish dorsal pelage and indistinct collar on throat; averaging smaller, and, relative to the condylobasilar length (CRL), having narrower interorbital breadth (IOB), breadth of palate between closest al-

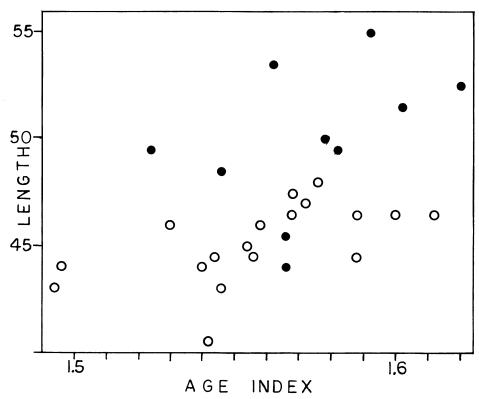


FIG. 22. Graph comparing males (dots) and females (circles) of *Ctenomys opimus* from near Huancaroma and illustrating the generally larger size (condylobasilar length of skull in this case) in males of a given degree of maturity or age, estimated as described in text.

veolar margins (PRB), and zygomatic breadth (ZYB). Based on direct comparison of skulls, C. opimus averaging smaller; incisors narrower, less procumbent, paler orange; nasals and rostrum narrower; zygomatic arches less massive, less widely flaring, bridge relative to position of anterior root more posterior; dorsal process of jugal more posterior; supraorbital process and ridge posterior to it present in both species, but less conspicuous in opimus; bullae much larger, lateral tubes placed more dorsally on bullae, gap between anterior processes about 2 rather than 3 or more mm; mesopterygoid fossa more acuminate and with large rather than small fenestrae visible on either side of the anterior basisphenoid suture.

In comparison with C. conoveri, C. opimus having long, fine, pale yellowish tan pelage with contrasting tips and gray bases of hairs both dorsally and ventrally; smaller; tail longer, hind foot longer, and ear longer, all

relative to HBL; and, relative to CRL, lesser palatilar length (PRL), ZYB, IOB, and greater breadth of dental span (DSB) and lambdoidal breadth (LAB). Based on direct comparison of skulls, C. opimus much smaller; breadth at postorbital constriction greater than at interorbital constriction, rather than the reverse; zygomatic arch more evenly bowed rather than with strong curvature of lateral profile in dorsal view of middle of arch; bullae relatively more inflated although inflated in both species; postorbital process conspicuous in both, but less extended in opimus; auditory tube relatively dorsal on bullae; anterior root of zygomatic arch projecting laterally rather than posterolaterally; supraoccipital plate more vertical rather than sloping anterodorsally to lambdoidal crest.

In comparison with Bolivian "C. minutus," C. opimus having paler, longer, laxer pelage; larger; hind foot longer relative to HBL; and, relative to CRL, having smaller

MXL and DSB and longer breadth of palate between closest alveolar margins (PRB). Based on direct comparison of skulls, *C. opimus* much larger; rostrum relatively longer; bullae smaller relative to size of skull.

In comparison with *C. steinbachi, C. opimus* without sharp border between venter and sides, and without distinct paler collar on throat; pale brown rather than glossy black; hair longer and softer; averaging smaller in HBL but not in CRL; longer skull, CRL, tail, and HF, relative to HBL; and, relative to CRL, lesser DSB and IOB. Based on direct comparison of skulls, *C. opimus* having rostrum narrower; eyes larger, supraorbital and dorsal jugal processes more posterior; interorbital area narrower; bullae larger; nasals narrower; zygomatic arches more widely bowed.

In comparisons of population samples within the species *C. opimus*, the Sajama series has a relatively longer tail than others, including the Rio Barros series.

Data on six samples of C. opimus were analyzed separately for males and females. Samples from six numbered localities used for figure 23 (and numbers of females in each sample) were: 1—near Huancaroma (19), 2 near Esperanza (3), 3-near Oruro (6), 4near Pomata Ayte (5), 5—near Cruce Ventilla (6), and 6—near ENDE camp (4). The percentages of correctly classified group memberships in an analysis of Canonical Discriminant Functions were: Group 1, 89.5; 2, 66.7; 3, 66.7; 4, 80.0; 5, 83.3; and 6, 100. Sufficient data on males were available for only two of the six geographic samples and the correctly predicted group memberships were: Group 1, 90.9, and 2, 100.

Although there is some geographic variation among local populations of *C. opimus* and the geographically most isolated sample (no. 6) is also the most isolated morphologically (fig. 23), we do not feel that recognition of more than one Bolivian subspecies is warranted at this time. The subspecies *C. o. luteolus* and *C. o. nigriceps*, known, respectively, from Argentina and Peru (following Cabrera, 1961: 554) need to be added to the analysis in order to evaluate the status of subspecies throughout the species' range.

DISTRIBUTION: Bolivian records are shown in figure 1. The species is reported to occur

north of Bolivia in Peru and south of Bolivia in Argentina, but we have not studied the specimens upon which these reports are based.

