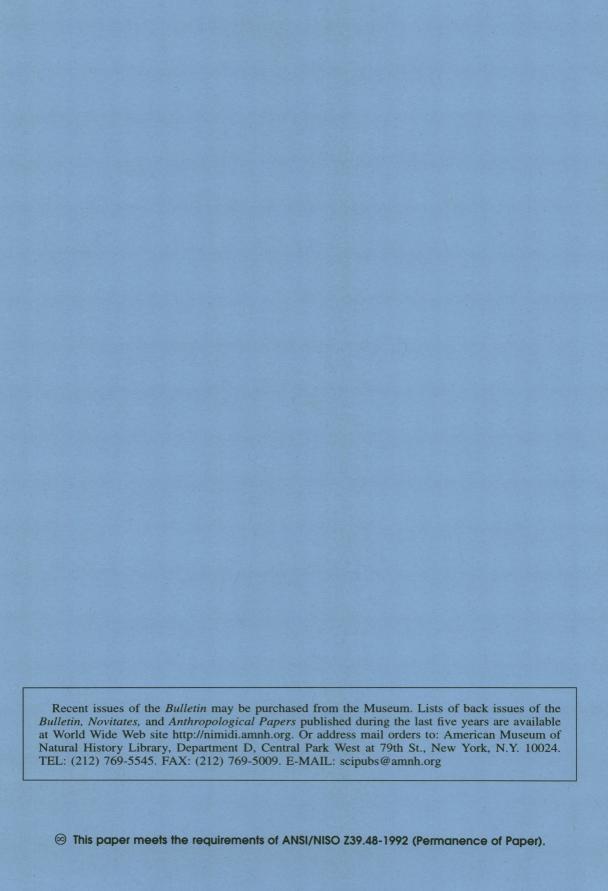
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ABSTRACT

This paper presents a phylogenetic analysis of the Asteropyginae Delo, 1935. Sixty-six characters and 39 taxa were employed in a cladistic analysis that produced a single most parsimonious tree that is presented as a hypothesis of asteropyginid relationships. In addition to the phylogeny generated, 11 new genera are diagnosed: Philipsmithiana, Coltraneia, Stummiana, Bellacartwrightia, Deloops, Tolkienia, Braunops, Armorigreenops, Kennacryphaeus, Modellops, and Hallandclarkeops, and 12 new species are described: Philipsmithiana hyfinkeli, P. burtandmimiae, Armorigreenops leoi, Pelitlina smeenki, Bellacartwrightia jennyae, B. whiteleyi, B. phyllocaudata, B. calderonae, Greenops widderensis, G. barberi, G. grabaui, and Kennacryphaeus harrisae.

Traditionally, asteropyginine taxa in the Appalachian and Michigan Basins of Eastern North America had been assigned to *Greenops* Delo, 1935, or *Greenops* (Neometacanthus) Richter and Richter, 1948. A core of five Eastern North American species—Greenops boothi (Green, 1837), G. widderensis, new species, G. chilmanae Stumm, 1965, G. grabaui, new species, and G. barberi, new species—can still be assigned to Greenops. All other asteropyginine taxa in Eastern North

America must be assigned to different genera. Species referable to *Neometacanthus* Richter and Richter, 1948, may be known from the Illinois Basin of Eastern North America, and a species referable to *Tolkienia*, new genus, may be known from the Michigan Basin of Eastern North America

The single most parsimonious cladogram was also used to investigate biogeographic patterns. In particular, the number of times that independent lineages of asteropyginines invaded from what are now Europe and North Africa (called Armorica herein) into Eastern North America during the Devonian was ascertained. Phylogenetic analysis suggested that there must have been at least four such invasions, with clades that had primitive European or North African affinities giving rise to Eastern North American taxa, although no species are shared between Armorica and Eastern North America. One of the lineages that invaded Eastern North America subsequently reinvaded Armorica. The timing of these invasions and their relation to overall patterns of faunal evolution in the Middle Devonian of Eastern North America are also discussed. These patterns suggest that the different taxa that make up a fauna often arrive during different time intervals, not all at once.

INTRODUCTION

The Asteropyginae Delo, 1935, form an important component of Devonian trilobite diversity. The group persisted from the Lockhovian to the Frasnian (Feist, 1991) or possibly Famennian (data herein) and consisted of approximately 225 species. Species occur in strata of Eastern North America (ENA), Armorica (comprising central Europe, northern Africa, Iberia, Turkey, and Afghanistan, although some reconstructions such as that of Van der Voo [1988] view Armorica as not including northern Africa), Burma, and Venezuela, and the group attained its peak diversity in the upper Emsian (Morzadec, 1992). This paper presents a phylogenetic analysis of a large component of the generic diversity within the asteropyginid clade. Special attention is paid to the evolutionary position of the Laurentian asteropyginines, and the putative monophyly of these taxa typically assigned to the genus *Greenops* in ENA is evaluated. The relationship of Armorican taxa to Laurentian taxa in ENA, the North American part of Oliver's (1976, 1977) Eastern Americas Realm, is also discussed. Twelve new species are described, 11 new genera are diagnosed, and a parsimony analysis of the Asteropyginae is conducted using 66 characters and 39 taxa.

Before phylogenetic patterns in the Asteropyginae can be analyzed, it is necessary to place that subfamily in a higher level phylogenetic context. Following the arguments and the character evidence presented by Edgecombe (1993) and Ramsköld and Edgecombe (1993), the Asteropyginae are assigned to the superfamily Acastacea Delo, 1935, family Acastidae Delo, 1935. When considering the subfamilial relationships within the Acastacea several authors (e.g., Struve, 1959; Pillet, 1961; Gandl, 1972; Eldredge, 1979; Timm, 1981; Smeenk, 1983; Tomczykowa, 1991; Edgecombe, 1991, 1993; Ramsköld and Edgecombe, 1993) have noted that the Acastavinae Struve, 1958a, appear to be closely related, or the sister taxon,

to the Asteropyginae, with both of these taxa typically placed in the Acastacea. The Acastavinae, as traditionally circumscribed, contains the genera Acastava Richter and Richter, 1954, Acastella Reed, 1925, Acastellina Richter and Richter, 1954, Acastocephala Shergold, 1966, Pelitlina Haas, 1968, Centauropyge Haas, 1968, Mimocryphaeus Gandl. 1972. Chimaerastella Gandl. 1972. and Acastopyge Tomczykowa in Tomczykowa and Witwicka, 1974. These taxa were treated as closely related because they possessed genal and pygidial marginal spines larger than those in Acastinae Delo, 1935, but not as large as those of the Asteropyginae (Ramsköld and Edgecombe, 1993). Edgecombe (1993) and Ramsköld and Edgecombe (1993) explicitly recognized that these taxa formerly assigned to the Acastavinae comprise a paraphyletic stem lineage to the Asteropyginae, although these authors did not define a monophyletic Acastavinae. Herein, a detailed analysis considering the formulation of a monophyletic Acastavinae is not pursued, but a species formerly placed within that paraphyletic grade is employed as an outgroup in the phylogenetic analysis.

There has been some disagreement on which taxa within the paraphyletic "Acastavinae" (using Wiley's [1979] quotation marks convention for paraphyletic taxa) are most closely related to true Asteropyginae and thus are the most apt choice as an outgroup for polarization of characters in an analysis of the ingroup Asteropyginae. Certain authors, such as Edgecombe (1991) and Ramsköld and Edgecombe (1993), have suggested that species of Acastava or Pelitlina are the last common ancestors of the Asteropyginae, whereas other authors have suggested that species of Acastella are immediately ancestral to the Asteropyginae (e.g., Timm, 1981; Smeenk, 1983; Tomczykowa, 1991). Finally, Gandl (1972) suggested that the Asteropyginae were polyphyletic, with part of the "clade" derived from his "Acastella" new species aff. tiro Richter and Richter, 1954, and the other part from other taxa in the "Acastavinae." The only one of these hypotheses of evolutionary relationship that is rigorously couched in terms of synapomorphous traits is that of Edgecombe (1991) and Ramsköld and Edgecombe (1993), and the conclusion of their analysis, that Acastava and Pelitlina are more closely related to the Asteropyginae than Acastella sensu stricto, is followed herein. Ramsköld and Edgecombe (1993) presented a series of characters that indicate that Acastella sensu stricto is sister to Scotiella Delo, 1935, to the exclusion of the other "Acastavinae," specifically Acastava and Pelitlina, which are themselves sister to the Asteropyginae.

When considering both Acastava and Pelitlina in relation to the Asteropyginae, it seems that species of Pelitlina bear at least two synapomorphous traits uniting this genus with the Asteropyginae to the exclusion of Acastava. The type of Acastava, A. atavus (Schmidt, 1907), has five pairs of small marginal denticles bordering the pygidium, and these denticles are also found in all of the species of Acastella, which in addition typically possess a long terminal pygidial spine. In both Pelitlina goltzi Haas, 1968, and Pelitlina smeenki, new species (formerly Acastava new species in Smeenk [1983]), there are five relatively broad pairs of marginal pygidial lappets bordering the pygidium. These broad marginal pygidial lappets are found in all members of the Asteropyginae, with five pairs of lappets always found except for a few exceptions in which four pairs are present, e.g., species of Neocalmonia Pillet, 1969. These shared characters are taken as evidence of evolutionary relationship. However, it is interesting to note the parallel development of this character in certain members of the Calmoniidae Delo, 1935, in particular, species of the "Metacryphaeus Group" (see Lieberman et al. [1991] and Lieberman [1993]). In addition to this synapomorphy of the pygidium, the cephalic border furrow extends onto the genal spine, partially bifurcating it, in both *Pelitlina* and the Asteropyginae (Edgecombe, 1991). Three characters are treated as synapomorphies of the Asteropyginae, with Pelitlina treated as a near basal member of the subfamily rather than a member of the "Acastavinae," and it is used as the outgroup in phylogenetic analysis of the Asteropyginae. A sister group relationship between the Asteropyginae and Pelitlina suggests that Pelitlina could be assigned to a new monotypic subfamily. This is a reasonable taxonomic solution; however.

it is not followed at this time because designation of a new subfamily will not improve our understanding of phylogenetic patterns within the Acastacea.

At this time, "Acastavinae" is not revised, and it is placed in the family Acastidae Delo, 1935, along with Asteropyginae and Acastinae, following Pillet (1961), Holloway and Neil (1982), Cooper (1982), and Edgecombe (1991). The Acastidae can be diagnosed by the transverse S1 and S2 (Edgecombe, 1991), and L1 shorter than L2 (Destombes and Henry, 1987; Edgecombe, 1991).

For purposes of phylogenetic analysis, Pelitlina smeenki, new species, was used to assess the primitive conditions of characters used to deduce evolutionary relationship except for the condition of one character. This character, number 24, describes the condition of the prosopon. In Pelitlina smeenki, the prosopon is dominated by fine granules, and in P. goltzi Haas, 1968, the prosopon is dominated by fine tubercles. This character was coded as "?" for the outgroup. Aside from this character, Pelitlina smeenki and P. goltzi are nearly identical except for: the condition of S2, which extends further laterally in P. goltzi; the lateral margins of the cephalon, which deflect more sharply posteriorly in P. goltzi; and the posterior portions of the pygidial lappets, which are pointed in P. smeenki but bluntly rounded in P. goltzi.

On the basis of phylogenetic analysis, species of Pelitlina can be discriminated from other members of Asteropyginae on the basis of their possession of the primitive state for the following characters, which map on the tree in the derived state at the node separating the rest of the Asteropyginae from Pelitlina: 4, 8, 9, 20, 26, 29, 33, 36, 46, 57, 65, and 66. All of these characters show some homoplasy in the Asteropyginae, with reversions to the primitive condition for certain taxa (see tables 1 and 2 for specific cases), except for characters 26 and 46. Thus, Pelitlina can be distinguished from all other members of the Asteropyginae by its very short genal spines and by the condition of the pygidial axis. In Pelitlina smeenki it constricts posterior to the third axial ring, whereas in all other members of the Asteropyginae it constricts at various positions behind the fourth axial ring.

Where possible, type species of genera within the Asteropyginae were used in the phylogenetic analysis. When types could not be obtained, generic assignments reflect the possession of characters that diagnose the genus. Generic names follow Struve (1959), Haas (1968, 1970), Haas and Mensink (1969), Gandl (1972), and Morzadec (1983, 1992). Morphological terminology follows Eldredge and Branisa (1980) and Lieberman et al. (1991).

Several of the taxa considered herein, including species in the genera Stummiana. new genus, Bellacartwrightia, new genus, Greenops, Asteropyge, and Kayserops, have a peculiar set of small holes on the thoracic and pygidial pleural fields. Stumm (1953) termed these holes circular pits. He identified them as present in Kayserops traversensis (Stumm, 1953), and he suggested that they were caused by the weathering of coarse tubercles. These pits actually pass all the way through the exoskeleton and are present in specimens that have tubercles preserved as well as in specimens of taxa that lack coarse tubercles. Thus, Stumm's (1953) claim that weathering of tubercles explains these morphological structures is untenable. Herein, we treat these holes as structurally and perhaps functionally similar to the anatomical features Ramsköld (1991) termed fenestrae. As Ramsköld (1991) recognized, there are odontopleurids, dalmanitids, and asteropyginines that bear these fenestrae. Each of these represent lineages that must have acquired these structures independently, and thus they cannot be viewed as homologous. We speculate that these fenestrae may have performed some sensory function, possibly with setae passing through them.

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ASTEROPYGINAE IN EASTERN NORTH AMERICA

The Asteropyginae represent an important component of the trilobite fauna found in the Middle Devonian Hamilton Group strata of ENA. ENA includes the Appalachian, Illinois, and Michigan Basins (see fig. 1) as well as the Hudson Bay lowlands (not shown and not considered). The Hamilton Group strata span the latest Eifelian through Givetian and have been the subject of intense stratigraphic study (e.g., Cooper et al., 1942; Brett, 1986; Brett and Baird, 1986; Rickard, 1989). The Hamilton Group is paradigmatically developed in New York, but coeval and very similar deposits, in terms of their faunas, are known from the Illinois and Michigan Basins. Stratigraphic columns for the Middle Devonian and the Hamilton and Traverse Groups are shown in figure 2. Although this trilobite fauna is not incredibly rich in diversity, individual specimens are in places very common, and trilobite taxa have an important role in paleontological, stratigraphic, and macroevolutionary studies. After species assigned to the genus *Phacops* Emmrich, 1839, those taxa assigned to the genus Greenops Delo, 1935, form the most abundant and important members of this trilobite fauna. For this reason, the phylogenetic relationships of these asteropyginids is of intrinsic interest and of direct relevance to hypotheses about the origin of the Hamilton Group fauna in particular and the origin of evolutionary faunas in general.

Burton and Eldredge (1974) substantiated an ancestral-descendant relationship between Phacops schlotheimi (Bronn) sensu lato in the latest Eifelian and Givetian of northwestern Africa and subspecies of Phacops rana (Green, 1832) in the earliest part of the Hamilton Group. This notion was originally developed in Hall and Clarke (1888) and later in Eldredge (1972). In addition, both Eldredge (1972) and Burton and Eldredge (1974) recognized the close relationship between Asteropyginae in the Hamilton Group of ENA and those found in northwest Africa. Spain, and Germany (Armorica). The pattern recognized by Eldredge (1972) and Burton and Eldredge (1974) for Phacops conforms with broader macroevolutionary hypotheses about the origin of major faunas in the fossil record discussed in Eldredge (1985, 1989, 1995) Lieberman (1994), Brett et al. (1990), and Brett and Baird (1995). For instance, Brett et al. (1990) and Brett and Baird (1995) suggested that the Paleozoic fossil record of New York consists of approximately 13 fossil faunas spanning the Ordovician to the Devonian. Each of these fossil faunas persists for roughly 5-10 million years and consists of about 200 largely unique "species-level" taxa; very few species are shared between faunas. A fauna appears in the record rela-



Fig. 1. A paleogeographic reconstruction of Eastern North America modified from Beaumont et al. (1988) showing the extent of the major tectonic basins during the Middle Devonian mapped onto a modern geographic map of the United States. "1" represents the Appalachian Basin, "2" the Michigan Basin, and "3" the Illinois Basin.

tively rapidly and vanishes with often greater rapidity. This truncation typically appears to be associated with an extinction event of varying intensity on a global scale.

In addition to the biostratigraphic patterns predicted above by Eldredge (1985, 1989), Brett et al. (1990), and Brett and Baird (1995) Brett, Baird, and Eldredge made important predictions about the genealogical relationships among the component taxa of underlying and superposed faunas. They suggested that the appearance of new species in an overlying fauna does not seem to be the by-product of in situ evolution or evolutionary turnover of clades within a particular geographic region. Rather, they claimed that new taxa appearing in a younger fauna generally are most closely related to taxa in a different geographic region rather than to the taxa in the underlying fauna. This, of course,

would indicate that taxa had invaded from a different geographic region. Thus, associated with transitions between faunas, these authors predicted that there should be a significant change in the biogeographic area relationships of component taxa in faunas.

The Asteropyginae appear to offer an excellent opportunity to test and validate these hypotheses of faunal evolution. Additional tests of these hypotheses, which are discussed later in this paper, can be found in Lieberman (1994). Only two species of asteropyginines, Bellacartwrightia pleione (Hall, 1861) and B. jennyae, new species, are known with certainty from the strata underlying the Hamilton Group and its equivalents in the Appalachian, Michigan, and Illinois Basins. In these underlying strata, referred to as the Onondaga Limestone in New York, the roughly coeval Amherstburg Formation in the Michigan Basin, and the Jeffersonville Limestone in the Illinois Basin, the bulk of the most closely related taxa all belong to the Dalmanitacea. These taxa, assigned to the genera Odontocephalus Conrad. 1840. Anchiopsis Delo, 1935, Coronura Hall and Clarke, 1888, and Trypaulites Delo, 1935, probably did not share a common ancestor with the Asteropyginae after the Ordovician. No other closer relatives of the Asteropyginae are known from ENA in the Middle Devonian. Thus, it is obvious that the Asteropyginae must have originated outside of ENA and subsequently invaded that region before or around the time of the deposition of the Onondaga Limestone and its equivalents, which are late Emsian through Eifelian in age. The actual timing of the invasions by the Asteropyginae into ENA, and their biogeographic sources, are discussed in greater detail in the sections of this paper on biogeography and faunal evolution.