SPECIMENS, 137: Dept. of La Paz: 1749/ 6847, Esperanza, near Mt. Sahama, 5 BM, 14 FMNH. Dept. of Oruro: 1739/6728, 2.5 km NE of Huancaroma, 1 AMNH, 2 MSB (one karyotyped); 1740/6727, 3.5 km E of Huancaroma, 23 AMNH, 2 MSB (including 55378, NK 11513 F, 11541 F, 11568 M, 14765 F, 14766 F, 14767 M, 14768 F, 14769 F); 1740/6729, Huancaroma, 1 MSB; 1740/ 6732, 3 km W of Huancaroma, Río Desaguadero, 1 MSB (NK 14776 F, 14777 M, 14782 M); 1741/6731, 11.2 km S and 0.8 km W Eucaliptus, 2 AMNH; 1759/6707, 2 mi E Oruro, 1 MVZ; 1759/6709, Oruro, 9 BM (Thomas, 1902); 1807/6900, Sajama, 2 FMNH; 1819/6759, 5 km W and 1 km N of Pomata Ayte, Río Barros, 7 AMNH (including 263043), 3 MSB (including 57202, NK 14547 M, 14550 F, 14557 M, 14558 F, 14559 F); 1825/6844, 30 km S and 25 km E of Sajama, 2 AMNH, 1 MSB (NK 14542 F, 14546 M); 1854/6647, Challapata, 4 BM (Thomas, 1902); 1908/6607, 7 km S and 4 km E of Cruce Ventilla, 3 AMNH, 2 MSB (including 263031, NK 14740 M, 14745 F, 14747 F, 14748 F); 1908/6607, 7 km S and 3 km E Cruce Ventilla (NK 14750 F, 14751 F); 1908/ 6607, 6 km S and 4 km E of Cruce Ventilla, 1 MSB; 1910/6625, 45 km distant and on a compass bearing of 123° from Challapata, 2 AMNH; 1911/6705, Pampa Aullaga, 3 BM (Thomas, 1902). **Dept. of Potosi:** 1853/6628, Livichuco, 1 BM (Thomas, 1902); 1935/6545, Potosí, 4 ANSP (19348 to 19351), 5 BM (1902.2.2.99 to 103); 1939/6540, Caricari Mountains, also spelled Karikari, 13 ANSP; 2028/6650, Uyuni, 13 ANSP; 2115/6545, Chocaya, 1 ANSP; 2210/6742, 2 km E of ENDE camp, Laguna Colorado, 3 AMNH, 2 MSB (NK 14571 F, 14572 F, 14573 F, 14583 M); 2210/6745, 7 km E of ENDE camp, Laguna Colorado, 1 AMNH; 2218/6744, 21 km by road SE of ENDE camp, Laguna Colorado, 1 AMNH (NK 14584 F).

REMARKS: Waterhouse, 1848: 275, referred to "numerous specimens" brought to England by Mr. Bridges "from the neighborhood of the town of Potosi" and to "others in which the colouring was somewhat richer," the latter comment suggesting the pos-

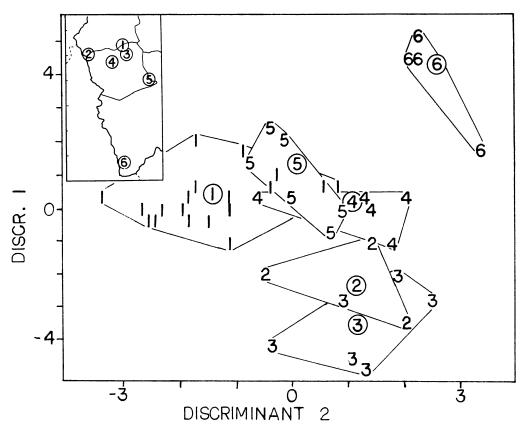


Fig. 23. Graph comparing samples of six different local populations of *Ctenomys opimus* as described in text. Values for individuals are indicated by uncircled numbers and sample centroids by circled numbers. The inset map of southwestern Bolivia shows the six geographic localities represented. The abscissa is the second discriminant function and the ordinate is the first discriminant function, based on analysis of females only.

sibility of C. frater. Waterhouse compared the skulls and found "no differences which would lead us to suppose there was more than one species amongst the Potosi specimens." The skull of one of these was figured on plate 8 (fig. 6) and it is clearly C. opimus and not C. frater. The skulls of these two species are sufficiently different that we suppose Waterhouse would not have made the above statement if both species were present in his sample; however, he did group the specimens from Potosí with specimens (from Maldonado, Uruguay, and elsewhere) that are not now considered to be conspecific. Waterhouse included measurements of four Bolivian specimens (external data on p. 276 and cranial data on p. 284) but cited no catalog numbers. We know of only four specimens of Ctenomys in the British Museum (Natural History) obtained by Bridges that came from, or might have come from, the vicinity of the city of Potosí. These are: 45.11.18.6 (this one simply from "Bolivia"), 45.11.18.7, 45.11.18.8 (the skull of which is numbered 45.12.8.2), and a skull with no registry number and with zygomatic breadth of 33.03 mm. We conclude that the four specimens noted are the four that Waterhouse provided measurements for, including the one figured, and that they are *C. opimus*. Waterhouse's measurements are either difficult to equate with ours or are not useful in distinguishing *C. opimus* from *C. frater*, the two species now known to occur at or near the city of Potosí.

Notes on Ctenomys conoveri, C. Stein-BACHI, AND "C. MINUTUS": Previously unidentified specimens reported earlier (Anderson et al., 1987) from Palo Marcado in

TABLE 1

Diploid Chromosome Numbers of Ctenomys

Country and species	2N			
ARGENTINA				
Ctenomys australis	46			
Ctenomys azarae	48			
Ctenomys latro	42			
Ctenomys magellanicus	36			
Ctenomys minutus	48, 50			
Ctenomys occultus	22			
Ctenomys opimus	26			
Ctenomys porteousi	48			
Ctenomys talarum	46, 48			
Ctenomys torquatus	68			
Ctenomys tuconax	61			
Ctenomys tucumanus	28			
Ctenomys haigi	50			
BRAZIL				
Ctenomys torquatus	44, 46			
Ctenomys minutus	46, 50, 54, 55			
Ctenomys flamarioni	48			
CHILE				
Ctenomys opimus	26			
Ctenomys robustus	26			
Ctenomys fulvus	26			
Ctenomys maulinus	26			
Ctenomys magellanicus	34			
URUGUAY				
Ctenomys torquatus	44			
Ctenomys pearsoni	56, 64, 70			
BOLIVIA				
Ctenomys opimus	26			
Ctenomys conoveri	48			
Ctenomys steinbachi	10			
Ctenomys boliviensis	36, 42, 44, 45, 46			
Ctenomys leucodon	36			
Ctenomys lewisi	56			
Ctenomys frater	52			

1) Argentina: Reig and Kiblisky, 1969; Pearson, 1984; Vidal-Rioja, 1985; 2) Brazil: Reig and Kiblisky, 1969; Freitas and Lessa, 1984; Freitas and Travi, 1982; 3) Chile: Gallardo, 1979; 4) Uruguay: Altuna and Lessa, 1985; Freitas and Lessa, 1984; Kiblisky et al., 1977; Novello and Lessa, 1986; 5) Bolivia: Anderson et al., 1987.

Tarija have been identified by Dr. Gerhard Storch, using characterizations given in our paper, as *C. conoveri*. Krumbiegel (1941: 131; quoted by Krieg, 1948: 119) cited specimens from Villa Montes, 21°15′S, 63°30′W, Tarija, under the name *Ctenomys leucodon* that are

C. conoveri, based on his description of the animals and on distributional considerations.

The legend of figure 8 in our earlier paper (Anderson et al., 1987) needs clarification. The male sex chromosomes of *Ctenomys steinbachi* were not mislabeled, as stated. The photo of the Y-chromosome is not to the same scale as the other chromosomes shown but is much more enlarged. The Y-chromosome is actually smaller than the X-chromosome.

We wish to stress here that the three small Bolivian specimens from near Roboré previously (Anderson et al., 1987) identified very tentatively as *Ctenomys minutus* are not of that species. Their identity will be dealt with in a later paper.

KARYOLOGICAL RESULTS

KARYOLOGY: Nondifferentially stained chromosomes from four nominal highland species, C. opimus (N = 29), C. leucodon (N = 3), C. frater (N = 5), and C. lewisi (N = 6) were examined. The diploid number (2N) varies among species (table 1), but intraspecific variation was not found in our samples, except for a questionable situation noted for C. leucodon. The number of autosomal arms (FN) range from 48 in C. opimus to 78 in C. frater.

Ctenomys opimus: Individuals from eight localities in Bolivia have a 2N of 26 and an FN of 48 (fig. 24). This karyotype is similar to that reported by Reig and Kiblisky (1969) for specimens collected in Jujuy Province, Argentina, and Gallardo (1979) for specimens collected in Tarapaca Province, Chile. The karyotype consists of three pairs of large metacentric, three pairs of medium metacentric, two pairs of medium subtelocentric, two pairs of small metacentric, and two pairs of small submetacentric autosomes. The morphology of the X-chromosome does not agree with the metacentric condition reported by Reig and Kiblisky (1969), but more closely resembles the submetacentric condition reported by Gallardo (1979). The Y-chromosome is minute and appears to be subtelocentric as reported by the above authors.

Ctenomys leucodon: Two females from two localities west of San Andres de Machaca have a 2N of 36 and an FN of 68 (fig. 24). This is

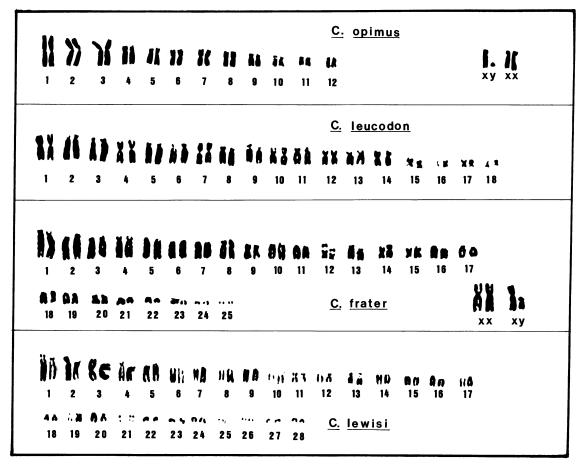


FIG. 24. Karyotypes (top to bottom) of *Ctenomys opimus* (catalog no. MSB 55376, NK 11568, and AMNH 260838, NK 11513, now in the Museo Nacional de Historia Natural in La Paz, for chromosomes of female); *C. leucodon* (AMNH 263028, NK 14793, female); *C. frater mordosus* (AMNH 263011, NK 14622, male; and 263012, NK 14610 for chromosomes of female); and *C. lewisi* (MSB 57186, NK 14653, female).