There has been a significant number of phylogenetic studies performed on the Asteropyginae, chiefly by European authors, e.g., Haas (1970), Gandl (1972), Timm (1981), Morzadec (1983), and Smeenk (1983). Most of these analyses used the principles of evolutionary taxonomy, which unites taxa on the basis of information about overall morphological similarity as well as vertical and horizontal distributions in the fossil record. None of these analyses consid-

		NEW Y	ORK	EUROPEAN STAGES	MAJOR LITHO- STRATIGRAPHIC UNITS IN ENA
	D		TAGHANIC		TULLY LIMESTONE
M	E		TIOUGHNIOGA	GIVETIAN	HAMILTON GROUP
D	v 0	ERIAN	CAZENOVIA	<u></u> ? ? ?	
D	N _		SOUTHWOOD	EIFELIAN	ONONDAGA LIMESTONE
E	A			7 7 7	
LOV	VER	ULSTERIAN	SAWKILL	EMSIAN	BOIS BLANC LS./ SCHOHARIE GRIT

G R O U P	FORMATION	MEMBER
	TULLY LIMESTONE	TULLY LIMESTONE / GILBOA
н		WINDOM SHALE KASHONG SHALE
	MOSCOW	MENTETH LIMESTONE
	MOSCOW	DEEP RUN SHALE
		TICHENOR LIMESTONE
A		JAYCOX SHALE
		SPAFFORD SHALE
М	LUDLOWVILLE	WANAKAH SHALE
	LODLOWVILLE	LEDYARD SHALE
,		CENTERFIELD LIMESTONE / STONE MILL
		BUTTERNUT
L	SKANEATELES	POMPEY
	OKAN EATEES	DELPHI STATION
т		STAFFORD LIMESTONE / MOTTVILE
o	MARCELLUS	OATKA CREEK SHALE/ CHITTENANGO
N		UNION SPRINGS / CHERRY VALLEY

Fig. 2. Stratigraphic columns modified from Cooper et al. (1942), Kirchgasser et al. (1985), Brett and Baird (1986), and Rickard (1989) for (left) Middle Devonian stages of Europe and Eastern North America, (middle) the Hamilton Group, and (right) the roughly coeval Traverse Group.

ered the North American asteropyginid species in great detail. In particular, these studies assumed that the ENA taxa represented a largely monophyletic, endemic radiation because they occurred in the same biogeographic region in sediments of the same age. The Asteropyginae of ENA were held to have diversified after the successful invasion of a single, or at most two, lineages, Greenops (Greenops) Delo, 1935, and Greenops

(Neometacanthus) Richter and Richter, 1948. Although the assumption that taxa are closely related on the basis of proximity in space and time and gestalt similarity may not be a bad one, the goal of this analysis was to use a criterion independent of geographic and stratigraphic occurrence to deduce phylogenetic relationship. The criterion used was morphology.

PHYLOGENETIC ANALYSIS

A phylogenetic analysis was conducted on several members of the Asteropyginae using 39 taxa and 66 characters. The characters used in the analysis are given in table 1, and a data matrix showing the codings for all taxa considered is given in table 2. The po-

G R O U P	HAMILTON	MICHICAN BACIN
ů	EQUIVALENT	MICHIGAN BASIN FORMATION
P	EGOIVALENT	FORMATION
1		THUNDER BAY LS.
	TULLY LS.	
T		PETOSKEY/POTTER FARM
l 1		
	MOSCOW	
R		
li		
Α		NORWAY POINT
	LUDLOWVILLE	
V		
		FOUR MILE DAM LS.
		FOOR MILE DAM LS.
E		
R	SKANEATELES	
	SKANEATELES	
		? ? ?
s		DOCK STREET CLAY
<u> </u>		
		GRAVEL POINT/ ALPENA LS.
E		NEWTON CREEK LS.
	MARCELLUS	GENSHAW
		FERRON POINT
ı		ROCKPORT QUARRY LS.
		BELL SHALE

Fig. 2. (Continued)

sition of these characters is shown visually on a trilobite exoskeleton in figure 3. One of the multistate characters, number 9, was treated as additive (ordered), because state 2—anterior margin of frontal globe transverse—appeared to be a special case of a rounded frontal glabellar lobe modified by the ventral deflection of the anterior cephalic border, and thus it represented a clear transformation series. A single most parsimonious cladogram of 489 steps was produced by PAUP 3.0q (Swofford, 1990) using a heuristic search with stepwise addition and 200 random replications (see fig. 4). The retention index for this tree is 0.48, and when uninformative characters are excluded, the consistency index is 0.21. A bootstrap analysis was also conducted to assess the relative support for particular nodes in the cladogram. The analysis used 100 replications with a heuristic search with branch swapping using

nearest neighbor interchanges that saved no more than five trees for each replication. The following nodes appeared in both the cladogram given in figure 4 and the cladogram produced by bootstrap analysis, and the number of times that these nodes were replicated in a 50% majority-rule consensus tree are: Greenops barberi/Greenops chilmanae = 0.54. Bellacartwrightia whitelevi/B. jennyae/B. phyllocaudata/B. calderonae = 0.66, B. whiteleyi/B. calderonae/B. phyllocaudata = 0.45, B. calderonae/B. phyllocaudata = 0.45, Pseudocryphaeus munieri/P. michelini = 0.31, Stummiana arkonensis/S. osismorum = 0.20, Breizhops lanceolatus/B. bigranulosus = 0.21, Kayserops inflatus/ Kayserops traversensis = 0.29, and Philips $mithiana\ burtandmimiae/P.\ hyfinkeli=0.48.$

This phylogeny suggested that there are two major clades within the Asteropyginae, and it can be compared with phylogenies presented by other authors such as Haas (1970), Gandl (1972), Arbizu (1979), Smeenk (1983) and Morzadec (1983) (see fig. 5).

Lesperérance and Desbiens (1995) have criticized the use of continuous variables in phylogenetic analysis, arguing that they represent arbitrarily divided discrete states. Of the characters utilized in this phylogenetic analysis, the following characters satisfy the criteria of continuous variables under Lesperérance and Desbien's (1995) definition: 6, 26, 46, 49, 57, and 60. Lesperérance (pers. comm., 1996) argued that when histograms were generated for the states of these characters, and the number of taxa possessing those states tabulated, they did in some cases approximate a normal distribution. He argued that this is strong evidence that these characters represent continuous variables and should not be included in phylogenetic analysis. The fact that a character and its states may be normally distributed across a clade is interesting, and points to possible clade-level properties of limits of variation in characters. However, these character states were invariant within species. Therefore, they potentially provide resolution of the phylogenetic relationships of these species. Similarly, the number of chambers in the heart varies across the vertebrates and has some particular distribution when the numbers of taxa are mapped by state. However, the fact that all

TABLE 1

Characters Used in Phylogenetic Parsimony Analysis, with Alternative Character States in Parentheses

"0" is the plesiomorphic state; "1"-"6" are apomorphic states. See table 2 for taxon/character matrix

- Interior margin of genal spine: (0) evenly deflected all the way back; (1) more strongly deflected laterally posteriorly.
- S3: (0) faintly sinusoidal, weak posterior deflection medially, weak lateral deflection anteriorly; (1) straight, anterolateral edge transverse or flexing posteriorly; (2) anterior portion convex anteriorly.
- S2: (0) roughly transverse, weakly convex anteriorly;
 (1) weakly declined posteriorly at lateral edge.
- 4. S1 crescent shaped, medial portion developed as: (0) straight line directed anteriorly; (1) smooth curve convex posteriorly.
- Posterior edge of visual surface: (0) anterior to transverse line tangent to anterior edge of L0; (1) posterior to transverse line tangent to anterior edge of L0.
- 6. Maximum number of lenses in a dorsoventral file on visual surface: (0) 8; (1) 13; (2) 4; (3) 10; (4) 7; (5) 6; (6) 9.
- Immediately before cephalic lateral border furrow meets posterior border furrow: (0) posterior border furrow is straight; (1) posterior border furrow flexes anteriorly.
- 8. Facial suture lateral to eye, before flexing posteriorly flexes anteriorly to point: (0) between distal tips of S1 and S2; (1) opposite S2; (2) opposite S1.
- Anterior margin of frontal glabellar lobe in dorsal view: (0) faintly pointed; (1) smoothly rounded; (2) transverse
- Distance between frontal lobe and facial suture at midline relative to laterally: (0) increases slightly; (1) constant.
- Anterior cephalic border medially: (0) does not flex dorsally beyond plane through lateral margins of frontal lobe; (1) flexes strongly dorsally.
- Anteriormost portion of palpebrum reaches: (0) point between medial tips of L3 and PMI; (1) opposite PMI.
- 13. Medially, anterior cephalic border in dorsal view: (0) pointed, lateral margins of border near midline forming a 90° angle; (1) rounded, margins forming approximately a 140° angle; (2) transverse.
- Fixigena: (0) evenly sloping from axial furrow to palpebrum; (1) with distinct bulge medially such that portion parallels dorsal plane.
- Ornamentation on medial part of L0: (0) absent; (1) tubercle; (2) spine.
- 16. Anterior cephalic border medially: (0) not deflected far anteriorly beyond margins of glabella; (1) deflected anteriorly, length (sag.) of at least L1 in front of facial suture.
- 17. Anterior cephalic border: (0) moderate or narrow lip; (1) developed as spatulate process, width (tr.) of process equivalent to width of L3 at its anterior edge; (2) developed as long spine, width (tr.) of spine less than width of medial portion of L1.

- 18. Anterior cephalic border: (0) not developed as spatulate process; (1) with lateral margins of process first diverging anteriorly then converge; (2) with lateral margins of process converging anteriorly.
- Posteromedian impression (PMI) developed on frontal lobe of cephalic exoskeleton as: (0) linear incision; (1) rounded depression; (2) PMI absent.
- Posterior margin of palpebrum relative to anterior margin of palpebrum deflected at an angle to transverse line that is: (0) significantly larger; (1) roughly equivalent.
- 21. L3: (0) flattened dorsally; (1) distinctly swollen.
- 22. Line from posterior to anterior edge of eye forms an angle to sagittal line that is: (0) approximately 30°; (1) approximately 45°; (2) approximately 5-10°.
- Change in cephalic topography where librigena meets lateral border furrow: (0) insignificant; (1) significant.
- 24. Adult prosopon dominated by: (0) fine tubercles; (1) fine granules; (2) coarse tubercles.
- Isolated patches of tubercles on L3: (0) absent; (1) present.
- 26. Genal spine developed as: (0) short triangular projection which does not extend posterior to second thoracic segment; (1) developed as long flange extending back to sixth or seventh thoracic segment; (2) developed as moderately long flange extending back to fourth thoracic segment; (3) developed as moderately long flange extending back to fifth thoracic segment; (4) developed as very long flange extending back to eighth thoracic segment.
- Plane from medial to lateral edges of genal spine: (0)
 evenly inclined; (1) medially flattened ledge, laterally
 evenly inclined.
- Medial edge of genal spine: (0) not significantly thickened relative to lateral edge; (1) significantly thickened relative to lateral edge.
- Lateral margin of genal spine: (0) parallels sagittal line; (1) in medial part forms a line that departs from sagittal line by about 15-20°.
- Lateral margin of genal spine at posterior end: (0) straight; (1) recurves posteriorly.
- 31. The raised ridge on the dorsal surface of the genal spine, when it flexes posteriorly at juncture of posterior border furrow and lateral border furrow: (0) is smoothly curved, convex distally; (1) forms sharp, dog legged kink.
- Axial furrows anterior to S1 relative to furrows posterior to S1, relative to sagittal line: (0) diverge more strongly; (1) diverge at same angle.
- Axial furrows at anterior half of L3 relative to posterior half, relative to sagittal line: (0) equally divergent; (1) less strongly divergent.

TABLE 1 (Continued)

- Portion of librigena between lateral margin of eye and lateral border furrow in dorsal view: (0) visible;
 (1) not visible.
- 35. Eye rests on: (0) sloping librigenal field; (1) vertical wall of librigena.
- 36. Visual surfaces in dorsal view (outgroup is ?): (0) not present within sagittal lines drawn tangent to lateral margins of frontal lobe; (1) present within sagittal lines drawn tangent to lateral margins of frontal lobe.
- Interior margin of genal spine relative to sagittal line:
 (0) deflected laterally 20°;
 (1) parallels lateral margin before deflecting laterally.
- 38. Broad ledge of anterior cephalic border (not considering medial projections) in dorsal view: (0) not visible anterior to anterolateral portions of frontal lobe; (1) visible anterior to anterolateral portions of frontal lobe.
- Ornament on medial part of pygidial and thoracic axial rings: (0) absent; (1) tubercle; (2) spine.
- Shape of terminal pygidial lappet: (0) tear-drop shaped; (1) subrectangular; (2) triangular; (3) very reduced or absent.
- 41. Terminal pygidial lappet, in dorsal view: (0) projects as far or further posteriorly than other pleural lappets; (1) does not project as far posteriorly as pleural lappets that project furthest posteriorly.
- Lateral margins of pygidial lappets: (0) curved; (1) straight.
- Medial margins of pygidial lappets: (0) curved; (1) straight.
- 44. Distal tips of pygidial lappets: (0) pointed; (1) at distal end of straight posterolateral margin that is inclined at 45° angle relative to sagittal line; (2) developed as blunt triangles.
- 45. Anterior margins of pygidial axial rings posterior to constriction of pygidial axis: (0) straight; (1) smoothly convex anteriorly.
- 46. Pygidial axis constricts, i.e., lateral margins of axis are roughly parallel, posterior to: (0) third axial ring; (1) fifth axial ring; (2) sixth axial ring; (3) fourth axial ring; (4) seventh axial ring.
- Pygidial pleural field flanking posterior part of pygidial axis: (0) faintly excavated; (1) prominently excavated.
- Anterior and posterior bands of pygidial pleural segments of: (0) equal elevation; (1) posterior band elevated above anterior band.
- Pygidial lappet that projects furthest posteriorly in dorsal view: (0) fifth; (1) fourth; (2) third.
- Tops of pygidial pleural segments: (0) flat; (1) rounded.
- Pygidial pleural furrows: (0) deeper than interpleural furrows; (1) same depth as interpleural furrows.
- Pygidial pleural furrows: (0) moderately incised, medially short (exsag.); (1) long (exsag.), almost as

- long as anterior pleural band; (2) developed as long (exsag.) excavated space, significantly longer than anterior pleural band.
- In dorsal view at contact of pygidial margin and lappets: (0) lappets of constant breadth; (1) lappets expand laterally.
- 54. At pygidial margins: (0) surface of pygidium is not dorsally inflated; (1) surface of pygidium dorsally inflates at contact of lappets and margins; (2) surface of pygidium dorsally inflates to form fringing ring.
- Laterally pygidial interpleural furrows: (0) arching at same angle as medially; (1) flex more strongly posteriorly; (2) flex anteriorly laterally.
- 56. In medial region of adjacent pygidial pleural segments: (0) posterior band of anterior segment equal in length (exsag.) to anterior band of posterior segment;
 (1) posterior band of anterior segment longer (exsag.) than anterior band of posterior segment.
- 57. Number of pygidial axial rings: (0) 9; (1) 13; (2) 11; (3) 15 or 16; (4) 18; (5) 14; (6) 12.
- 58. Margin of terminal pygidial lappet: (0) not rectangular; (1) subrectangular, convex posteriorly; (2) subrectangular, triangular posteriorly; (3) rectangular, transverse posteriorly.
- 59. Terminal pygidial lappet: (0) not triangular; (1) sharp triangle; (2) blunt triangle.
- 60. Terminal pygidial lappet: (0) broad anteriorly, 1.5 to 2 times width (tr.) of posterior part of pygidial axis; (1) narrow anteriorly, 1-1.1 times width (tr.) of posterior portion of pygidial axis; (2) very broad 3-4 times width (tr.) of posterior part of axis; (3) very narrow, 0.5 times width (tr.) of posterior part of axis.
- 61. Isolated patches of coarse or fine tubercles on L2: (0) absent; (1) present.
- 62. Anterior margins of first five pygidial axial rings going from axial furrows to midline: (0) evenly convex and do not arch strongly anteriorly; (1) straight, then strongly arched anteriorly medially.
- 63. Longest pygidial lappet (excluding terminal lappet): (0) short, length (sag.) from proximal to distal tip equal to length (sag.) of first two pygidial axial rings; (1) long, length (sag.) from proximal to distal tip equal to the length (sag.) of at least the first three pygidial axial rings.
- 64. Number of pairs of pygidial pleural lappets: (0) 5; (1) 4.
- 65. Number of transverse rows of circular fenestrae on anterior band of thoracic pleural segments (outgroup?): (1) 1; (2) 2; (3) none.
- 66. Conformation of circular fenestrae on posterior band of thoracic pleural segments (outgroup ?): (1) a few fenestrae close to axis, do not extend further than half of the pleural field; (2) one transverse row of fenestrae; (3) two transverse rows of fenestrae; (4) fenestrae absent.

TABLE 2

Character States for Taxa Used in Phylogenetic Parsimony Analysis

Characters and alternative states are given in table 1. Missing data are represented by "?"

	1 1234567890	1111111111 1234567890	22222223 1234567890	333333334 1234567890	444444445 1234567890	555555555 1234567890	666666 123456
Pelitlina smeenki	00000000	000000	00000200	000200	00000000	00000000	000022
Treveropyge prorotundifrons	0001020110	001	2000000	0100	0	0000060	000022
rseudocrypnaeus michelini P munieri	0010001		21002010	11001000	1110100	000120	001022
Tolkienia granulispina	2001112210	110000	35550	01220200	02011	11001	001022
Braunops obsoletus	001	8	00002011	000011	110	10000	001013
B. prostellans ,	0101020000	0	00002001	100010	0000030100	0000	001033
Hallandclarkeops cantarmoricus	1111	10	21111101	001010	110	0202114011	1010?3
Neocalmonia quadricosta	011	000	01211101	001010	0000	0000215010	111122
Bellacartwrightia phyllocaudata	1001061011	0001210011	10	12	1010121001	0100113010	001034
B. pleione	25252525	252525255	6. 6	2.5	11011	0100103011	201012
B. Jennyae Pkitolowi	1011131001	0000210001	1010	1100110122	1010021101	0100015011	10101
B. calderonae	1000001011	0001210011	10	117	0100	0100115011	101034
Coltraneia celtica	0011111221	1100000010	10	100	1000120001	0100003022	001022
Bradocryphaeus afghanica	0111140111	0101000010	01	000	1000711121	0100012013	1010??
Heliopyge turolensis	01110?0101	0000100011	11	110	00011	0001012001	1010??
Paracryphaeus hercyniae	02111?0000	000200022	01	10	00213	0000110303	1000??
Pilletina aequisulcata	0001041121	2000120001	01	0010010001	0002010001	1000003202	0010??
Breizhops lanceolatus	0111051201	0000110000	000	0011011101	0111010000	0000012201	000012
B. chaconae	0111040111	0101000001	00	0010011001	0012010001	0000112201	220000
B. bigranulosus	0111020210	0000100010	000	1101101001	0011010101	0001012201	1000??
Philipsmithiana burtandmimiae	0101041100	0000010011	00	0010111102	1010010001	0100003100	101034
F. nytnkeli Modellong etmosi	0201061011	00001000000	9 8	0010101001	1111010101	0100103100	100034
Momentops struvel	0101001111	0000100001	2 6	0010001001	1000110001	0200002100	010055
neometacaninus sp. att. siettijer Asterovvge punctata	0101140111	0010010010	\sim	0010011103	1110110011	0001113020	011012
Rhenops babini	2001100211	000000000	00	2010011002	1002111000	0000103020	0110??
Rhenops, new species	0001141010	0100200001	110	1010111112	11111110?0	0000102030	201022
Greenops chilmanae	011115101?	0010100011	0000131000	0110110002	1010110000	0000102011	111011
G. grabaui	0101150011	00000	00003100	0010101001	1012110000		010011
G. widderensis	0101150011	0001	20013100	0010101001	1010110000		101011
G. barberi	0110050010	0001	20003000	0010111002	1010110000	0000002011	111011
Deloops haasi	0211141211	0002	0000021001	00000	0012120000	0000105200	0100?3
Stummana osismorum	0111141010	0000	00002000	11100100	1012110000	0000002101	111017
3. arkonensis	0111041110	T000	2222T000	11010701	1002110101		11011
Armorigreenops leoi Kennacrvphaeus harrisae	1111101210	0110200011	0000010101	0110001012	1010010001	0000102021	7 0
Kayserops inflatus	0201100210	0002	111010	01111101	1100111101		102
K. traversensis	1114101	0002	0002120102	0111101012	10101101?1	000001?021	101012

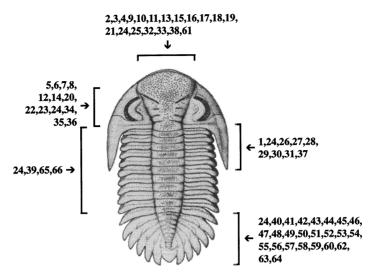


Fig. 3. An illustration of a hypothetical species of asteropyginine trilobite modified from Ludvigsen (1979) showing the approximate position of characters used in the phylogenetic analysis.

humans and chimps possess a four-chambered heart whereas all sharks do not is a valid synapomorphy providing evidence that chimps and humans share a sister group relationship to the exclusion of sharks. Thus, although Lesperérance and Desbiens (1995) made an important point—that variation

within characters should be quantified—as long as character states are invariant within terminal taxa they still can and should be used to provide evidence for common descent. Vrba et al. (1994) also demonstrated how truly continuous variables can be used to determine phylogenetic relationships.

SYSTEMATIC PALEONTOLOGY

SUPERFAMILY ACASTACEA Delo, 1935

FAMILY ACASTIDAE Delo, 1935

SUBFAMILY ASTEROPYGINAE Delo, 1935

PELITLINA HAAS, 1968

Type Species: Pelitlina goltzi.

Assigned Taxa: Pelitlina smeenki, new species.

DIAGNOSIS: Cephalic border furrow bisects genal spine; S3 faintly sinusoidal; posterior edge of visual surface anterior of transverse line tangent to L0; distance between frontal lobe and facial suture at midline relative to lateral edges increases slightly; lateral margin of genal spine at posterior end straight; five pygidial pleural lappets; pygidial margin constricts posterior of third axial ring; axial

furrows at anterior half of L3 relative to posterior half equally divergent; nine pygidial axial rings; posterior pygidial terminus teardrop shaped.

DISCUSSION: The two species in the genus are united by at most the unique possession of only two characters: the nine pygidial axial rings and the teardrop-shaped pygidial terminus. The other characters are diagnostic in combination. In addition, the first of these two characters is developed in several closely related species such as Acastella tiro Richter and Richter, 1954, and it is also found in the more distantly related Paracryphaeus hercyniae Gandl, 1972. Thus, this genus is difficult to diagnose, and is more noteworthy for the characters it lacks rather than those it uniquely possesses. The genus in many ways fits the description of a metataxon propounded by Gauthier et al. (1988), and it is certainly due to the lack of autapomorphies that

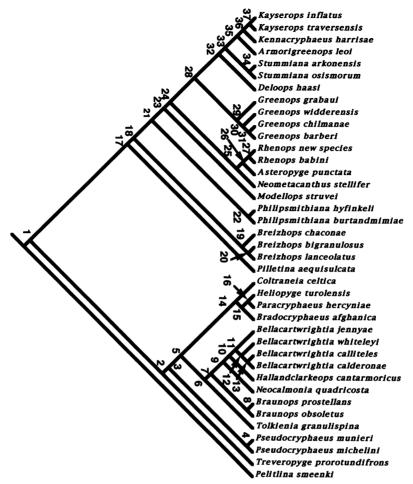


Fig. 4. The single most parsimonious tree of length 474 steps produced using a heuristic search with stepwise addition and 100 random replications by Paup 3.0q (Swofford, 1990). The retention index is 0.44, and when uninformative characters are excluded, the consistency index is 0.22. Character states at nodes optimized using ACCTRAN are as follows, with characters numbered as in table 1 and character states in parentheses: Node 1: 4(1), 8(1), 9(1), 11(0), 17(0), 18(0), 20(1), 21(0), 26(2), 29(1), 33(1), 36(1), 46(1), 51(0), 53(0), 54(0), 57(2), 64(0), 65(1), 66(2); **Node 2**: 19(1), 22(2), 31(1), 41(1), 66(3); Node 3: 23(1), 52(1), 60(1), 63(1); Node 4: 8(0), 29(0), 40(1), 58(2); Node 5: 5(1), 6(1), 48(1), 50(1), 56(1); Node 6: 19(0), 36(0); Node 7: 6(3), 16(1), 22(0), 38(1), 40(2), 59(2); Node 8: 2(1), 5(0), 8(0), 23(0), 41(0); Node 9: 25(1), 26(1), 27(1), 36(1), 57(5), 61(1); Node 10: 7(1), 14(1), 22(2), 43(1), 46(2); Node 11: 1(1), 3(1), 8(0), 10(1), 15(2), 27(0), 28(1), 29(0), 35(1), 39(2), 66(3); Node 12: 19(1), 34(1), 65(2); Node 13: 3(0), 5(0), 6(0), 55(1), 65(3), 66(4); Node 14: 3(1), 10(1), 12(1), 20(0), 26(0), 45(1), 60(2); Node 15: 2(1), 6(4), 24(2), 25(1), 49(2), 60(3), 61(1); Node 16: 9(0), 12(0), 20(1), 26(1), 38(1), 52(0); Node 17: 6(4), 7(1), 10(1), 40(1), 44(1), 50(1), 58(2); Node 18: 2(1), 26(1), 27(1), 29(0), 37(1), 43(1); Node 19: 3(1), 56(1), 60(1); Node 20: 6(2), 8(2), 15(1), 19(0), 27(0), 34(1); Node 21: 36(0), 41(1), 58(1); Node 22: 25(1), 30(1), 35(1), 52(1), 57(3), 61(1), 65(3), 66(4); Node 23: 45(1), 62(1); Node 24: 5(1), 19(1), 44(0), 55(1); Node 25: 22(2), 27(0), 28(1), 36(1), 63(1); Node 26: 38(1), 40(2), 42(1), 57(3), 58(0), 59(2); **Node 27**: 2(0), 8(0), 19(0), 27(1), 31(1), 44(1), 47(1), 50(0); **Node 28**: 8(0), 26(2), 44(2), 50(0), 66(1); **Node 29**: 6(5), 7(0), 13(1), 26(3), 35(1); **Node 30**: 22(2), 25(1), 44(0), 61(1), 63(1); Node 31: 3(1), 10(0), 36(1), 40(2), 58(0), 59(1), 60(1); Node 32: 3(1), 14(1), 15(1), 30(1), 44(2), 66(3); Node 33: 10(0), 27(0), 32(1), 39(1), 60(1); Node 34: 12(1), 13(1), 66(1); Node 35: 6(0), 14(0), 26(1), 28(1), 40(2), 58(0), 59(2); Node 36: 30(0), 35(1), 36(1), 44(2), 50(1), 57(1); Node 37: 2(2), 19(2), 24(1), 25(1), 34(1), 48(1), 62(0), 63(1).

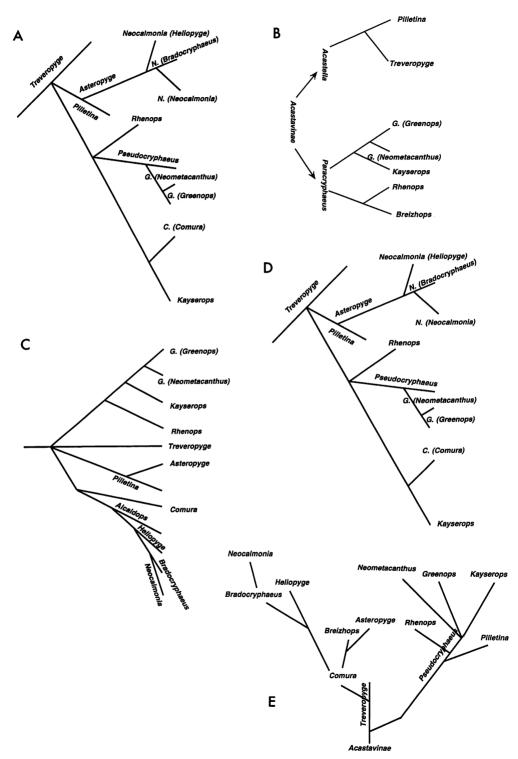


Fig. 5. Five scenarios for asteropyginid evolution hypothesized by (A) Haas (1970), (B) Gandl (1972), (C) Arbizu (1979), (D) Smeenk (1983), and (E) Morzadec (1983).

Pelitlina, Acastava, and other acastavinids have been traditionally treated as ancestral taxa by evolutionary taxonomists. The possibility exists that this genus is paraphyletic, but the two unique characters (or one) and the other characters given in combination do suggest a means of uniquely diagnosing it as a monophyletic taxon. More detailed work on other acastavinids is needed to rigorously define genera, but this work will not be pursued herein because Pelitlina is simply employed as an outgroup. Ingroup Asteropyginae can be separated from this and other outgroups by the following characters (state of each character given in parentheses): 4(1), 8(1), 9(1), 20(1), 26(2), 29(1), 33(1), 36(1), 46(1), 57(2), 65(1), and 66(2); however, all of these characters do show reversal within the Asteropyginae.

The two species assigned to this genus are very similar: differing principally in the condition of the prosopon, which is dominated by fine granules in *Pelitlina smeenki* but is dominated by fine tubercles in *P. goltzi* Haas, 1968; the condition of S2, which extends further laterally in *P. goltzi*; the lateral margins of the cephalon, which deflect more sharply posteriorly in *P. goltzi*; and the posterior margins of the pygidial lappets, which are pointed in *P. smeenki* and weakly rounded in *P. goltzi*. *Pelitlina goltzi* is known from the Upper Emsian of northwestern Turkey, and *P. smeenki* is known from the Lower Emsian of northern Spain.

Pelitlina smeenki, new species

Acastava n. sp. Smeenk, 1983: pl. 28, figs. 1-6.

TYPES: Holotype Rijksmuseum van Geologie en Mineralogie, Leiden, the Netherlands, RGM 339515, Smeenk (1983: pl. 28, fig. 1).

DIAGNOSIS: Same as for genus and prosopon of fine granules; posterior margins of pygidial lappets pointed.

ETYMOLOGY: Named for Z. Smeenk, who figured this species in his excellent monograph on Devonian trilobites of northern Spain.

DESCRIPTION: Cephalic length (sag.) 50% of width. Axial furrow nearly straight anterior to S1, diverging forward at about 25°, narrow, shallow; posterior to S1 diverging at

10°. Cranidial anterior border short exsagittally and medially, medially weakly upturned triangular process. Preglabellar furrow shallow and narrow. Cranidial anterior border lengthens slightly medially, developed as narrow lip. Facial suture anterior of eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length equal to width across frontal lobe; frontal lobe ellipsoid, about 60% of length of glabella; L1-L3 flat (sag.); frontal lobe at level of posterior glabellar region; anterior part of frontal lobe moderately declined forward, faintly pointed in dorsal view. Posteromedian cephalic impression (PMI) shallow linear incision. S3 shallow, weakly sinusoidal, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening distally, outer edge nearly straight along axial furrow. S2 roughly transverse, weakly convex anteriorly, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial portion developed as straight line directed anteriorly, contacting axial furrow laterally. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly medially, deep. L0 65% of width of frontal lobe, moderately arched (tr.), longest sagittally. Large Eye Index 0.55. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite S1, anterior to transverse line tangent to L0; eye raised slightly above frontal lobe; maximum number of eight lenses in a dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to internal margin of eye. Postocular fixigenal field 30% of length of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, weakly flexing posteriorly distally, anterior margin nearly straight, of relatively equal length (exsag.), longest posterior to palpebrum, shallowing distally. Genal spine developed as short triangular projection, cephalic lateral border furrow bisects spine; lateral and medial edges of genal spine evenly inclined; lateral margin of genal

spine parallels sagittal line; lateral margin of genal spine at posterior end straight; interior margin of genal spine deflected laterally. Lateral border furrow narrow, shallow; lateral border weakly widens posteriorly. Anterior margin of cephalic doublure bluntly pointed. Prosopon of fine granules.

Thorax not known.

Pygidium broadly triangular in outline excluding marginal lappets, length about 75% of width without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, diverging at 20° angle anterior to third pygidial axial ring; roughly parallel posterior to third pygidial axial ring. Axis 30% of pygidial width anteriorly, with nine rings; rings of nearly equal length distally and sagittally, longest sagittally; anterior and posterior edge of axial rings posterior to third axial ring from midline to distal edge first straight, then flexing posteriorly, then anteriorly, then slightly posteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved; medial margins curving; distal tips pointed. Axial terminus teardrop shaped. projecting further posteriorly than other lappets; postaxial region 20% of pygidial length (sag.). Pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; anterior and posterior pleural segments of equal elevation; tops of segments flat; pleural furrows deeper than interpleural furrows, moderately incised, short (exsag.). Laterally pygidial interpleural furrows arch at same angle as medially. In medial region of adjacent pleural segments, posterior band of anterior segment equal in length (sag.) to anterior band of posterior segment.

DISCUSSION: This species was used as the outgroup in the phylogenetic analysis and is known from the Lower Emsian of northern Spain (Smeenk, 1983). Smeenk (1983) merely figured the specimen and did not discuss or describe it. Thus, a description is given here.

TREVEROPYGE STRUVE, 1958b

TYPE SPECIES: Asteropyge (Asteropyge) prorotundifrons Richter and Richter, 1943.
ASSIGNED TAXA: None (see below).
DIAGNOSIS: See Struve (1959) and T. prorotundifrons below.

DISCUSSION: Several species appear to share the combination of characters that the type of the genus, Treveropyge prorotundifrons, possesses, and they may belong to this genus. These species include Asteropyge wallacei Termier and Termier, 1950, Treveropyge wallacei procerospinosa Gandl, 1972, Echinopyge cathamma Haas, 1968, Cryphaeus boopis Richter, 1909, Cryphaeus drevermanni Richter, 1909, and possibly ?Treveropyge henryi Arbizu, 1979, and ?Phacops rotundifrons Emmrich, 1839. However, until material of these taxa is examined in detail, such conclusions must await further testing.

Both Haas (1970) and Morzadec (1983) concluded that Treveropyge was one of the basal members of the asteropyginid clade (see fig. 5a, e). This is not entirely supported by the analysis conducted herein, as Treveropyge is itself sister to one of the two main clades of asteropyginines. Haas (1970) viewed Treveropyge as part of a paraphylum sister to the rest of the asteropyginines, which in turn consisted of branches off of a basal split between the genus Pilletina Haas, 1970 and its descendants, which included Neocalmonia Pillet, 1969, Heliopyge Haas and Mensink, 1969, Bradocryphaeus Haas and Mensink, 1969, and Asteropyge Hawle and Corda, 1847, and the genus Pseudocryphaeus Pillet, 1954, and its descendants, which included Rhenops Richter and Richter, 1943 Neometacanthus Richter and Richter, 1948, Greenops Delo, 1935, Comura Richter and Richter, 1926, and Kayserops Delo, 1935. Morzadec (1983) viewed asteropyginid evolution as involving two branches that split early in the Devonian. He felt these two basal branches consisted of a paraphyletic Treveropyge, which gave rise to several other taxa including Bradocryphaeus, Neocalmonia, Heliopyge, and Asteropyge, and a paraphyletic Pseudocryphaeus, which gave rise to Rhenops, Neometacanthus, Kayserops, and Greenops. These conclusions about a dichotomous basal branch in the Asteropyginae are supported, but instead Treveropyge and Pseudocryphaeus are part of only one of the major branches. However, the branch containing Treveropyge also contains Bradocryphaeus and Neocalmonia. Smeenk (1983) concluded (see fig. 5d) that Treveropyge was

part of a basal polytomy in the Asteropyginae along with Pilletina and a lineage that evolved into Pseudocryphaeus and Paracryphaeus. Although greater resolution than this classic "star phylogeny" sensu Edgecombe (1992) was possible in this analysis, the relatively close relationship between Treveropyge and Pseudocryphaeus was replicated here. Gandl (1972) (see fig. 5b) suggested that portions of Treveropyge were sister to certain species of Metacanthina Pillet, 1954. Here one of the species he assigned to Metacanthina, M. munieri (Oehlert), is treated as closely related to Treveropyge and is assigned to Pseudocryphaeus. Partially matching Gandl's (1972) conclusion, Treveropyge appears to be sister to *Pseudocryphaeus*, and it is discussed in greater detail below.

Treveropyge prorotundifrons (Richter and Richter, 1943)

Asteropyge (Asteropyge) prorotundifrons Richter and Richter, 1943: 175, pl. 7, figs. 1-3.