the first reported karyotype for this species. The karyotype is entirely biarmed with a fairly continuous gradation in size among the large and medium size chromosomes. These include four metacentric pairs and ten submetacentric or subtelocentric pairs. The remaining four small pairs appear to be metacentric. Cells of one male were examined and each of the 11 cells counted had 35 chromosomes. It is unclear whether this is an intraspecific polymorphism or results from variation in the sex chromosomes. Further banding studies and samples from other localities are needed to address this question.

Ctenomys frater: Individuals from two localities within the range of the subspecies C. f. mordosus have a 2N of 52 and an FN of 78 (fig. 24). This is the first reported karyotype for this species. The autosomal complement comprises two pairs of large subtelocentric chromosomes, 12 pairs of medium to small submetacentric to subtelocentric chromosomes, nine pairs of medium to small acrocentrics, and two pairs of micro or dot chromosomes. The X-chromosome is large and submetacentric and the Y-chromosome is small and submetacentric. Secondary constrictions were noted in chromosome pair 12. The constrictions presumably correspond to regions of nucleolar organization, although silver staining was not conducted.

Ctenomys lewisi: Individuals from the type

SKD

Dim.a	C. frater		C. opimus		C. leucodon		C. lewisi	
	26.1.1.126 Male	2.2.2.113 Female	55378 Male	57202 Female	1901.6.7.56 Male	263028 Female	26.1.1.116 Male	26.1.1.122 Female
H & B	192	160	251	201	200	172	150	133
TAI	76	72	89	81	85	79	69	71
HF	35	35	45	41	34	34	37	37
EAR	_	9	11	9	. 8	4	_	_
WT	_	_	457	280	_	_	_	_
CRL	41.8	40.8	55.2	49.7	47.3	44.4	47.8	45.2
CIL	46.6	45.0	59.3	53.9	52.4	49.4	55.4	50.7
PRL	21.2	21.0	29.2	24.1	23.8	22.9	25.2	24.5
MXL	11.6	10.6	12.4	11.2	11.0	11.5	11.2	11.6
DSB	10.8	10.4	10.2	11.5	10.6	11.4	10.5	9.9
PRB	2.0	2.2	3.0	2.7	2.6	2.3	2.1	2.0
ZYB	28.2	30.7	37.5	33.1	34.2	31.3	32.8	30.8
IOB	10.0	10.5	14.1	12.0	12.3	9.5	12.7	11.9
LAB	27.4	28.2	34.4	31.7	31.5	28.7	29.8	28.9

18.2

18.3

TABLE 2

Measurements of Four Species of Bolivian Highland Ctenomys

16.4

20.1

15.1

locality have a 2N of 56 and an FN of 74 (fig. 24). This is also the first reported karyotype for this species. All six individuals examined were females so the morphology of the sex chromosomes could not be determined. Four pairs are large and submetacentric to subtelocentric, 7 pairs medium to small and submetacentric to subtelocentric, 3 pairs large and acrocentric, 11 pairs medium to small and acrocentric, and 3 pairs micro chromosomes. Ctenomys lewisi and C. frater are the only Bolivian Ctenomys with microchromosomes although these appear to be common among some of the species of Ctenomys with higher diploid numbers (Kiblisky et al., 1977).

DISCUSSION

The genus *Ctenomys*, although found throughout the highlands of Bolivia, generally exhibits a patchy distribution typical of subterranean rodents. *Ctenomys opimus* ranges throughout the altiplano, while *C. leucodon* is restricted to the northern altiplano, and *C. frater* and *C. lewisi* occur in highlands southeast of the altiplano.

The wide geographic range of *C. opimus* is not accompanied by substantial morphometric or chromosomal variation. Analyses

of existing specimens suggest that in C. opimus some degree of morphological divergence has occurred among populations, but not to the extent seen in some other subterranean rodents such as North American pocket gophers of the genus *Thomomys* (as reflected by the numerous subspecies mapped by Hall, 1981). The Ctenomys opimus specimens used in this study are chromosomally monomorphic in populations from relatively distant parts of the range. Karyotypes of \hat{C} . opimus from Chile (Gallardo, 1979) and Argentina (Reig and Kiblisky, 1969) are similar to those of the Bolivian specimens, except for a possible slight variation in the X-chromosome morphology reported by Reig and Kiblisky (1969).