Treveropyge prorotundifrons (Richter and Richter). Struve, 1959: O483, figs. 382, 1a, b.

Treveropyge prorotundifrons iberica Gandl, 1972: 121, pl. 10, figs. 6-10.

Treveropyge prorotundifrons prorotundifrons Richter and Richter. Gandl, 1972: 120, text figs. 16–18.

Treveropyge iberica Gandl. Smeenk, 1983: 400, pl. 6, figs. 7–9, pl. 7, figs. 1, 2.

?Treveropyge prorotundifrons iberica Gandl, 1972: 121, pl. 9, figs. 8-10.

TYPES: Holotype defined in Richter and Richter (1943: pl. 7, fig. 3).

DIAGNOSIS: Medial portion of S1 smooth curve directed anteriorly; lateral to eye facial suture, before flexing posteriorly, flexes anteriorly to point opposite S2; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; cephalic posterior border furrow longest (exsag.) posterior to palpebrum; demarcation between librigena lateral to eye and lateral border furrow weak; cephalic axial furrows diverge anteriorly at constant angle; genal spine developed as short triangular spine that does not extend posterior to second thoracic segment; 12 pygidial axial rings; anterior and posterior pleural segments of equal elevation when viewed laterally; terminal pygidial lappet teardrop shaped; posteriormost pair of pygidial pleural lappets project further posteriorly than terminal pygidial lappet; lateral and medial margins of pygidial lappets curved.

DISCUSSION: This species, the type of the genus, is known from the lower Emsian of northern Spain and the Eifelian of Germany, and was used to represent the phylogenetic position of *Treveropyge* in the analysis. On the basis of the absence of the derived state for several characters, it has a basal position in one of the major clades of the Asteropyginae.

The subspecies of *T. prorotundifrons* recognized by Gandl (1972), *prorotundifrons* and *iberica*, do not appear to be distinguishable on the basis of any morphological features, and are treated as synonymous here. Any differences that may exist between specimens hailing from the two different distributional areas—the former, *T. prorotundifrons prorotundifrons*, in the Eifelian of Germany, and the latter in the Lower Emsian of northeastern Spain—are so slight that, although possibly suggesting allopatric or temporal differentiation, they would not affect the coding of character states used in phylogenetic analysis.

It could not be ascertained whether the specimens shown by Gandl (1972: pl. 9, figs. 8–10) and called meraspids of *Treveropyge* prorotundifrons iberica are correctly identified. In these specimens, the pygidial terminus extends posterior to the fifth pair of pygidial lappets, whereas in T. prorotundifrons the lappets extend significantly further than the pygidial terminus. In addition, the pygidial terminus is teardrop shaped in typical T. prorotundifrons, whereas it is triangular in the specimens in Gandl's (1972) plate 9. Moreover, these specimens have significantly fewer pygidial axial rings as well as much blunter pygidial lappets. Finally, the lateral margins of the pygidial axial rings constrict posterior to the third pygidial axial ring such that the axial furrows are parallel posterior to this point in the putative meraspids, whereas in holaspid T. prorotundifrons the pygidial axis sharply constricts posterior to the fifth axial ring.

If these pygidia are actually meraspids (they appear to all be M9) of *T. prorotun-difrons*, then we can compare morphoclines in the transformation of character states dur-

ing ontogenetic development to morphoclines suggested by outgroup analysis. According to outgroup analysis, primitively the pygidial axis is held to constrict, i.e., the lateral margins of the axis are roughly parallel, posterior to the third axial ring (character 46). One of the derived states of this character is to have the constriction posterior to the fifth axial ring. In this case, ontogenetic and outgroup polarizations match. According to outgroup analysis, primitively the pygidial terminus projects further posteriorly than the other pygidial lappets (character 41). This transformation series is also supported by the possible ontogenetic transformation series provided by T. prorotundifrons. Of course, the number of pygidial axial rings (character 57) is lower during earlier stages of ontogeny, and it is also lowest in the outgroup taxon employed. However, the transformation of at least one character does not agree with the polarities suggested by outgroup analysis. Primitively, the shape of the terminal pygidial lappet (character 40) is held to be teardrop shaped. In T. prorotundifrons, it transforms from triangular to teardrop shaped in the transition from putative meraspid to holaspid.

Treveropyge prorotundifrons shares many similarities with T. wallacei procerospinosa Gandl, 1972, including the teardrop-shaped terminal pygidial spine, the number of pygidial axial rings, and the condition of the pygidial lappets. They differ chiefly in the length of the terminal pygidial spine, which projects further posteriorly than the pygidial lappets in T. wallacei procerospinosa but not in T. prorotundifrons.

PSEUDOCRYPHAEUS PILLET, 1954

Type Species: *Phacops michelini* Rouault, 1851.

Assigned Taxa: Cryphaeus munieri Oehlert, 1877.

DIAGNOSIS: Cephalic posterior border furrow longest (sag.) posterior to medial visual surface of eyes; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; PMI developed as rounded depression; posterior and anterior margins of fixigena deflected such that they intersect transverse line at roughly equivalent angle; line from posterior to anterior edge of eye intersects sag-

ittal line forming 5° angle; demarcation between librigena lateral to eve and lateral border furrow prominent; genal spine developed as moderately long flange extending back to fourth thoracic segment; raised ridge on dorsal surface of genal spine flexes posteriorly at juncture of posterior border furrow and cephalic border furrow and forms sharp, doglegged kink; cephalic axial furrows diverge at constant angle; axial furrows opposite anterior half of L3 relative to posterior half less strongly divergent; terminal pygidial lappet triangular; pygidial axis constricts posterior to fifth ring; pygidial pleural furrows almost as long (exsag.) as anterior pleural band; lateral margins of terminal pygidial lappet anteriorly parallel, posteriorly convergent and triangular.

DISCUSSION: Three species traditionally assigned to Pseudocryphaeus were subjected to phylogenetic analysis. These were the type of the genus, P. munieri (Oehlert, 1877), and Braunops prostellans (Richter and Richter, 1939). Braunops prostellans is separated by several nodes on the cladogram from the type of the genus, and differs from Pseudocryphaeus michelini in the condition of the following characters: 2, 9, 14, 16, 19, 22, 23, 28, 34, 36, 38, 40, 42–44, 46, 48, 52, 56, 58, 59, 62, and 63. Therefore, this species must be assigned to a different genus, and it is discussed in greater detail below. Pseudocryphaeus munieri differs from P. michelini in the condition of several characters, including: 4, 6–8, 14, 28, 34, 42–44, 57, 60, 62, and 63.

According to the analysis conducted herein, the genus appears to have a basal phylogenetic position in one of the major clades within the Asteropyginae. Treveropyge is sister to this clade, and this accords well with its position in the evolutionary tree of Haas (1970) (see fig. 5a). However, Haas (1970) thought that all genera in the Asteropyginae other than Treveropyge were derived from either Pseudocryphaeus or Pilletina. These genera he placed as a near basal bifurcation in the evolution of the asteropyginines. In his evolutionary tree, *Pseudocryphaeus* is wildly paraphyletic, giving rise directly to at least four genera: Rhenops, Greenops, Neometacanthus, and Comura. A close relationship between Pseudocryphaeus and Greenops, Neometacanthus, and Rhenops is not supported here. As redefined here, Pseudocryphaeus is para- or polyphyletic, but inclusion of some taxa within the genus that had formerly been assigned to it such as Braunops prostellans would make Pseudocryphaeus paraphyletic. In addition, Haas' (1970) hypothesis that Pseudocryphaeus and several other asteropyginid genera should be sister to a lineage containing among other taxa Neocalmonia, Heliopyge, Bradocryphaeus, and Asteropyge is not supported. As discussed above, Morzadec (1983) (see fig. 5e) thought that *Pseudocryphaeus* was the most primitive taxon in one of the basal branches of the Asteropyginae, and he also viewed it as paraphyletic and suggested that it gave rise to same the genera Haas (1970) suggested as well as to the genus Pilletina. The basal position of Pseudocryphaeus in one of the maior branches within the Asteropyginae is supported, but the overall topology of asteropyginid evolution Morzadec (1983) advocated is not. Pilletina is discussed further below. Arbizu (1979) suggested (see fig. 5c) that Pseudocryphaeus was paraphyletic and gave rise to Kayserops, Rhenops, and Greenops, while also being part of a basal polytomy along with Treveropyge, Pilletina, and Comura. Again, the relatively basal position of Pseudocryphaeus is supported herein; however, it does not appear to belong to a basal polytomy, only a basal bifurcation. Gandl (1972) suggested (see fig. 5b) that Pseudocryphaeus and Paracryphaeus were sister taxa, with both of these taxa related to a clade derived from a different segment of the "Acastavinae" than Treveropyge was. As mentioned above, Gandl's (1972) diphyletic origin of the Asteropyginae is not supported by the analysis conducted herein, as one genus within the "Acastavinae," Pelitlina, shares the most synapomorphous character states with the asteropyginid clade. Until another acastavine is found to share a greater or equal number of synapomorphies with the asteropyginines, and also to contain characters uniting it with Pseudocryphaeus and other asteropyginines to the exclusion of Treveropyge, Gandl's (1972) entire perspective on asteropyginine phylogeny must be called into question.

Without access to specimens and addition-

al, detailed phylogenetic analysis, the generic status of species traditionally assigned to *Pseudocryphaeus*, e.g., *P. jonesi* (Oehlert, 1877), *P. vernensis* Morzadec, 1971, *P. oehlerti* Morzadec, 1971, *P. baconnierensis* Morzadec, 1971, and *P. spatulaeformis* Morzadec, 1971, cannot be evaluated at this time.

Pseudocryphaeus michelini (Rouault, 1851)

Pseudocryphaeus michelini (Rouault). Struve, 1959: O482, figs. 381, 3a, b; Morzadec, 1971: 169, pl. 15, figs. 1-20, text fig. 7, see for extended synonymy; Gandl, 1972: 91.

Pseudocryphaeus cf. michelini (Rouault). Smeenk, 1983: pl. 2, figs. 7, 8.

Types: Lectotype defined in Morzadec (1971: pl. 15, fig. 20).

DIAGNOSIS: S2 weakly declined posteriorly at lateral edge; medial portion of S1 developed as smooth curve convex posteriorly; maximum of eight lenses in dorsoventral file on visual surface; medial edge of genal spine significantly thickened relative to lateral edge; librigenae between lateral margins of eyes and lateral border furrow not visible in dorsal view; terminal pygidial lappet subrectangular, triangular at distal end, projecting further posteriorly than pygidial pleural lappets; lateral and medial margins of pygidial lappets straight; nine pygidial axial rings.

DISCUSSION: This species is known from the Athyris undata limestones, Siegenian (?Pragian), of the Massif Armorican. The illustration in the treatise (Struve, 1959) is inaccurate in its depiction of the terminal pygidial lappet, which should be rectangular anteriorly, triangular posteriorly.

Pseudocryphaeus munieri (Oehlert, 1877)

Cryphaeus munieri Oehlert, 1877: 582, pl. 9, fig. 3, not fig. 3a.

Pseudocryphaeus munieri (Oehlert). Haas, 1970: 119, pl. 4, figs. 8, 9; Morzadec, 1971: 171, pl. 16, figs. 1–13, text fig. 13 (see for extended synonymy).

TYPES: Lectotype defined in Morzadec (1971: pl. 16, fig. 5).

DIAGNOSIS: Maximum of 10 lenses per dorsoventral file on visual surface; immediately before cephalic lateral border furrow meets posterior border furrow, posterior border furrow flexes anteriorly; lateral to eye, facial suture before flexing posteriorly flexes anteriorly to point opposite S1; demarcation between librigena lateral to eye and lateral border furrow prominent; medial edge of genal spine not significantly thickened relative to lateral edge; medial portion of S1 developed as straight line directed anteriorly; lateral and medial margins of pygidial lappets curved; terminal pygidial lappet triangular posteriorly; last pair of pygidial lappets projects further posteriorly than pygidial terminal lappet.

DISCUSSION: This species is known from the Athyris undata limestones, Siegenian (?Pragian), of the Armorican Massif. Gandl (1972) figured a pygidium (pl. 7, fig. 1) that he referred to as Metacanthina munieri and viewed as synonymous with Pseudocryphaeus munieri. However, Gandl's specimen differs from P. munieri in the condition of several characters. In particular, the pygidial terminus extends further posteriorly than the pygidial pleural lappets in Gandl's specimen, whereas in P. munieri the lappets extend further posteriorly. In addition, the sides of the pygidial terminus at its anterior end are parallel in P. munieri, whereas they are convergent in Gandl's specimen. Finally, the lateral edges of the pleural lappets in P. munieri show greater curvature than they do in Gandl's figured specimen. Therefore, Gandl's specimen clearly represents a different, and possibly new, species. Because of the limited morphological information, it is difficult to ascertain whether this pygidium belongs to Pseudocryphaeus or to some other genus.

TOLKIENIA. NEW GENUS

Type Species: Comura (Comura) granulispina Morzadec, 1983.

ASSIGNED TAXA: Comura (Comura) cancer Morzadec, 1983, Comura nova Arbizu, 1979, and Greenops sp. A Stumm, 1953.

DIAGNOSIS: Anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; pygidial pleural lappets bulge laterally in dorsal view at pygidial margin; pleural lappets very long; terminal pygidial lappet teardrop shaped, does not extend as far posteriorly as other pleural lappets; prosopon of fine

tubercles; lateral and medial margins of pygidial pleural lappets curved; distal tips of pygidial lappets pointed; pygidial axis constricts posterior of sixth axial ring; pygidial interpleural furrows flexing at same angle laterally and medially; tops of pygidial pleural segments rounded; pleural furrows long (exsag.), almost as long as anterior pleural band; in medial region of adjacent pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; fourth pygidial lappet projects furthest posteriorly; anterior margins of pygidial axial rings posterior to sixth axial ring straight; terminal pygidial lappet narrow; anterior cephalic border not deflected strongly anteriorly.

ETYMOLOGY: Named for the author J. R. R. Tolkien.

DISCUSSION: This genus is known from the lower Couvinian (?Eifelian) of the Cantabrian Mountains of Spain, from the upper Emsian of the Armorican Massif, and possibly from the Givetian of the Michigan Basin of ENA. There are taxa that authors in the past have assigned to Comura Richter and Richter, 1926, but that are removed from that genus herein because they do not possess all of the diagnostic characters of Comura. Some of these forms are assigned to *Tolkienia*. For instance, contra the diagnosis of Comura, in C. granulispina Morzadec, 1983, C. kervezensis Morzadec, 1983, and C. cancer Morzadec, 1983, medially the pygidial axial rings do not flex strongly posteriorly. In addition, they do not possess a prominent medial tubercle or spine on the pygidial axial rings, which is diagnostic of Comura. Comura nova Arbizu, 1979, also differs in characters diagnostic of Comura. It is possible that if subjected to phylogenetic analysis, the aforementioned species might map out as very closely related to the type of Comura, C. cometa, and thus they could of course be assigned to Comura if an emended diagnosis were presented. However, neither the type of Comura nor other adequately preserved species matching the diagnosis of that genus could be obtained for the purposes of phylogenetic analysis.

Alcaldops is very similar to the type species of Comura. The chief difference between the two genera appears to be that al-

though species of Alcaldops do have a single medial spine or tubercle present on the pygidial axial rings, these tubercles, as well as the medial portion of the pygidial axial rings. do not arch strongly posteriorly. In addition, the type species of Alcaldops, A. alcaldei Arbizu, 1979, does not possess a prominent spine on L0, unlike Comura tuberculata Morzadec, 1969. Until studies can be carried out on species of Alcaldops, particularly the type of the genus, it is treated as still valid, although it was not analyzed herein. However, the possibility exists that the genus may contain species both nested within and outside of species traditionally assigned to Comura, and therefore it would be either no longer valid or in need of substantial revision.

Although Comura could not be subjected to phylogenetic analysis herein, it is probable that it is closely related to Tolkienia. In particular, both genera have: very long pygidial pleural lappets that are rounded; the lateral and medial margins of the pleural lappets curved; the posterior portion of the pleural segments elevated above the anterior portion; the teardrop-shaped, narrow terminal pygidial lappet; and, in the medial region of adjacent pleural segments, the posterior region of the anterior segment is longer (sag.) than the anterior portion of the posterior segment. Arbizu (1979), Smeenk (1983), and Morzadec (1983) suggested that there was a close relationship between Neocalmonia Pillet, 1969, Heliopyge Haas and Mensink, 1969, Bradocryphaeus Haas and Mensink, 1969, and Comura Richter and Richter, 1926. This claim could not be verified herein. However, Tolkienia maps out on the cladogram in figure 4 within a larger clade including these genera, although it is not sister to any of them.

There exists a single pygidium from the Potter Farm Formation (Taghanic), upper Givetian, of the Michigan Basin in ENA that may be referable to *Tolkienia*. This specimen, referred to as *Greenops* sp. A by Stumm (1953), lacks nearly all of the diagnostic characteristics of the pygidium of *Greenops* and cannot be assigned to that genus henceforth. Although poorly preserved, it appears to possess some of the diagnostic characteristics of the genus *Tolkienia*, and it

is tentatively placed in that genus awaiting the recovery of additional specimens. In particular, it has the very long pygidial pleural lappets with curved lateral and medial margins characteristic of *Tolkienia*. In addition, as in *Tolkienia*, the terminal pygidial lappet is very narrow; laterally the pygidial interpleural furrows arch at the same angle as medially; the pleural field flanking the terminal portion of the pygidial axis is faintly excavated; it may have a triangular terminal pygidial lappet; and the pygidial pleural lappets expand slightly where they reach the pleural field.

Tolkienia granulispina Morzadec, 1983, was used in the phylogenetic analysis because it was the most completely preserved taxon available of this new genus.

Tolkienia granulispina (Morzadec, 1983)

Comura (Comura) granulispina Morzadec, 1983: 134, pl. 21, figs. 1-7.

TYPES: Holotype defined in Morzadec (1983: pl. 21, fig. 2).

DIAGNOSIS: Distance between frontal glabellar lobe and facial suture increases slightly from anterolateral margins to midline: S3 faintly sinusoidal; S2 roughly transverse, convex anteriorly; maximum of 13 lenses per dorsoventral file on visual surface: tubercle on L0 present; PMI developed as linear incision; posterior margin of fixigena relative to anterior margin deflected at an angle to transverse line that is roughly equivalent; medial tubercles or spines on pygidial and thoracic axial rings present; medial margins of pygidial pleural lappets curving laterally; anterior margins of pygidial axial rings posterior to sixth axial ring straight; pygidial pleural fields flanking posterior part of pygidial axis faintly excavated; 11 pygidial axial rings.

DISCUSSION: This species is known from the upper Emsian of the Armorican Massif. The PMI was treated as being developed as a linear incision in this species instead of as a rounded depression. This is because the PMI is rectangular, although not significantly so, being almost square; however, this is treated as showing greater concordance with the linear character state than with the rounded character state.

Morzadec (1983: fig. 33) held that this species was directly ancestral to *Tolkienia cancer* (Morzadec, 1983) and that this ancestral lineage gave rise to several species of *Comura* as well as to species of *Asteropyge* Hawle and Corda, 1847, such as *A. punctata* Steininger, 1831. *Tolkienia cancer* and *T. granulispina* appear to be closely related, and the relationship between *Tolkienia* and *Comura* could not be evaluated herein, although it probably is a close one. However, *Asteropyge punctata* appears to be only distantly related to *T. granulispina*, and the two taxa are separated by several nodes on the cladogram given in figure 4.

BRAUNOPS. NEW GENUS

Type Species: Kayserops obsoletus Gandl, 1972.

ASSIGNED TAXA: Kayserops brevispinosus Gandl, 1972, Pseudocryphaeus ogivalis Morzadec, 1976, Asteropyge (Metacanthus) prostellans Richter and Richter, 1939, and possibly Pseudocryphaeus asteriferus Haas, 1968.

DIAGNOSIS: S3 straight, anterolateral edge flexes posteriorly; posterior edge of visual surface anterior to transverse line tangent to L0; fixigena evenly sloping from axial furrow to palpebrum; facial suture lateral of eye, before flexing posteriorly flexes anteriorly to point between distal tips of S1 and S2; distance between frontal lobe and facial suture at midline relative to laterally increases slightly; medially anterior cephalic border pointed in dorsal view; posterior margin of palpebrum relative to anterior margin deflected at an equivalent angle to transverse line; L3 flattened dorsally; axial furrows diverge less strongly opposite anterior half of L3 than opposite posterior half of L3; anterior cephalic border developed as broad ledge, visible anterior to medial parts of frontal lobe; S2 roughly transverse, weakly convex anteriorly; lateral margins of pygidial lappets curved, distal extremities of lappets pointed; pleural lappets long; anterolateral margins of first five pygidial axial rings evenly convex; anterior margins of pygidial axial rings posterior to fifth ring straight; tops of pygidial pleural segments rounded; pygidial pleural furrows long (exsag.); laterally pygidial interpleural furrows arch at same angle as medially; posterior part of pygidial pleural segments elevated above anterior part.

ETYMOLOGY: Named for Chris Braun, academic compatriot.

DISCUSSION: The species in this genus are known from the lower and upper Emsian of northern Spain, from the upper Emsian of the Massif Armorican, and from the upper Siegenian (Pragian) and Emsian of northwestern Turkey. The most completely preserved species of the genus, Braunops obsoletus (Gandl, 1972), and B. prostellans (Richter and Richter, 1939), were used to code character states for this genus. Some of the species in this genus had been assigned by Gandl (1972) and Smeenk (1983) to the genus Kayserops Delo, 1935, but it is clear that this generic assignment cannot be upheld because these species do not match the diagnosis of the genus presented by Delo (1935), nor do they resemble the type of the genus, Cryphaeus kochi Kayser, 1883 in many important features. In particular, they lack the highly convex cephalon; also Kayserops is supposed to lack a well-developed anterior border, but they have a prominent one. In addition, the species assigned to Braunops do not have L2 and L3 swollen and rising above the frontal lobe, nor is L1 narrow and depressed, as is diagnostic of Kayserops. However, in characteristics of the pygidium, some of the species of Braunops do bear a resemblance to those of Kayserops. In particular, the long thin pleural lappets curving at their lateral margins and pointed posteriorly as well as the medial axial tubercles are characteristic of K. kochi, although the first two of these character states are plesiomorphic. Moreover, some species of Braunops do have the condition of the eyes rising high above the glabella characteristic of K. kochi. These similarities are treated as convergences here-

Other species in the genus *Braunops*, such as *B. prostellans*, have in the past been assigned by authors such as Richter and Richter (1939) to *Asteropyge*. Phylogenetic topology uncovered herein indicates that *B. prostellans* is distantly related to the type of *Asteropyge*. Haas (1968) suggested that *B.*

prostellans should instead be grouped with Pseudocryphaeus. Although this species cannot be placed within that genus without either making it polyphyletic or having it encompassing a very large number of species, the species B. prostellans is included within a larger clade sister to Pseudocryphaeus.

A significant number of the characters supporting the genus *Braunops* are in the primitive state, and *B. prostellans*, which appears in the fossil record in the Siegenian (Pragian), has a large number of primitive character states. However, parsimony analysis conducted herein suggests that *Braunops* is not one of the basal members of the Asteropyginae, but rather sits on a side arm in one of the two main branches in the Asteropyginae.

Gandl (1972) suggested that what is recognized herein as *Braunops*, new genus, was closely related to *Greenops* (*Neometacanthus*) stellifer. On the basis of phylogenetic analysis this is not supported herein, and *Neometacanthus* and *Greenops* are separated on the cladogram given in figure 4 by several nodes from *Braunops*. Smeenk (1983) suggested that his *Kayserops*, here equivalent to *Braunops*, was derived from stem lineage *Paracryphaeus* Gandl, 1972. Phylogenetic analysis suggests that *Paracryphaeus* is distantly related to *Braunops*.

Braunops obsoletus (Gandl, 1972)

Kayserops n. sp. aff. diadema Carls and Gandl, 1968: 463.

Kayserops obsoletus Gandl, 1972: 128, pl. 12, figs. 1-7.

?Kayserops obsoletus Gandl. Smeenk, 1983: 403, pl. 8, figs. 7-12, text fig. 4.

TYPES: Holotype defined in Gandl (1972: pl. 12, fig. 7).

DIAGNOSIS: Same as for genus and: anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; anterior cephalic border medially flexes dorsally; tubercle on L0 developed as long spine; genal spine extends back to fourth thoracic segment; plane from medial to lateral edges of genal spine evenly inclined, medial edge thickened relative to lateral edge; lateral margin of genal spine forms line that departs from sagittal line by 15–20°; raised ridge on dorsal surface

of genal spine at juncture of posterior and lateral border furrows is smoothly curved, convex distally; axial furrows diverge at same angle anterior and posterior of S1; interior margin of genal spine deflected laterally, more strongly deflected laterally posteriorly: tubercle on medial portion of pygidial and thoracic axial rings; terminal pygidial lappet triangular, projects further posteriorly than other lappets; medial margins of pygidial lappets straight; pygidial axis constricts posterior to fifth axial ring; pygidial pleural field flanking posterior portion of axis prominently excavated; in medial region of adjacent pleural segments, posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known with certainty from the lower Emsian of the Celtiberian Mountains of northeastern Spain. It may also be present in the Emsian of the Massif Armorican as well as in the upper lower Emsian of the southern Cantabrian Mountains of northern Spain. The specimens that Smeenk (1983) figured from the La Vid Formation (upper lower Emsian) in the southern Cantabrian Mountains of northern Spain are nearly identical to the type specimens illustrated by Gandl (1972) from the lower Emsian of the Celtiberian Mountains of northeastern Spain. However, there are some slight differences between the specimens they figure, particularly in the number of axial rings: 10 in Smeenk's (1983) specimens versus 11 in Gandl's (1972) types. It could not be ascertained whether or not these differences are within the range of variation for this species, and because of this, Smeenk's (1983) specimens are only tentatively assigned to B. obsoletus.

Counts of the maximum number of lenses in a dorsoventral file on the visual surface could not be obtained for Gandl's (1972) specimens, although they could be obtained from Smeenk (1983). In these specimens there are a maximum of nine lenses per file. This character, number 6, was coded with a question mark for the purposes of phylogenetic analysis, in recognition that Smeenk's (1983) specimens might not be conspecific with Gandl's (1972).

Investigation of the photographs in Gandl (1972: pl. 12) suggests that *B. obsoletus* may

have structures homologous to the fenestrae found in some species of the asteropyginines in ENA and described as characters 65 and 66. These structures were scored as present, but this conclusion awaits further study of original specimens.

Braunops prostellans (Richter and Richter, 1939)

Cryphaeus pectinata Huffner, 1918: 371, pl. 30, fig. 5.

Asteropyge (Metacanthus) prostellans Richter and Richter, 1939: 17, pl. 17, figs. 14-15.

Pseudocryphaues prostellans (Richter and Richter). Haas, 1968: 151, pl. 35., figs. 13-15, text fig. 44.

DIAGNOSIS: S1 medially developed as smooth curve convex posteriorly; anterior margin of frontal glabellar lobe in dorsal view faintly pointed; anterior cephalic border medially does not flex dorsally; genal spine extends back to fourth thoracic segment; plane from medial to lateral edges of genal spine evenly inclined; lateral margin of genal spine in medial portion forms line that departs from sagittal line by 15-20°; raised ridge on dorsal surface of genal spine at juncture of posterior and lateral border furrows forms sharp, doglegged kink; cephalic axial furrows diverge at same angle anterior and posterior to S1; interior margin of genal spine deflected laterally; broad ledge of anterior cephalic border not visible anterior to anterolateral parts of frontal lobe; terminal pygidial lappet triangular; medial margins of pygidial lappets curved; pygidial axis constricts posterior to fourth axial ring; pygidial pleural field flanking posterior part of pygidial axis faintly excavated; in medial region of adjacent pygidial pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment.

Discussion: This species was formerly assigned to the genus *Pseudocryphaeus*, but it differs from the type species of the genus in the condition of several characters including: 2, 9, 14, 16, 19, 22, 23, 28, 29, 36, 38, 40, 42–44, 46, 48, 52, 56, 58, 59, 62, and 63 and thus can no longer be assigned to that taxon.

NEOCALMONIA PILLET, 1969

TYPE SPECIES: Neocalmonia quadricosta Pillet, 1969.

ASSIGNED TAXA: Several taxa, including: Neocalmonia (Neocalmonia) malistana Haas and Mensink, 1969, Neocalmonia (Neocalmonia) brinckmanni Haas and Mensink, 1969, Neocalmonia (Neocalmonia) imperfecta Pillet, 1969, Neocalmonia (Neocalmonia) batillifera Haas and Mensink, 1969, Neocalmonia (Neocalmonia) kakari Haas and Mensink, 1969, Neocalmonia (Neocalmonia) thaumata Haas and Mensink, 1969, and Neocalmonia cantabrica Smeenk, 1983.

DIAGNOSIS: S3 faintly sinusoidal; lateral to eye, facial suture, before flexing posteriorly, flexes anteriorly to point opposite S2; prominent change in cephalic topography where librigena contacts lateral border furrow; isolated patches of coarse tubercles present on L2, L3, and frontal lobe; medially, genal spine flattened, laterally, evenly inclined; visual surfaces, in dorsal view, present within sagittal lines drawn tangent to distal edges of frontal glabellar lobe; S2 transverse, weakly convex anteriorly; S1 crescent shaped, medially straight; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view: anterior cephalic border pointed medially in dorsal view, projecting medially and laterally far beyond frontal lobe; fixigena evenly sloping from axial furrow to palpebrum; PMI developed as linear incision; posterior margin of palpebrum relative to anterior margin deflected at angle to transverse line that is significantly larger; distal tips of pygidial lappets pointed; pygidial axis constricts posterior to fifth axial ring; pygidial pleural field flanking posterior portion of axis faintly excavated; posterior portion of pleural segment elevated above anterior portion; fourth pygidial pleural lappet projects furthest posteriorly; tops of pygidial pleural segments flat; pygidial interpleural furrows flex anteriorly laterally; posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; pygidial lappets long (exsag.); four pairs of pygidial lappets present.

DISCUSSION: The species in this genus are known from the Frasnian of Afghanistan and the lower Givetian of northern Spain. Several authors, including Haas and Mensink (1969) and Haas (1970), have suggested that Neocalmonia be divided into several different subgenera such as Neocalmonia (Neocalmonia), Neocalmonia (Bradocryphaeus), and Neocalmonia (Heliopyge). Herein, these subgeneric classifications are not followed. Instead, each of these subgenera is elevated to full generic status, with Neocalmonia now comprising what had traditionally been treated as Neocalmonia (Neocalmonia).

Smeenk (1983) figured and described a new species from the lower Givetian of the Cantabrian Mountains of northern Spain that he referred to as Neocalmonia cantabrica. Although the cephala of his species are homogeneous, there appear to be differences in the pygidia he recovered, with some possessing four pairs of pleural lappets and others with five pairs. In the latter, the posteriormost pair appears connected to the terminal pygidial lappet. Smeenk (1983) commented that this latter condition was also present in Bradocryphaeus afghanica Haas and Mensink, 1969, and he viewed his N. cantabrica as transitional between Bradocryphaeus and typical Neocalmonia, which possesses four pairs of pleural lappets. However, Smeenk (1983) recognized that the types of Bradocryphaeus and Neocalmonia do differ in the condition of several characters. At present, it is difficult to ascertain whether or not Smeenk's (1983) suite of specimens belongs to a single species or two species. There are clear differences between some of the pygidia he illustrates, even though these pygidia are poorly preserved. For the present, the holotype of N. cantabrica (Smeenk, 1983: pl. 17, fig. 3) as well as another specimen illustrated by Smeenk (1983: pl. 17, fig. 4) are treated as members of a valid species of Neocalmonia. However, some of the pygidia Smeenk (1983) illustrated, as well as all of the cephala he figured, appear to bear several characters diagnostic of Bradocryphaeus rather than Neocalmonia. In particular: the lateral edge of S2 is directed posteriorly; there are five pairs of pygidial pleural lappets, with the fifth pair extending from the terminal lappet; the third lappet projects furthest posteriorly; and the medial portion of S1 is developed as a smooth curve convex posteriorly. Therefore, these pygidia and cephala are treated as Bradocryphaeus new

species. Smeenk (1983) may be correct about the transitional position of *N. cantabrica* between what he claims to be the ancestral *Bradocryphaeus* Haas and Mensink, 1969, and the derived *Neocalmonia*, but his conclusion is not supported by the phylogenetic analysis presented herein, which suggests that *Neocalmonia* is not especially closely related to *Bradocryphaeus* Haas and Mensink, 1969, or to *Heliopyge* Haas and Mensink, 1969.

Several authors have commented on the phylogenetic relationships of Neocalmonia. Haas and Mensink (1969) asserted that Bradocryphaeus was derived from Asteropyge Hawle and Corda, 1847, in the late Eifelian, and Bradocryphaeus gave rise to Heliopyge in the early Givetian and Neocalmonia in the late Givetian. This phylogenetic topology is duplicated in Haas (1970) (see fig. 5a). On the basis of the phylogenetic analysis conducted herein, Asteropyge appears to be distantly related to all of these genera. Part of Haas and Mensink's (1969) phylogenetic conclusions are supported in that Bradocryphaeus and Heliopyge are closely related, although not sister taxa. However, Neocalmonia is part of a larger clade that is in turn sister to a clade including Bradocryphaeus and Heliopyge, and thus Neocalmonia is not closely related to either of these genera. Arbizu (1979) suggested that Alcaldops gave rise to the genus *Heliopyge*, which gave rise to *Bra*docryphaeus, which in turn gave rise to Neocalmonia (see fig. 5c). Alcaldops was not analyzed herein because species of this genus were not available. However, this genus does appear to be very closely related to Comura and Tolkienia, as discussed above. On the basis of the phylogenetic analysis presented herein, Tolkienia, new genus, is relatively closely related to Neocalmonia, with these genera being separated by two nodes on the cladogram given in figure 4. Again, Arbizu's (1979) phylogeny does partially disagree with the scheme of evolutionary relationships depicted herein, in terms of the close relationship he posits between Bradocryphaeus, Heliopyge, and Neocalmonia. Morzadec's (1983) phylogeny is very similar to that of Arbizu (1979), differing solely in substituting Comura for Alcaldops (see fig. 5e). Smeenk (1983) asserted that Comura gave rise to Bradocryphaeus, which in turn gave

rise first to Alcaldops and then to Neocalmonia (see fig. 5d). According to Smeenk (1983), Alcaldops also gave rise to Heliopyge. Smeenk (1983) felt that this entire clade had a separate origin from the rest of the Asteropyginae. Smeenk (1983) did not speculate on what might be the predecessor of this separate lineage. This phylogenetic perspective of Smeenk (1983) is not supported herein.

The two lineages sister to the clade containing Neocalmonia are known to first appear in the late Siegenian (Pragian) and the Emsian, respectively. This implies a gap from the late Siegenian to the lower Givetian in the fossil record, when species in the lineage leading to Neocalmonia should have been recovered but are unknown. This gap could be due to inadequate sampling of that time interval, but this appears unlikely. However, such species may have persisted in regions that cannot be sampled, either because they were not preserved in the fossil record or because they are in inaccessible areas. Finally, as discussed above, under the heading of Braunops prostellans, the actual age of first appearance was found to show some discordance with the predicted age of first appearance. In addition, several putative reversals to the primitive character state were concentrated at the node defining Braunops. If these primitive character states were misidentified as reversals, it is possible that the gap between the age of first appearance of Braunops and the first appearance of Neocalmonia could be an artifact of mistakes in cladogram topology. However, as such issues require additional analysis, they await further exploration by other authors.

Neocalmonia quadricosta Pillet, 1969

Neocalmonia quadricosta Pillet, 1969: 144, figs. A, B, B1.

Neocalmonia (Neocalmonia) quadricosta Pillet. Haas and Mensink, 1969: 175, pl. 5, figs. 1-7, text fig. 6.

TYPES: Holotype defined in Pillet (1969: fig. B1).

DIAGNOSIS: Line from posterior to anterior edge of eye forms 30° angle with sagittal line; maximum of 10 lenses per dorsoventral

file on visual surface; genal spine developed as long flange extending back to sixth thoracic segment; anterior margins of first five pygidial axial rings from axial furrow to midline straight, then strongly arch anteriorly; prosopon of coarse tubercles; lateral and medial margins of pygidial pleural lappets curving; pygidial pleural lappets project further posteriorly than terminal pygidial lappet; terminal pygidial lappet broad, triangular; 14 pygidial axial rings.