16.9

17.6

16.9

Ctenomyids are closely related to Octodontidae (including the genera Octodontomys, Octodon, Spalacopus, Aconaemys, Tympanoctomys, and Octomys). Pascual et al. (1965) and others have included Ctenomys (as the subfamily Ctenomyinae) in the Octodontidae and placed the other genera in the Octodontinae. Chromosomal variation in the octodontid genera (Spalacopus cyanus 2N = 58, Octodon degus 2N = 58, Octodontomys gliroides 2N = 38) was thought to be less than in Ctenomys until recent studies of other octodontids revealed extremely differ-

a Dimension. Abbreviations are spelled out in text.

ent karyotypes in *Tympanoctomys barrae* (2N = 102; Contreras and Torres-Mura, 1987) and *Octodon lunatus* (2N = 78; Spotorno et al., 1988). Together, the Octodontidae and Ctenomyidae range from 10 in *C. steinbachi* to 102 in *T. barrae*, and thus span nearly the entire range of diploid chromosomal variation known for mammals. The causes of this extreme variation are not clear.

Reig (1970) and Reig et al. (1972) hypothesized that the contrast in population structure between Ctenomys and the octodontid genera (e.g., Spalacopus) explained the increased chromosomal variability found in tuco tucos. These authors suggested that the subterranean Ctenomys have solitary habits and a low capacity for dispersal which has led to reduced gene flow and increased differentiation in this group, while the lack of chromosomal variation in octodontid species was due to increased dispersal and gene flow. It seems clear, with the recently reported karyotypes of Octodon lunatus and T. barrae. that a difference in population structure between Ctenomys and the octodontids is not a sufficient explanation for the karyotypic diversity found in these groups.

Diploid chromosomal numbers among the seven species of Bolivian Ctenomys range from 10 in C. steinbachi to 56 in C. lewisi. Intraspecific chromosomal variation also may be substantial as has been noted for C. boliviensis (Anderson et al., 1987), with five chromosomal forms known, although two of these may be distinct species. This amount of intraspecific variation is similar to that found in Spalax ehrenbergi (Wahrman et al., 1969, 1985) and Thomomys bottae (Patton, 1972; Hafner et al., 1983) complexes.

In general, however, species of *Ctenomys* exhibit less intraspecific chromosomal variation than other subterranean rodents. It is our subjective judgment, based on familiarity with *Thomomys* and other geomyids in North America, that intraspecific morphological variation also is less in *Ctenomys*, and that this difference will not disappear with further taxonomic revision. However, considering the complexity of the situation and our relatively greater ignorance of *Ctenomys*, this judgment is best considered to be a hypothesis for further testing. Chromosomal polymorphisms have been reported for only five *Ctenomys*

species (table 1), but the paucity of reported intraspecific polymorphisms may be due to limited sampling as relatively few species have been examined over wide geographic areas. It is also possible that populations that seem identical based on nondifferentially stained chromosomes will be found to differ structurally when examined using chromosome banding techniques, as has been noted for the subterranean Insectivora (Yates and Moore, in press).

Considerable care must be taken when attempting to draw systematic conclusions from the analysis of standard karyotypes (Baker et al., 1987), however, a few preliminary generalizations may be made from the data now available for Bolivian tuco tucos. Based strictly on diploid number, some species (e.g., C. boliviensis and C. steinbachi) that are geographically closest to each other do not appear to be sister taxa, while other geographically proximate species (e.g., C. lewisi and C. frater) are very similar karyotypically. If similarity of diploid number is representative of the history of chromosomal evolution in this group, then the historical biogeography of Ctenomys in Bolivia is more complex than a simple lowland/highland dichotomy. Although we had no a priori theoretical basis for predicting elevational or geographic correlations with diploid number, we thought it worth examining whether such correlations were noteworthy in the present case. Correlations are not noteworthy among the seven species that were examined. Low diploid numbers (< 30) are found in both the lowlands (C. steinbachi) and highlands (C. opimus) and the same is true for the higher diploid numbers.

Chromosomal variation among fossorial rodents (see Nevo, 1979; and Patton and Sherwood, 1983, for reviews) has received considerable attention in the literature, however, relatively little is understood about the mechanisms that allow this variation. Recent use of banding techniques has elucidated some of the types of rearrangements that have occurred in *Thomomys* (Patton and Sherwood, 1982; Barros and Patton, 1985), *Geomys* (Qumsiyeh et al., 1988), and *Spalax* (Wahrman et al., 1985). Preliminary G- and C-bands from Bolivian tuco tucos (Cook and Yates, unpubl. data) reveal that substantial changes,

including fission/fusion events and non-Robertsonian rearrangements, have taken place within the Bolivian group. Although the process-level explanations for chromosomal differentiation in subterranean rodents have generally focused on population structure (Patton, 1985) or environmental correlates (Nevo et al., 1988), Sites and Moritz (1987) listed other processes that might be important. Additionally, factors intrinsic to the genome (Greenbaum et al., 1986; Meyne et al., 1989, in press) may have a role in chromosomal evolution.

The extreme chromosomal variation found in *Ctenomys* (2N = 10-70), wide geographic distribution, and large number of species provide a prime mammalian example of explosive diversification with rapid rates of evolution as the earliest known fossils of *Ctenomys* are from the Pleistocene (Mones and Castiglioni, 1979; Frailey et al. 1980; Patterson and Wood, 1982). Detailed studies of this genus should provide powerful insights into the processes responsible for extensive chromosomal evolution.