DISCUSSION: This species is known from the Frasnian of Afghanistan. It was chosen for phylogenetic analysis because it is the type of the genus.

HALLANDCLARKEOPS, NEW GENUS

TYPE SPECIES: Kayserops? cantarmoricus Morzadec and Arbizu, 1978.

ASSIGNED TAXA: Asteropyge renaudae Morzadec, 1969, Heliopyge troaonensis Morzadec, 1983, and Asteropyge quelernensis Morzadec, 1983.

ETYMOLOGY: Named by combining the names of J. Hall and J. M. Clarke, who both contributed so much to our knowledge of the Devonian of North America.

DIAGNOSIS: Fixigena with distinct bulge medially; S3 faintly sinusoidal; cephalic posterior border furrow flexes anteriorly laterally; S2 roughly transverse, weakly convex anteriorly; posterior edge of visual surfaces anterior to transverse line tangent to L0; distance between frontal lobe and facial suture increases from lateral edge to midline; anterior cephalic border medially flexes dorsally, deflected far anteriorly medially, developed as spatulate process; PMI developed as linear incision; posterior margin of palpebrum relative to anterior margin deflected at angle to transverse line that is roughly equivalent; prosopon of fine granules; isolated patches of coarse tubercles on L3 and L2 present; genal spine developed as long flange extending back to seventh thoracic segment; medially genal spine developed as flattened ledge, laterally evenly inclined; cephalic axial furrows diverge at same angle anterior and posterior to S1; terminal pygidial lappet triangular; pygidial lappets project further posteriorly than terminal lappet; medial margins of pygidial lappets straight, lateral margins curved; py-

gidial axis constricts posterior to sixth axial ring; anterior margins of pygidial axial rings posterior to sixth axial ring smoothly convex anteriorly: pygidial pleural field flanking posterior part of axis moderately excavated; posterior part of pleural segment elevated above anterior part; tops of pygidial pleural segments rounded; pygidial pleural furrows developed as long (exsag.) excavated spaces; at pygidial margin surface of pygidium dorsally inflates to form fringing ring; laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pygidial pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; terminal pygidial lappet narrow (tr.); pygidial lappets long (exsag.).

DISCUSSION: This genus is known from the Couvinian (roughly Eifelian) and Couvinian-Givetian transition in the Cantabrian Mountains of northern Spain as well as from the Eifelian of the Massif Armorican. Authors such as Morzadec and Arbizu (1978) and Arbizu (1979) suggested that what is here treated as the type species of Hallandclarkeons. new genus, belonged to the genus Kayserops Delo, 1935. However, H. cantarmoricus (Morzadec) lacks several of the diagnostic characteristics of Kayserops, and H. cantarmoricus cannot be considered to be closely related to the type of Kayserops. Later, Morzadec (1983) changed his mind and treated H. cantarmoricus as well as the closely related Asteropyge quelernensis Morzadec, 1983, as species of Asteropyge. Again, H. cantarmoricus differs from the type species of Asteropyge in the condition of several characters, including those diagnostic of Asteropyge, and the two taxa cannot be considered closely related. Other species herein assigned to Hallandclarkeops had been assigned by Morzadec (1983) to Heliopyge Hans and Mensink, 1969.

The phylogenetic position of only one species of *Hallandclarkeops* was assessed: *H. cantarmoricus*, the type of the genus. This species is sister to *Bellacartwrightia*, a genus of ENA asteropyginines. It appears in the fossil record around the same time as *Bellacartwrightia* does. Although the other species assigned to *Hallandclarkeops* were not subjected to phylogenetic analysis, it is be-

lieved that they are all very closely related to *H. cantarmoricus* because they share a significant number of apomorphies with that species.

Hallandclarkeops cantarmoricus (Morzadec and Arbizu, 1978)

Kayserops? cantarmoricus Morzadec and Arbizu, 1978: 925, pl. 1, figs. 1-9; Arbizu, 1979: 76, pl. 5, figs. 1-4.

Neocalmonia (Heliopyge) n. sp. Gandl in Carls, 1979: 38.

Asteropyge cantarmorica (Morzadec and Arbizu). Morzadec, 1983: 132, pl. 19, figs. 1-6, 8, pl. 20, figs. 1-4 (see for complete synonymy).

TYPES: Holotype figured in Morzadec (1983: pl. 20, fig. 1).

DIAGNOSIS: Line from posterior to anterior edge of visual surface forms 5° angle with sagittal line; maximum of 10 lenses per dorsoventral file on visual surface; lateral margins of spatulate process at anterior edge of cephalon first diverge anteriorly, then converge; 18 pygidial axial rings.

DISCUSSION: This species is known from the Eifelian of the Massif Armorican, Morzadec (1983) suggested that this species was sister to Asteropyge punctata (Steininger, 1831), the type of the genus Asteropyge, and both were derived from what he perceived as a paraphyletic Comura lineage. However, as mentioned above, this species is distantly related to Asteropyge punctata, differing in the condition of many characters used in phylogenetic analysis. According to the phylogenetic analysis, this species is most closely related to Bellacartwrightia, new genus, a genus containing several species, some of which were formerly mistakenly classified as members of Greenops. (However, there probably are other species of Hallandclarkeops that may be more closely related to Bellacartwrightia but that could not be considered in the phylogenetic analysis.) Illustrations in Morzadec (1983) suggest that this species may possess fenestrae on the thoracic pleural segments. It was questionably scored as having these until the actual specimens can be examined.

All articulated cephalothoraxes of this species have largely obscured thoracic regions. Thus, the exact thoracic segment that the genal spines extended back to (character 26)

had to be estimated. This estimated position, the seventh segment, was used in the phylogenetic analysis.

BELLACARTWRIGHTIA, NEW GENUS

TYPE SPECIES: Bellacartwrightia jennyae, new species.

DIAGNOSIS: S3 faintly sinusoidal; facial suture lateral to eye, before flexing posteriorly flexes anteriorly to point between distal tips of S1 and S2: distance between frontal lobe and facial suture at midline relative to distance between frontal lobe and facial suture at lateral margins of frontal lobe equivalent; anteriormost portion of palpebrum reaches point between medial tips of L3 and PMI; medially, anterior cephalic border in dorsal view pointed; 8-10 lenses in a dorsoventral file on visual surface; anterior cephalic border developed as moderate lip, medially deflected anteriorly for length (sag.) of L1 in front of facial suture; broad ledge of anterior cephalic border visible anterior to anterolateral parts of frontal lobe; posterior margin of palpebrum relative to anterior margin deflected at angle to transverse line that is roughly equivalent; L3 flattened dorsally; significant change in cephalic topography where librigena meets lateral border furrow; isolated patches of tubercles on L3; plane from medial to lateral edges of genal spine evenly inclined; medial edge of genal spine significantly thickened relative to lateral ledge; lateral margin of genal spine parallels sagittal line; raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border furrow forms sharp, doglegged kink; interior margin of genal spine evenly deflected laterally, more strongly deflected posteriorly; tubercles or spines on medial portion of thoracic and pygidial axial rings; terminal pygidial lappet triangular, does not project as far posteriorly as fifth pair of pleural lappets; 14-16 pygidial axial rings; fifth pair of lappets project furthest posteriorly; medial margins of pygidial lappets straight, distal extremities pointed; pygidial pleural field flanking posterior part of pygidial axis prominently excavated; tops of pygidial pleural segments rounded; pygidial pleural furrows long (exsag.), almost as long as anterior pleural band; anterolateral margins of first five pygidial axial rings evenly convex; pygidial lappets long (exsag.) equal to the length of the first three pygidial axial rings (sag.).

Assigned Taxa: Bellacartwrightia whiteleyi, new species, B. phyllocaudata, new species, B. calderonae, new species, Cryphaeus calliteles Green, 1837, and Dalmanites (Cryphaeus) pleione Hall, 1861.

ETYMOLOGY: Named for Paulyn Cartwright, BSL's wife.

DISCUSSION: This genus is based on several species that have traditionally been assigned to the genus Greenops (e.g., Delo, 1935, 1940; Babcock and Speyer, 1987; Speyer, 1988, etc.), although sometimes (e.g., Ludvigsen, 1979; Morzadec, 1992) they had been treated as belonging to the subgenus Neometacanthus Richter and Richter, 1948. As discussed below under the heading of Neometacanthus, species of Bellacartwrightia differ substantially from the type of Neometacanthus, N. stellifer, in the condition of several characters, and they are significantly distanced from one another on the cladogram shown in figure 4. Thus, they cannot be treated as congeners.

In addition, the members of this genus are phylogenetically distant from species assigned to true *Greenops* Delo, 1935. Thus, past authors had been lumping very distinct phylogenetic lineages within the Asteropyginae when they assigned all ENA asteropyginines to the genus *Greenops*. These two Middle Devonian genera have not shared a common ancestor since, at latest, the Siegenian (?Pragian), based on an analysis of ghost lineages. To treat these species as members of a genus *Greenops* would necessitate placing all of the asteropyginines within the genus *Greenops*.

The presence of this genus in ENA and its phylogenetic position relative to *Greenops* have implications for the appearance of the Hamilton Group fauna in ENA. Typically, most of the asteropyginine species in ENA were viewed as a largely endemic radiation (e.g., Hall and Clarke, 1888; Delo, 1940; Stumm, 1953; Morzadec, 1983, 1992; etc.). However, the phylogenetic analysis conducted herein calls this contention into question. Rather, it suggests that the appearance of the asteropyginines in ENA was the by product

of multiple biogeographic events involving invasion by Armorican lineages into ENA, and the ENA asteropyginines can be partitioned into small groups of closely related species nested within Armorican species, with these groups separated by significant phylogenetic distance. A method is discussed below for using phylogenetic analysis in conjunction with the principle of parsimony to resolve biogeographic area relationships. The method indicates that invasions occurred between the Emsian and the Givetian involving lineages entering into ENA from Armorica. The evidence suggests that the invasions represent at least four independent biogeographic events. However, whether or not they occurred contemporaneously is difficult to ascertain.

As discussed by Delo (1940), the species originally described as Cryphaeus collitelus Green, 1837, was based on types from strata near Huntingdon, Pennsylvania, that were subsequently lost. Authors such as Hall (1876) and Hall and Clarke (1888) recovered material from New York that they claimed was conspecific with that species, and they assigned this material to Dalmanites (Cryphaeus) boothi var. calliteles (apparently the original spelling of collitelus was a typographical error). Delo (1940) supported their conclusions. For several reasons, the decisions of Hall and Clarke (1888) and Delo (1940) regarding these New York specimens must be questioned.

First, these authors did not make an adequate attempt to recover material from Green's (1837) type locality to verify if the New York material differed from the Pennsylvania material. Until that is done, one cannot be sure that the material Hall and Clarke (1888) illustrated, which subsequent authors such as Grabau and Shimer (1910) and Delo (1940) also figured, actually represented that species. Instead of following Hall and Clarke's (1888) taxonomic assignment of much of their material to D. (C.) boothi var. calliteles, we are at present making excursions to recover asteropyginine material from Green's (1837) type locality. When such material is recovered it will be possible to erect a neotype for Cryphaeus calliteles. At this time, our analysis cannot incorporate specimens that could be assigned to calliteles.

Hall and Clarke (1888) are unlikely to be correct in emphasizing the exact match between specimens from New York that they treated as calliteles and true specimens of that species from the type locality in Pennsylvania because they assigned several different New York Hamilton Group species to D. (C.) boothi var. calliteles. At this time, the different species from New York State assigned by Hall and Clarke (1888) to D. (C.) boothi var. calliteles are assigned to two new species. A subsequent paper (Lieberman and Kloc, in prep.) will address the issue of the species calliteles in greater detail.

Delo's (1940) analysis of *Greenops boothi* var. *calliteles* largely followed that of Hall and Clarke (1888), and for that reason his assignments also cannot be accepted.

If there are similarities between the Pennsylvanian material and the bulk of the aster-opyginid material Hall and Clarke (1888) and Delo (1940) assigned to calliteles, then it is likely that a redefined calliteles based on a neotype would be assigned to Bellacartwrightia, so tentatively calliteles is treated as a taxon referable to that genus.

Two of the species of Bellacartwrightia analyzed herein are not known with certainty from rocks older than the Givetian portion of the Erian; however, Hall and Clarke (1888) claimed that a specimen which on the basis of its morphology we assert belongs to either B. phyllocaudata or B. jennyae was found in the Onondaga Limestone, and would therefore be pre-Givetian. We could not verify this stratigraphic assignment, and it is treated as questionable. However, B. pleione (Hall, 1861) is known from the Jeffersonville Limestone (Southwoodian), equivalent to the Onondaga Limestone, from the Falls of the Ohio, Kentucky, in the Illinois Basin, and it also occurs in the Hamilton Group and possibly the Onondaga Limestone of the Appalachian Basin. Thus, the genus Bellacartwrightia ranges from the Southwoodian (Eifelian) to the Taghanic (Givetian), with the latest occurrences in the Appalachian Basin.

Bellacartwrightia pleione was not included in the phylogenetic analysis because of the paucity of adequately preserved material, and the absence of cephalic remains. However, based on the diagnostic pygidial characteristics of all other species of Bellacart-

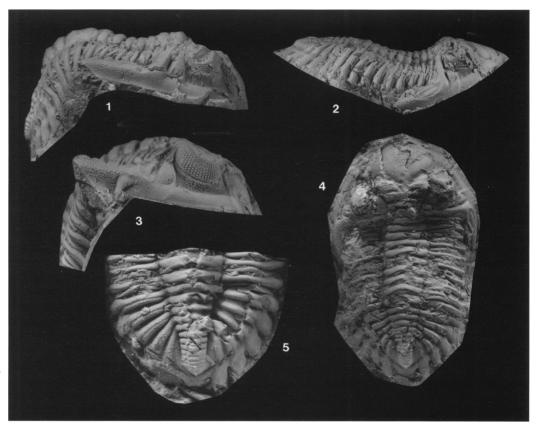


Fig. 6. Philipsmithiana hyfinkeli, new species, Middle Devonian, Morocco. 1. AMNH 45261, lateral view of entire specimen, ×1.3. 2, 4. AMNH 45262, lateral and dorsal views of entire specimen, ×1.5. 3, 5. AMNH 45263, lateral view of cephalon and portion of thorax and dorsal view of pygidium and portion of thorax, ×2.5.

wrightia, it appears likely that pleione should be placed within this genus. Recovery of additional material would make it possible to incorporate this species into phylogenetic analysis and determine its phylogenetic position with greater confidence.

Bellacartwrightia jennyae, new species

Figures 14.1, 14.2, 16, 20.3-20.6, 20.8-20.11

Dalmanites (Cryphaeus) boothi var. calliteles (Green). Hall and Clarke, 1888: 45, pl. 16, figs. 8, 11-13, 19-21, 22, pl. 16a, figs. 9, 11, 13-15, pl. 24, fig. 15.

Neometacanthus calliteles (Green). Ludvigsen, 1979: 80, fig. 57d.

Greenops boothi (Green). Delo, 1940: pl. 10, fig. 17; Eldredge, 1971a: pl. 14, figs. 4, 5.

Greenops boothi, var. calliteles (Green). Delo, 1940: pl. 10, fig. 19.

?Dalmanites (Cryphaeus) boothi, var. calliteles (Green). Hall and Clarke, 1888: 45, pl. 16a, fig. 16.

TYPES: Holotype AMNH 45312, a complete specimen from the Centerfield Limestone (Tioughniogan), Ludlowville Formation, Hamilton Group, Middle Devonian, Brown's Creek, York, New York State; paratypes AMNH 45310, a complete individual from the same locality as the holotype, and AMNH 36998, a complete individual from the Centerfield Limestone, Centerfield, New York.

DIAGNOSIS: S2 weakly declined posteriorly at lateral edge; S1 developed medially as smooth curve convex posteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; anterior margin of frontal glabellar lobe in dorsal

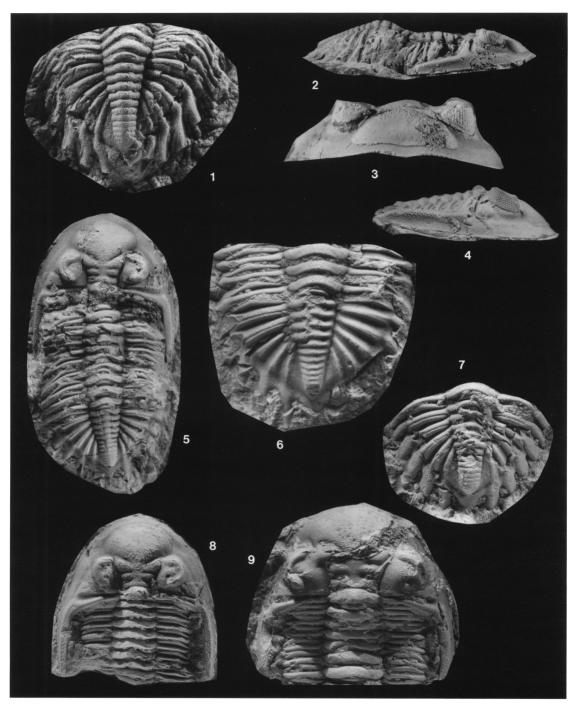


Fig. 7. Asteropyge and Philipsmithiana spp. 1. Asteropyge punctata (Steininger, 1831), Middle Devonian, Eifel District, Germany, YPM 6643, dorsal view of pygidium and portion of thorax, ×2.75. 2–9. Middle Devonian, Morocco. 2–6, 8. Philipsmithiana burtandmimiae, new species. 2, 3, 5. AMNH 45268, lateral, anterior and dorsal views of entire specimen, ×1.8, ×1.8, and ×2.3. 4, 8. AMNH 45269, lateral and dorsal views of cephalon and portion of thorax, ×1.9. 6. AMNH 45270, dorsal view of pygidium, ×2.0. 7, 9. Philipsmithiana finkeli, new species, AMNH 45261, dorsal view of pygidium and dorsal view of cephalon and portion of thorax, ×1.5 and ×2.

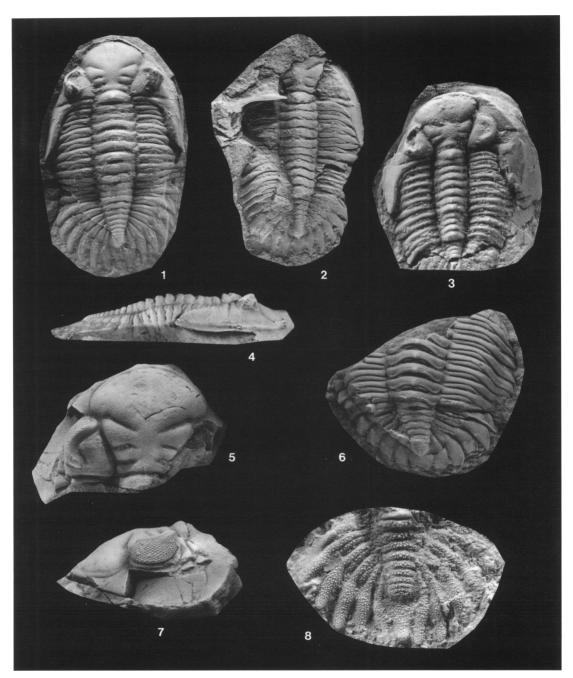


Fig. 8. Greenops, Kennacryphaeus, and Stummiana spp. 1, 2, 4. Greenops barberi, new species, Wanakah Shale, Ludlowville Formation, Father's Camp, Lake Erie, New York. 1, 4. AMNH 45256, dorsal and lateral views of latex cast taken from external mold of entire specimen, ×2.5. 2. AMNH 45257, dorsal view of nearly complete specimen, ×3. 3. Greenops new species from the Kashong Shale, Middle Devonian, Moscow, New York, AMNH 45255, dorsal view of cephalothorax and partial pygidium, ×3.5. 6. Greenops grabaui, new species, Hamilton Group, New York, AMNH 45317, dorsal view of thoracopygidium, ×3.5. 5, 7. Kennacryphaeus harrisae, new species, ?Mottville Member, Skaneateles Formation, Fitch Road, New York, AMNH 45248, dorsal and lateral views of partial cephalon, ×2.5. 8. Stummiana arkonensis (Stumm, 1953), Alpena Limestone (Cazenovian), the Huron Point Cement Company, Alpena, Michigan, AMNH locality 3059, AMNH 45249, dorsal view of pygidium, ×4.

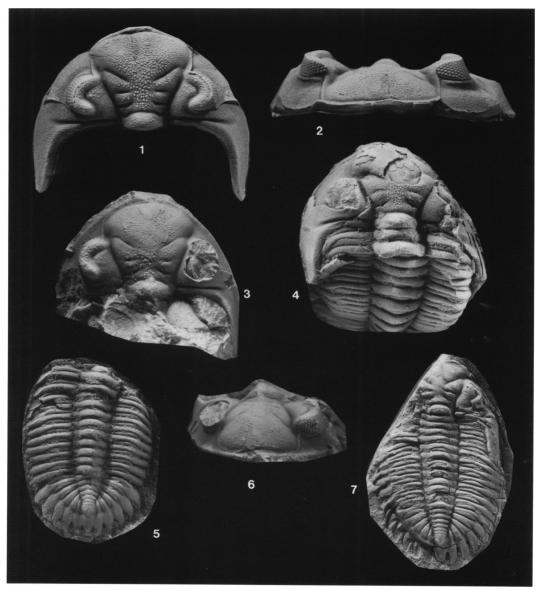


Fig. 9. Greenops spp. 1, 2. Greenops widderensis, new species, ?Widder Formation, Traverse Group, Arkona, Ontario, Canada, ROM 1731H, dorsal and anterior views of cephalon, ×4.5. 3–7. Greenops grabaui, new species. 3, 6. Hamilton Group, shore of Lake Erie, Hamburg, New York, YPM 33778, dorsal and anterior views of cephalon, ×5.75. 4. YPM 33788, dorsal view of cephalothorax, ×3. 5, 7. Greenops grabaui, new species, Hamilton Group, New York. 5. AMNH 45274, dorsal view of thoracopygidium, ×3.5. 7. AMNH 45275, dorsal view of nearly complete specimen, ×2.5.

view faintly pointed; spine on L0; PMI developed as linear incision; line from posterior to anterior edge of eye forms 5-10° angle to sagittal line; maximum of 10 lenses per dorsoventral file on visual surface; cephalic axial furrows anterior to S1 diverge at same

angle, at anterior half of L3 relative to posterior half equally divergent; visual surfaces in dorsal view present within sagittal line drawn tangent to frontal lobe; one transverse row of fenestrae on anterior and posterior bands of thoracic pleural segments; genal spine developed as long spine extending back to eighth thoracic segment; lateral margins of genal spine recurve posteriorly at posterior end; spine on medial portion of pygidial and thoracic axial rings; pygidial axis constricts posterior to sixth axial ring; anterior margin of pygidial axial rings posterior to constriction of pygidial axis straight; posterior parts of pygidial pleural segments elevated above anterior parts; laterally pygidial interpleural furrows arch at same angle as medially; in medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; terminal pygidial lappet narrow; lateral margins of pygidial pleural lappets curved; 14 pygidial axial rings.

ETYMOLOGY: Named for Ms. Jenny Lieberman, BSL's sister.

MATERIAL: AMNH 28910, 36998, 45295, 45296, 45310–45312; NYSM 4289–4294, 4297, 4298, 4300, 4301, 4303, 4304; YPM 33763, 74585.

DESCRIPTION: Cephalic length (sag.) 55% of width. Axial furrow nearly straight anterior to S0, diverging forward at about 25°, narrow, shallow. Cranidial anterior border developed as moderate lip deflected moderately anterior to facial suture. Preglabellar furrow shallow and narrow. Cranidial anterior border of constant length (sag.) from lateral to medial edges of frontal lobe. Facial suture anterior to eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly, then posteriorly laterally. Glabellar length equal to width across frontal lobe; frontal lobe ellipsoidal, about 65% of length of glabella; frontal lobe at level of posterior glabellar region; anterior part of frontal lobe moderately declined forward, faintly pointed in dorsal view. PMI shallow linear incision. S3 shallow, weakly sinusoidal, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, bearing isolated tubercles, lengthening distally, outer edge forming smaller angle with sagittal line at anterior end than at posterior end. S2 weakly declined posteriorly at lateral edge, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial part developed as smooth curve convex posteriorly, laterally contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly medially in dorsal view, deep. L0 55% of width of frontal lobe, moderately arched (tr.), longest sagittally, bearing spine on medial portion. Large Eye Index 0.55. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite S0, posterior to transverse line tangent to L0; eye raised prominently above frontal lobe; maximum of 10 lenses per dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 15% of length of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, weakly flexing anteriorly distally, anterior margin nearly straight. Genal spine developed as very long flange extending back to eighth thoracic segment, cephalic border furrow bisects spine; lateral and medial edges of genal spine evenly inclined; medial edge of genal spine significantly thickened and swollen relative to lateral edge, deflected more strongly laterally posteriorly; lateral margin of genal spine parallels sagittal line; lateral margin of genal spine at posterior end recurves; interior margin of genal spine deflected laterally. Raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border furrow forms sharp, doglegged kink. Lateral border furrow narrow, shallow, lateral border weakly widens posteriorly. Anterior margin of cephalic doublure bluntly pointed. Prosopon of fine tubercles.

Hypostome parabolic, width (tr.) at midlength about 110% of sagittal length. Course of anterior margin (hypostomal suture) weakly convex, with moderate wings positioned far forward. Lateral margin tapering behind anterior wings at approximately 30° angle relative to sagittal line. Middle body bean shaped in outline, broadest anterior to midlength, anterior margin poorly defined; anteriolateral margins defined by smooth break in slope, grading down to rather flattened anterior wing. Convexity (tr.) of middle border moderate sagittally; faintly convex (sag.) anterior and posterior lobes separated at about

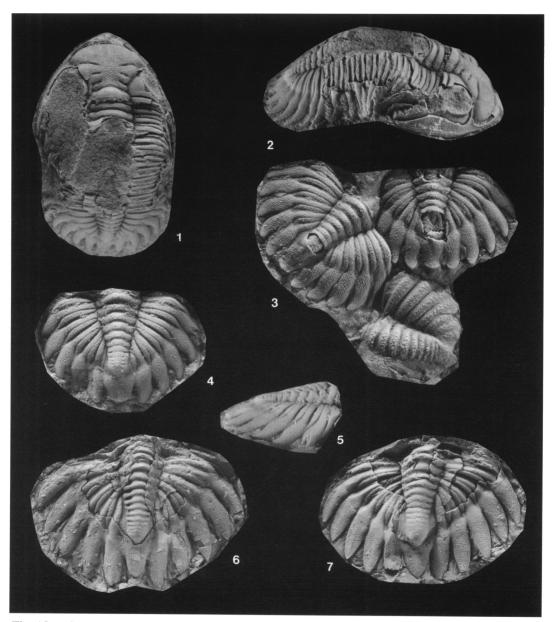


Fig. 10. Greenops and Bellacartwrightia spp. 1-5. Greenops grabaui, new species, Hamilton Group, New York. 1, 2. Near 18 Mile Creek, YPM 33875, dorsal and lateral views of entire specimen, ×4.5.

3. Near 18 Mile Creek, YPM 33783, 33784, dorsal view of two pygidia and one partial pygidium of Phacops rana (Green, 1832), YPM 33785, ×3.5. 4, 5. AMNH 45254, dorsal and lateral views of pygidium, ×3.2. 6, 7. Bellacartwrightia phyllocaudata, new species, Moscow Formation, Hamilton Group, Jaycox Run Creek, near Avon, Geneseo Township, Livingston County, New York. 6. AMNH 45244, dorsal view of pygidium, ×2. 7. AMNH 45243, dorsal view of pygidium, ×1.75.

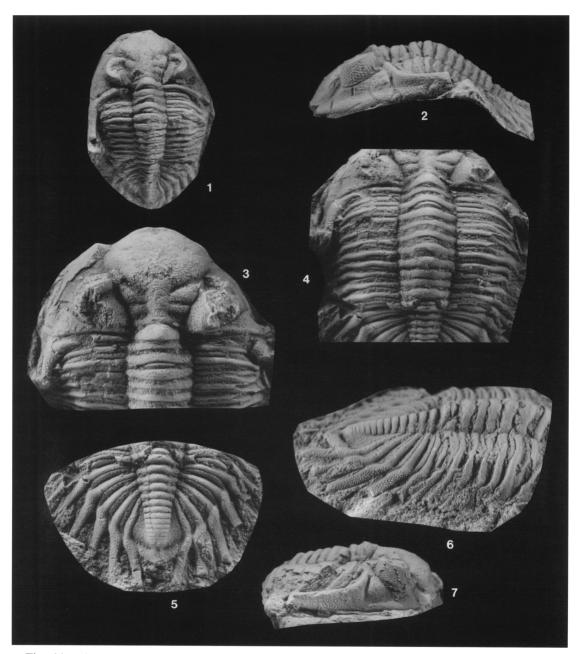


Fig. 11. Asteropyge punctata (Steininger, 1831). 1–6. Calceola Beds (Eifelian), Gees Gerolstein, Eifel District, Germany, 1, 2. AMNH 18767, dorsal and lateral views of entire specimen, $\times 3$ and $\times 3.5$. 3–6. AMNH 18768, dorsal view of cephalon, dorsal view of thorax, and dorsal and lateral views of pygidium and portion of thorax, $\times 4$, $\times 3.5$, $\times 3.75$, and $\times 3.75$. 7. Eifel District, Germany, YPM 6643, lateral views of cephalon and portion of thorax, $\times 2.75$.

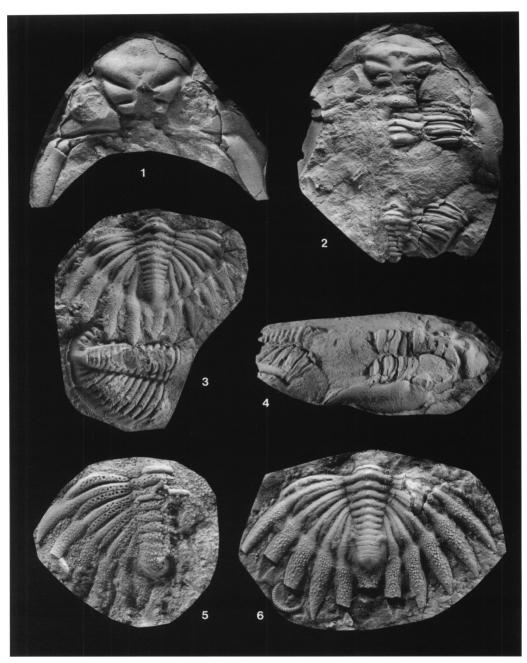


Fig. 12. Bellacartwrightia and Kayserops spp. 1–4. Bellacartwrightia phyllocaudata, new species, Deep Run Member, Moscow Formation, Jaycox Run Creek, near Avon, Geneseo Township, Livingston County, New York. 1. AMNH 45247, dorsal view of cephalon, ×2.5. 2, 4. AMNH 45246, dorsal and lateral views of nearly complete specimen, ×1.75. 3. AMNH 45245, dorsal view of pygidium and partial pygidium of Monodechenella macrocephala (Hall, 1861) (AMNH 44760), ×2.25. 5. Kayserops traversensis (Stumm, 1953), Traverse Group, Four Mile Dam Limestone (Tioughniogan), Four Mile Dam, Alpena, Michigan, dorsal view of partial pygidium, AMNH 45250, ×4.25. 6. Bellacartwrightia whiteleyi, new species, ?Ludlowville Formation, Canandaigua Lake, New York, AMNH 45251, dorsal view of pygidium, ×2.5.



Fig. 13. Bellacartwrightia spp. 1–3. Bellacartwrightia pleione (Hall, 1861). 1, 2. Deep Run Member, Moscow Formation, Jaycox Run Creek, near Avon, Geneseo Township, Livingston County, New York, AMNH 45252, dorsal and lateral views of pygidium, ×2.5. 3. Jeffersonville Limestone, falls of the Ohio River, near Louisville, Kentucky, AMNH 4248, dorsal view of holotype pygidium, ×2.5. 4. Bellacartwrightia calderonae, new species, ?Windom Shale (Tioughniogan), Moscow Formation, Kashong Glen, west side of Seneca Lake, Bellona, New York, AMNH 29215, dorsal view of entire specimen, ×1.5. 5. Bellacartwrightia sp. aff. jennyae, new species, from the Windom Shale, Moscow Formation, western New York, YPM 33789, dorsal view of thoracopygidium, ×3.

65% of length by broad, shallow middle furrow. Maculae small, low, circular swellings, positioned back about 55% of sagittal length of hypostome on middle furrow, displaced medially relative to lateral border furrow 25% of width of hypostome at midlength. Posterolateral-posterior border furrows nar-

row, shallow. Posterolateral border of constant length (exsag., sag.), 10% of length (sag.) of hypostome. Posterior margin rounded, apparently lacking spines.

Thorax of 11 segments. Axial ring about 25% of width of thorax, of equal length (exsag., sag.), at distal ends and medially



Fig. 14. *Bellacartwrightia jennyae*, new species, Centerfield Limestone, Ludlowville Formation, Centerfield, New York. 1, 2. Dorsal and lateral views of paratype specimen, AMNH 36998, ×2.75.

flexed forward. Ring furrow of equal length (exsag., sag.), moderately incised, flexing forward medially and distally. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; scattered tubercles around medial part of axial ring, centered around medially placed, enlarged spine; single circular fenestra flanks each side of medial spine; at lateral margin of axial rings, circular fenestrae present. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.), outer part of pleu-

rae more strongly flexed downward, straight in posterior segments. Posterior band long (exsag.), anterior margin straight, deflected at constant, small angle posteriorly; length (exsag.) shortens as length of pleural furrow increases medially, after approximate midpoint, length of pleural furrow decreases laterally, length of posterior band increases, becomes planar; posterior surface declined posterodorsally, has one transverse row of circular fenestrae near axis: anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially anterior margin transverse; deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally, with one transverse row of circular fenestrae.

Pygidium broadly triangular in outline excluding marginal lappets, length (sag.) about 65% of width (tr.) without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 20 degree angle anterior to sixth pygidial axial ring, roughly parallel posterior to sixth pygidial axial ring. Axis 30% of pygidial width anteriorly, with 14 rings; rings of nearly equal length distally (exsag.) and medially (sag.), bearing medial spine; anterior and posterior edge of axial rings posterior to sixth axial ring from midline to distal edge convex, then transverse. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips pointed. Axial terminus triangular, does not project as far posteriorly as other lappets; postaxial region 20% of pygidial length (sag.). Pygidial pleural field flanking posterior part of pygidial axis moderately excavated; anterior band of pleural segment elevated above posterior band; tops of segments rounded; pleural furrows deeper than interpleural furrows, long (exsag.). Laterally pygidial interpleural furrows arch at same angle as medially. In medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment.

DISCUSSION: This species is known from the following localities and stratigraphic horizons in the Appalachian Basin of ENA: the Centerfield Limestone (Tioughniogan), Ludlowville Formation, Buffalo Creek, Blossom,

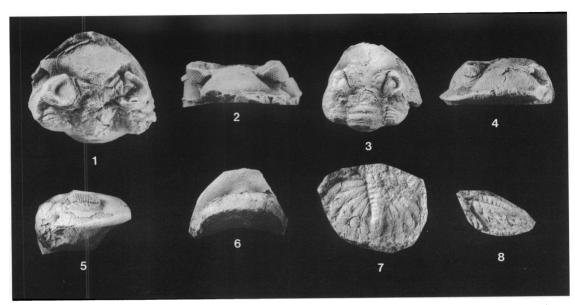


Fig. 15. Rhenops new species, Frasnian or Famennian of Venezuela. 1, 2. Dorsal and anterior views of cephalon, $\times 1.1$. 3–6. Dorsal, anterior, lateral, and ventral views of cephalon, $\times 1$ 7, 8. Anterior and lateral views of pygidium, $\times 1.2$. All housed in the Department of Geology, University of Rio de Janeiro, Brazil.

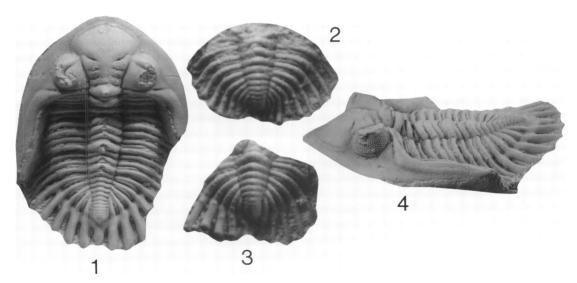


Fig. 16. Bellacartwrightia jennyae, new species, Centerfield Limestone, Ludlowville Formation, Centerfield, New York. 1, 4. NYSM 4293, dorsal and lateral views of complete specimen, ×1.5. 2. NYSM 4290, dorsal view of meraspid (M9) pygidium, ×11. 3. NYSM 4289, dorsal view of meraspid (M6) pygidium, ×10.