A synthetic and eclectic approach is needed to help resolve the systematic relationships within the Ctenomyidae. In addition to expanded chromosome analyses (including G- and C-band studies) and the morphometric analyses presented in this paper, recent studies of protein electrophoretic variation (Sage et al., 1986), sperm morphology (Feito and Gallardo, 1982; Feito and Barros, 1982; Altuna et al., 1985; Vitullo et al., 1988), and phallic morphology (Altuna and Lessa, 1985) provide data sets that are useful in addressing the systematics of this diverse group. Brooks (1988) reviewed the evidence for phyletic coevolution in host/parasite systems and he and Kluge (1989) concluded that parasites may be used to develop strong hypotheses of host relationships. Phylogenetic relationships of the nematode parasites of the Bolivian tuco tucos have recently been proposed by Gardner (1988) and provide additional testable hypotheses of the host relationships. Due to the wide geographic range and large number of species found in this group, Ctenomys may be an excellent model to use in developing hypotheses of both biogeographic and faunal relationships in Bolivia and the southern cone of South America.

REFERENCES

Altuna, C. A., and E. P. Lessa

1985. Penial morphology in Uruguayan species of *Ctenomys* (Rodentia: Octodontidae). J. Mammal, 66: 483–488.

Altuna, C. A., C. F. Novello, and E. P. Lessa

1985. Notas sobre la morfología espermática de *Ctenomys rionegrensis* (Rodentia, Octodontidae) del Uruguay. Brenesia 24: 397–401.

Anderson, S.

1985. Lista preliminar de mamiferos bolivianos. Cuadernos, Acad. Nac. Cienc. Bolivia, vol. 65, Cienc. Naturaleza, no. 6, Mus. Nac. Hist. Nat., Zoologica, no. 3, pp. 5-16, December.

Anderson, S., T. L. Yates, and J. A. Cook

1987. Notes on Bolivian mammals 4: the genus *Ctenomys* (Rodentia: Ctenomyidae) in the eastern lowlands. Am. Mus. Novitates 2891: 20 pp.

Baker, R. J., M. B. Qumsiyeh, and C. S. Hood 1987. Role of chromosomal banding patterns in understanding mammalian evolution. *In* H. H. Genoways (ed.), Current mammalogy 1: 67–96. New York: Ple-

num.

Barros, M. A., and J. L. Patton

1985. Genome evolution in pocket gophers (genus *Thomomys*) III. Flourochrome revealed heterochromatin heterogeneity. Chromosoma 92: 337–343.

Blainville, H. M. D. de

1826. Sur une nouvelle espèce de Rongeur Fouisseur du Brèsil. Nouv. Bull. Soc. Philomatique Paris 3: 62–64 (not seen). Also published with same title in Ann. Sci. Nat., Zool. Paleontol. 9: 97–104.

Brooks, D.

1988. Macroevolutionary comparisons of host and parasite phylogenies. Ann. Rev. Ecol. Syst. 19: 235-259.

Cabrera, A.

1961. Catalogo de los mamiferos de America del Sur. Rev. Mus. Argentino Cienc. Nat. "Bernardino Rivadavia," Buenos Aires, 4(2): xxii + 308-732.

Contreras, L. C., and J. C. Torres-Mura

1987. Tympanoctomys, un mamifero deserticola con el numero diploide mas alto. Arch. Biol. Med. Exp. 20: pages unnumbered, resumen no. R-190.

Feito, R., and C. Barros

1982. Ultrastructure of the head of *Ctenomys* maulinus spermatozoon with special reference to the nucleus. Gamete Res. 5: 317–321.

Feito, R., and M. Gallardo

1982. Sperm morphology of the Chilean species of *Ctenomys* (Octodontidae). J. Mammal. 63: 658–661.

Frailey, D., K. E. Campbell, and R. G. Wolff

1980. Additions to the knowledge of *Hippocamelus, Ctenomys, Myocaster* from the Middle Pleistocene of the Tarija Basin, Bolivia. Occas. Pap. Mus. Nat. Hist. Univ. Kansas 85: 1–14.

Freitas, T. R. O., and E. P. Lessa

1984. Cytogenetics and morphology of *Ctenomys torquatus* (Rodentia: Octodontidae). J. Mammal. 65: 637–642.

Freitas, T. R. O., and V. H. Travi

1982. Estudo citotaxonomico e distribuicao geografica de tres especies do genero *Ctenomys* no Rio Grande do Sul (Rodentia-Ctenomyidae). Cienc. Cult. 34: 755 (Proceedings). [Not seen; cited in S. Kasahara and Y. Yonenaga-Yassuda. 1984. Rev. Genet. 7(3): 509–533.]

Gallardo, M.

Las especies chilenas de Ctenomys (Rodentia, Octodontidae). I. Estabilidad cariotípica. Arch. Biol. Med. Exp. 12: 71-82.

Gardner, S. L.

1988. Coevolution and zoogeography of helminth parasites and their rodent hosts in Bolivia. Ph.D. diss., University of New Mexico. 180 pp.

Greenbaum, I. F., D. W. Hale, and K. P. Fuka 1986. Synaptic adaptation in deer mice: a cellular mechanism for karyotypic orthoselection. Evolution 40: 208–213.

Hafner, J. C., D. J. Hafner, J. L. Patton, and M. F. Smith

1983. Contact zones and the genetics of differentiation in the pocket gopher, *Thomomys bottae* (Rodentia: Geomyidae). Syst. Zool. 32: 1–20.

Hall, E. R.

1981. The mammals of North America, 2nd ed. New York: Wiley, vol. 1, pp. 1–600 + 90.

Honacki, J. H., K. E. Kinman, and J. W. Koeppl 1982. Mammal species of the world. Lawrence, Kansas: Allen Press and Assoc. Syst. Colls., ix + 694 pp.

Kiblisky, P., N. Brum-Zorrilla, G. Perez, and F. A. Saez

1977. Variabilidad cromosómica entre diversas poblaciones uruguayas del roedor cavador del género *Ctenomys* (Rodentia, Octodontidae). Mendeliana 2: 85-93.

Kluge, A. J.

1989. A concern for evidence and a phylogenetic hypothesis of relationships among *Epicrates* (Boidae, Serpentes). Syst. Zool. 38: 7-25.

Krieg, H.

1948. Zwischen Anden und Atlantik. Munchen: Carl Hanser Verlag, 492 pp.

Krumbiegel, I.

1941. Die Säugetiere . . . Dr. Kriegs. 13.Kammraten (Ctenomys). Zool. Anz.135: 18-26.

Lessa, E. P., and J. A. Cook

1989. Interspecific variation in penial characters in the genus *Ctenomys* (Rodentia: Octodontidae). J. Mammal. 70: 856–860.

Mares, M. A., and R. A. Ojeda

1982. Patterns of diversity and adaptation in South American hystricognath rodents. In M. A. Mares and H. H. Genoways (eds.), Mammalian biology in South America. The Pymatuning Symposia in Ecology, Pymatuning Laboratory of Ecology, University of Pittsburgh, Spec. Publ. Ser., 6: 393-432.

Meyne, J., R. J. Baker, H. H. Hobart, T. C. Hsu, O. A. Ryder, O. G. Ward, J. E. Wiley, D. H. Wurster-Hill, T. L. Yates, and R. C. Moyzis

In press. Distribution of non-telomeric sites of the (TTAGGG)n telomere sequence in vertebrate chromosomes. Chromosoma.

Meyne, J., R. C. Ratcliff, and R. K. Moyzis

1989. Conservation of the human telomere sequence (TTAGGG)n among vertebrates. Proc. Natl. Acad. Sci. USA 86: 7049-7053.

Mones, A., and L. R. Castiglioni

1979. Additions to the knowledge on fossil rodents of Uruguay (Mammalia: Rodentia). Palaeontol. Z. 53(1-2): 77-87.

Neveu-Lemaire, M., and G. Grandidier

1911. Notes sur les mammiferes des hauts plateaux de l'Amerique du Sud. Paris: Le Sondier, viii + 127 pp.

Nevo, E.

1979. Adaptive convergence and divergence of subterranean mammals. Ann. Rev. Ecol. Syst. 10: 269–308.

Nevo, E., M. Corti, G. Heth, A. Beiles, and S. Simson

1988. Chromosomal polymorphisms in subterranean mole rats: origins and evolutionary significance. Biol. J. Linn. Soc. 33: 309-322.

Novello, A. F., and E. P. Lessa

1986. G-band homology in two karyomorphs

of the *Ctenomys pearsoni* complex (Rodentia: Octodontidae) of neotropical fossorial rodents. Z. Saeugetierkd. 51: 378–380.

Osgood, W. H.

1916. Mammals of the Collins-Day South American Expedition. Field Mus. Nat. Hist. Zool. 10: 199–216, + 2 plates.

Pascual, R., J. Pisano, and E. J. Ortega

1965. Un nuevo Octodontidae (Rodentia, Caviomorpha) de la formacion Epecuen (Plioceno Medio) de Hidalgo (Provincia de la Pampa). Ameghiniana Rev. Asoc. Paleontol. Argentina 4: 19–30.

Patterson, B., and A. E. Wood

1982. Rodents from the Deseadan Oligocene of Bolivia and the relationships of the Caviomorpha. Bull. Mus. Comp. Zool. Harvard Coll. 149: 371-543.

Patton, J. L.

1972. Patterns of geographic variation in karyotype in the pocket gopher, *Thomomys bottae* (Eydoux and Gervais). Evolution 26: 574-586.

1985. Population structure and the genetics of speciation in pocket gophers, genus *Thomomys*. Acta Zool. Fennica 170: 109-114.

Patton, J. L., and S. W. Sherwood

1982. Genome evolution in pocket gophers (genus *Thomomys*) I. Heterochromatin variation and speciation potential. Chromosoma 85: 149-162.

1983. Chromosome evolution and speciation in rodents. Ann. Rev. Ecol. Syst. 14: 139–158.

Pearson, O. P.

1984. Taxonomy and natural history of some fossorial rodents of Patagonia, Southern Argentina. J. Zool. London 202: 225-237.

Qumsiyeh, M. B., C. Sanchez-Hernandez, S. K. Davis, J. C. Patton, and R. J. Baker

1988. Chromosomal evolution in *Geomys* as revealed by G- and C-band analysis. Southwest. Nat. 33: 1-13.

Reig, O. A.

1970. Ecological notes on the fossorial octodont rodent *Spalacopus cyanus* (Molina). J. Mammal. 51: 592-601.

1986. Diversity patterns and differentiation of high Andean rodents. In F. Vuilleumier and M. Monasterio (eds.), High altitude tropical biogeography, pp. 404– 442. New York, Oxford: Oxford Univ. Press.

Reig, O. A., and P. Kiblisky

1969. Chromosome multiformity in the genus

Ctenomys (Rodentia: Octodontidae). A progress report. Chromosoma 28: 211–244.

Reig, O. A., O. A. Spotorno, and D. R. Fernandez
1972. A preliminary survey of chromosomes in populations of the Chilean burrowing octodont rodent Spalacopus cyanus
Molina (Caviomorpha, Octodontidae).
Biol. J. Linn. Soc. 4: 29-38.

Rusconi, C.

1928. Dispersión geográfica de los Tuco-tucos vivientes (Ctenomys) en la región neotropical. An. Soc. Argent. Est. Geogr. "Gaea" 3: 235–250.

Sage, R. D., J. R. Contreras, V. G. Roig, and J. L. Patton

1986. Genetic variation in the South American burrowing rodents of the genus *Ctenomys* (Rodentia: Ctenomyidae). Z. Saeugetierkd. 51: 158-172.

Sites, J. W., Jr., and C. Moritz

1987. Chromosomal evolution and speciation revisited. Syst. Zool. 36: 153–174.

Spotorno, A., L. Walker, L. Contreras, J. Pincheira, and R. Fernandez-Donoso

1988. Cromosomas ancestrales en Octodontidae y Abrocomidae. Soc. Genet. Chile., pages unnumbered, resumen no. R-65.

Thomas, O.

1900. Descriptions of new rodents from western South America. Ann. Mag. Nat. Hist., ser. 7, 5: 383-387.

1902. On mammals collected by Mr. Perry O. Simons in the southern part of the Bolivian Plateau. Ibid., ser. 7, 9: 222-230.

1919. Two new rodents from Tartagal, Salta, N. Argentina. Ibid., ser. 9, 4: 154–156.

1925. The Spedan Lewis South American Exploration. I. On mammals from southern Bolivia. Ibid., ser. 9, 15: 575-582.

1926. The Spedan Lewis South American Exploration. II. On mammals collected in the Tarija Department, southern Bolivia. Ibid., ser. 9, 17: 318–328.

1927. A selection of lectotypes of American rodents in the collection of the British Museum. Ibid., ser. 9, 19: 545-554.

Vidal-Rioja, L.

1985. Chromosome polymorphism in *Ctenomys talarum* (Rodentia: Octodontidae). Caryologia 38: 169–178.

Vitullo, A. D., E. R. S. Roldan, and M. S. Merani 1988. On the morphology of spermatozoa of tuco-tucos, *Ctenomys* (Rodentia: Ctenomyidae): new data and its implications for the evolution of the genus. J. Zool., London, 215: 675-683. Wagner, A.

1848. Beiträge zur Kenntniss der Arten von Ctenomys. Arch. Naturg. Jahrg. 14, Bd. 1: 72–78.

Wahrman, J., R. Goitein, and E. Nevo

1969. Geographic variation of chromosome forms in *Spalax*, a subterranean mammal of restricted mobility. *In* K. Benirschke (ed.), Comparative mammalian cytogenetics, pp. 30–48. New York: Springer-Verlag.

Wahrman, J., C. Richler, R. Gamperl, and E. Nevo 1985. Revisiting *Spalax*: Mitotic and meiotic chromosome variability. Israel J. Zool. 33: 15–38.

Waterhouse, G. R.

1848. A natural history of the mammalia. Vol.

2, Rodentia. London: Hippolyte Bailliere, 500 pp. + 22 plates.

Woods, C. A.

1984. Hystricognath rodents. In S. Anderson and J. K. Jones, Jr. (eds.), Orders and families of Recent mammals of the world, pp. 389-446. New York: Wiley.

Yates, T. L., and D. W. Moore

In press. Speciation and evolution in the family Talpidae (Mammalia: Insectivora). *In* Subterranean mammals, E. Nevo and O. Reig, eds.

Yepes, J.

1930. Los roedores octodontinos con distribucion en la zona cordillerana de Chile y Argentina. Rev. Chilena Hist. Nat. 34: 321-331.

Recent issues of the Novitates may be purchased from the Museum. Lists of back issues of the Novitates, Bulletin, and Anthropological Papers published during the last five years are available free of charge. Address orders to: American Museum of Natural History Library, Department D, Central Park West at 79th St., New York, N.Y. 10024. THIS PUBLICATION IS PRINTED ON ACID-FREE PAPER.