New York; the Centerfield Limestone, Brown's Creek, York, New York; and ?the Onondaga Limestone (Southwoodian), Batavia, New York. Thus, it persists for a significant time in ENA, and passes through at least two stage boundaries without going extinct.

The existence of this species in ENA appears to have engendered some taxonomic confusion. It is figured in some of the earliest

monographic treatments of Devonian fossils from New York, such as Hall and Clarke (1888), as well as in popular accounts of trilobite fossils, such as Ludvigsen (1979). However, in all of these, as well as in Delo (1940), it has been consistently misidentified as *calliteles* (Green), with varying generic assignments. In fact, the bulk of the specimens in the collections of the NYSM that Hall and Clarke (1888) assigned to D. (C.) boothi var. calliteles belong to Bellacartwrightia jennyae.

Bellacartwrightia jennyae is closely related to B. phyllocaudata, a species that also has consistently been misidentified as calliteles. However, B. phyllocaudata and B. jennyae can be distinguished by several character states. In particular, in B. jennyae S2 is weakly declined posteriorly at its lateral edge, whereas in B. phyllocaudata S2 is roughly transverse and convex anteriorly; in B. jennyae the posterior edge of the visual surfaces is posterior to a transverse line tangent to L0, whereas in B. phyllocaudata the visual surfaces are anterior to a transverse line tangent to L0; in B. jennyae the PMI is developed as a linear incision, whereas in B. phyllocaudata it is developed as a rounded depression; in B. jennyae the genal spine extends back to the eighth thoracic segment, whereas in B. phyllocaudata the genal spine extends back to the sixth thoracic segment; in B. jennyae the posterior part of the pleural segment is elevated above the anterior part of the pleural segment, whereas in B. phyllocaudata they are of equal elevation; in B. jennyae laterally the pygidial interpleural furrows arch at the same angle as medially whereas in B. phyllocaudata they flex more strongly posteriorly; in B. jennyae there are 14 pygidial axial rings whereas in B. phyllocaudata there are 15 or 16 pygidial axial rings; in B. jennyae the terminal pygidial lappet is relatively narrow anteriorly, whereas in B. phyllocaudata it is relatively broad; in B. jennyae there is one transverse row of circular fenestrae on the anterior and posterior parts of the thoracic pleural segments, whereas these fenestrae are absent in B. phyllocaudata. None of these characters in these taxa showed any intergradation, and because these two taxa represent two diagnosably distinct units, it was of course necessary to treat

them as separate taxa. There appear to be at least two new species that are more closely related to *B. phyllocaudata* than *B. jennyae* is. In addition, there is at least one new species of *Bellacartwrightia* from the Windom Shale that is very similar to *B. jennyae* that is not described herein. However, it is figured as *B.* sp. aff. *jennyae* (fig. 13.5). It will be described in a forthcoming publication (Lieberman and Kloc, in prep.). *Bellacartwrightia jennyae* is relatively rare, but in certain units, such as the Centerfield Limestone, particularly where it outcrops near Blossom, New York, it is quite abundant.

Eldredge (1971a: pl. 14, fig. 4) figured a hypostome that is treated as belonging to this species. It is not associated with any specimens, but it is from a locality where B. jennyae is the only known asteropyginid species. It is therefore placed in that species and described. The other complete specimen Eldredge (1971a) illustrated (AMNH 36998) as G. boothi bears the hallmark characters of B. jennyae and is treated as belonging to that species. In particular, this specimen has the long pygidial lappets distally pointed, with the pleural segments prominently rounded, the spines on the medial portions of the axial rings, and the long, medially flattened genal spines.

Ludvigsen (1979) illustrated a slab from the NYSM showing two specimens that he assigned to Neometacanthus calliteles. These specimens were assigned by Hall and Clarke (1888) to D. (C.) boothi var. calliteles, but as discussed above they can no longer be placed within that species because a valid type for calliteles does not exist. A neotype for Bellacartwrightia calliteles will be erected in a subsequent publication. None of the specimens from the Hamilton Group of New York appear to be valid exponents of that species, and, indeed, Hall and Clarke's (1888) concept of calliteles included several species. The specimens Ludvigsen (1979) illustrated belong to B. jennyae. The species cannot be assigned to Neometacanthus Richter and Richter, 1948, because Bellacartwrightia is distantly related to Neometacanthus. A concept of Neometacanthus including jennyae would necessitate placing all asteropyginid species within that genus. Ludvigsen (1979) suggested that what are here named as

Greenops widderensis, new species, and B. jennyae were closely related; however, these taxa are distantly related on the basis of the phylogeny presented in figure 4 and must be treated as constituting totally independent lineages in ENA.

Several meraspid pygidia have been recovered from the Centerfield Limestone at Centerfield, New York and figured in Hall and Clarke (1888). They assigned these meraspids to D. (C.) boothi var. calliteles, and one of these is also figured in Delo (1940). Putting aside the problems with whether or not calliteles actually occurs in New York, the only species of Bellacartwrightia recovered from this horizon is B. jennyae, and therefore these meraspids are tentatively assigned to that species. Further evidence for this assignment is that the pygidial pleural lappets bear a greater resemblance to those of B. jennyae than to B. phyllocaudata or B. pleione. Being that the pygidia of B. jennyae, B. whiteleyi, new species, and B. calderonae, new species, are all quite similar, it is conceivable that these meraspids might belong to one of those taxa, but the distributional data are treated as decisive. For these reasons, these meraspids are treated as belonging to B. jennyae. In Hall and Clarke's (1888) illustrations, only five pairs of pleural lappets were shown on the pygidia; however, counts of the NYSM specimens figured (4289, 4290, and 4301) produced values that range between six and nine. To correct this inaccuracy, some of these meraspids are illustrated herein (fig. 16.2, 16.3).

Bellacartwrightia phyllocaudata, new species

Figures 10.6, 10.7, 12.1-12.4, 17, 18

Dalmanites (Cryphaeus) boothi (Green). Hall and Clarke, 1888: 44, pl. 16a, figs. 5-7.

Dalmanites (Cryphaeus) boothi var. calliteles (Green). Hall and Clarke, 1888: 45, pl. 16, figs. 16, 17, pl. 16a, fig. 17.

Cryphaeus boothi var. calliteles (Green). Grabau and Shimer, 1910: 330, fig. 1647c.

Greenops boothi (Green). Delo, 1935: 415, fig. 33; Shimer and Shrock, 1944: 647, pl. 274, fig. 12.

Greenops boothi var. calliteles (Green). Delo, 1940: 88, pl. 10, fig. 16.

Greenops (Greenops) boothi var. calliteles (Green). Ellison, 1965: pl. 18, fig. 9.

Greenops sp. Speyer, 1988: 134, fig. 10b. ?Dalmanites (Cryphaeus) boothi (Green). Hall and Clarke, 1888: 44, pl. 16, figs. 5-7. ?Dalmanites boothi (Green). Hall, 1876: pl. 16, fig. 7.

TYPES: Holotype AMNH 45230, a complete individual from the Deep Run Member (Tioughniogan), Moscow Formation, Hamilton Group, Middle Devonian, Jaycox Run Creek, upstream of Nations Road, south of Avon, Geneseo Township, Livingston County, New York, AMNH locality 3041; paratypes AMNH 45231, a pygidium; 45232, a cephalon; 45233, a cephalon; and 45241, a pygidium; all from the same locality as the holotype.

DIAGNOSIS: Maximum of nine lenses per dorsoventral file on visual surface of eye; fixigena with distinct bulge medially; cephalic posterior border furrow longest (exsag.) posterior to medial visual surface of eyes, deep laterally; S2 roughly transverse, anterior edge weakly convex; posterior edge of visual surfaces anterior to transverse line tangent to L0; visual surfaces in dorsal view present within sagittal line drawn tangent to frontal lobe; S1 medially developed as smooth curve convex posteriorly; line from posterior to anterior edge of eye forms an angle with sagittal line that is approximately 5-10°; anterior margin of frontal glabellar lobe in dorsal view smoothly rounded; PMI developed as rounded depression; spine present on L0; genal spine developed as long flange extending back to sixth thoracic segment; lateral margin of genal spine at posterior end straight; axial furrows anterior to S1 diverge more strongly, but anterior to L3 less strongly divergent; circular fenestrae on thoracic pleural segments absent; medial spine on pygidial and thoracic axial rings present; pygidial axis constricts posterior to sixth axial ring; 15 or 16 pygidial axial rings; in medial region of adjacent pygidial pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; lateral margins of pygidial lappets curved; anterior margins of pygidial axial rings posterior to constriction of pygidial axis smoothly convex anteriorly; anterior and posterior parts of pygidial pleural segments of equal elevation; laterally pygidial inter-

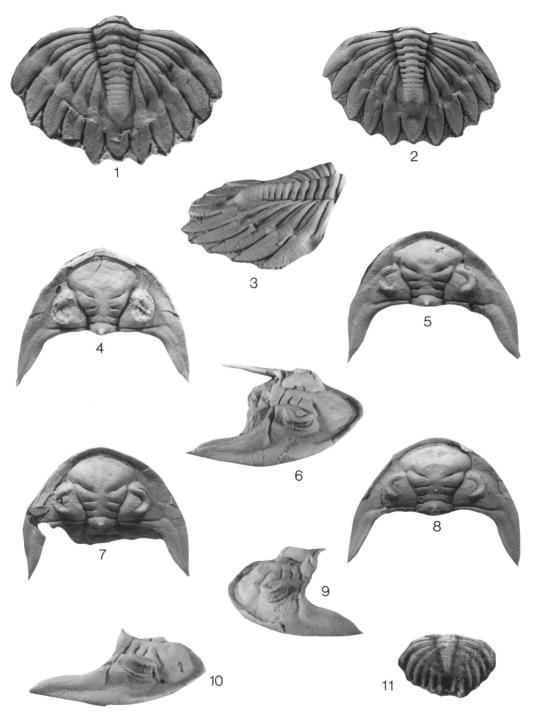


Fig. 17. Bellacartwrightia phyllocaudata, new species, Deep Run Member, Moscow Formation, Jaycox Run Creek, near Avon, Geneseo Township, Livingston County, New York. 1. AMNH 45241, dorsal view of paratype pygidium, ×2.3. 2, 3. AMNH 45231, dorsal and lateral views of paratype pygidium, ×1.5. 4. AMNH 45233, dorsal view of paratype cephalon, ×1.9. 5, 10. YPM 35808, dorsal and lateral views of cephalon, ×1.8. 6, 7. AMNH 45237, lateral and dorsal views of cephalon, ×1.5. 8, 9. AMNH 45232, dorsal and lateral views of paratype cephalon, ×1.2. 11. AMNH 45236, dorsal view of meraspid (M6) pygidium, ×7.

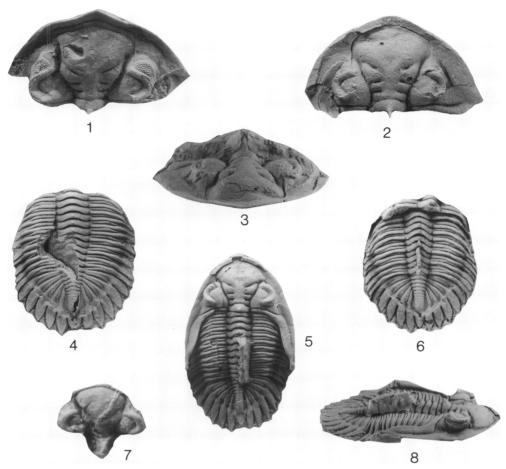


Fig. 18. Bellacartwrightia phyllocaudata, new species, Deep Run Member, Moscow Formation, Jaycox Run Creek, near Avon, Geneseo Township, Livingston County, New York. 1. AMNH 45235, dorsal view of cephalon, $\times 3$. 2, 3. AMNH 45239, dorsal and anterior views of cephalon, $\times 1.7$. 4. AMNH 45238, dorsal view of cephalothorax, $\times 1.5$. 5, 8. AMNH 45230, dorsal and lateral views of complete holotype individual, $\times 1.9$. 6. AMNH 45240, dorsal view of cephalothorax, $\times 1.6$. 7. AMNH 45234, dorsal view of small holaspid cephalon, $\times 8$.

pleural furrows flex more strongly posteriorly; terminal pygidial lappet broad.

ETYMOLOGY: This species is named by combining *phyllo* for leafy with *caudata* for tail, describing the leafy appearance of the pygidium of this species.

MATERIAL: AMNH 45230–45240, 45241–45247; NYSM 4285, 4287, 4295, and 4296; YPM 35808, ?33859.

DESCRIPTION: Same as for *Bellacartwrightia jennyae* except: cephalic length (sag.) 50% of width (tr.). Axial furrow diverging more strongly anterior to S1, diverging forward at about 45°, narrow, shallow. Glabellar length equal to 95% of width across frontal lobe;

frontal lobe ellipsoid, about 60% of length of glabella; anterior part of frontal lobe smoothly rounded in dorsal view. PMI developed as rounded depression. L3 wedge shaped, bearing isolated tubercles, lengthening distally, outer edge forms smaller angle with sagittal line at anterior end than at posterior end. S2 roughly transverse. L0 45% of width of frontal lobe. Large Eye Index 0.45. Posterior edge of eye opposite S0, anterior to transverse line tangent to L0; maximum of nine lenses per dorsoventral file on visual surface. Postocular fixigenal field 20 percent length of eye. Genal spine developed as very long flange extending back to sixth or seventh

thoracic segment; lateral margin of genal spine at posterior end straight.

Thoracic axial ring about 30% of width of thorax. Circular fenestrae absent.

Pygidial length about 60% of width without lappets. Axis with 15 or 16 rings. Anterior margins of pygidial axial rings posterior to constriction of axis smoothly convex anteriorly. Terminal pygidial lappet broad anteriorly. Postaxial region 20% of pygidial length (sag.). Anterior and posterior bands of pleural segments of equal elevation. Laterally pygidial interpleural furrows arch more strongly posteriorly.

DISCUSSION: This species is known from the following localities in the Hamilton Group in the Appalachian Basin of ENA: Windom Shale, Moscow Formation (Tioughniogan), borrow pit near Hatch's Lake, at intersection of Bradley Brook and Soule roads, 3.6 miles south of intersection of Bradlev Brook Road and New York Route 26, near Eaton, New York, AMNH locality 3031, and Earlville State Lands, New York; Deep Run Member, Moscow Formation, Jaycox Run. south of Avon, New York, AMNH locality 3041; Moscow Formation, Deep Spring Quarry; Moscow Formation, Canandaigua Lake, New York; Lower Mahantango Formation, Pennsylvania Route 191, about 4 miles north of Stroudsburg, Pennsylvania, AMNH locality 3065; "Centerfield" Member (Tioughniogan), Mahantango Formation, vicinity of Pohopoco Creek, in road cut on county road 2.5 miles east of Mahoning Valley exit of the northeast extension of the Pennsylvania Turnpike, near Forest Inn, Pennsylvania, AMNH locality 3068; Donation Member, Mahantango Formation, road cut along U.S. Route 22, opposite Huntingdon, Pennsylvania, AMNH locality 3072; upper Frame Member, borrow pit along road, south flank of Mahanoy Ridge, east from Pennsylvania Route 34, south of New Bloomfield, Pennsylvania, AMNH locality 3071; and ?the Onondaga Limestone (Southwoodian), near Buffalo, New York.

The material that Hall and Clarke (1888) assigned to *Dalmanites* (*Cryphaeus*) boothi var. calliteles (Green, 1837), as mentioned above, cannot be assigned to that species because the original types were lost, and the material Hall and Clarke (1888) used was not

from the type locality and cannot be demonstrably shown to be identical to Green's (1837) type material. However, over the years, several authors have followed Hall and Clarke (1888) and recognized a boothi var. calliteles in the Hamilton Group of New York, engendering much taxonomic confusion because the material they figured, the bulk of which is housed in the NYSM, actually belongs to several different species. The majority of the material they illustrated and collected actually belongs to B. jennyae, new species. However, some of the specimens classically treated as calliteles, i.e., those illustrated by Delo (1940) as Greenops boothi var. calliteles, and those illustrated by Grabau and Shimer (1910) as Cryphaeus boothi var. calliteles are assigned herein to a new species, B. phyllocaudata. Of three specimens Delo (1940) illustrated and assigned to Greenops boothi var. calliteles (pl. 10, figs. 13, 16, 19), one is a meraspid pvgidium that, as mentioned above, probably belongs to B. jennyae (Delo's [1940] fig. 19), another, his fig. 13, is a new species from the Kashong Shale of the Hamilton Group that will be described in a forthcoming paper (Lieberman and Kloc, in prep.), and Delo's (1940, fig. 16) can be assigned to B. phyllocaudata.

Some specimens of B. phyllocaudata are known from Huntingdon, Pennsylvania, at AMNH locality 3072, but this is not equivalent to Green's (1837) locality. When material is recovered from Green's (1837) type locality, it will be possible to designate a neotype for B. calliteles. Until that time, that species name must be avoided. Bellacartwrightia phyllocaudata and B. jennyae differ in the condition of several characters, including: the pygidial pleural lappets, which are rounded in B. jennyae and flattened in B. phyllocaudata; the greater relative length (exsag.) of the genal spines in B. jennyae; the greater relative incision of the pygidial pleural furrows in B. jennyae; the relatively broader terminal pygidial lappet in B. phyllocaudata; the anterior and posterior portions of adjacent pygidial pleural segments are of equal elevation in B. phyllocaudata, whereas in B. jennyae the posterior part is elevated above the anterior portion; and in B. phyllocaudata, there are a maximum of

nine lenses in a dorsoventral file on the visual surfaces, whereas in *B. jennyae* there are a maximum of 10 lenses in a dorsoventral file.

Although Bellacartwrightia phyllocaudata and G. boothi (Green, 1837) differ in the condition of a significant number of characters, there also has been considerable confusion regarding the identification and discrimination of specimens of B. phyllocaudata from G. boothi. However, the diagnostic characters for the two taxa presented herein may offer a way of consistently determining the identity of asteropyginid fossils in ENA.

Ellison (1965) suggested that his Greenops (G.) boothi var. calliteles—actually what is illustrated is a specimen of B. phyllocaudata—and G. boothi are sexual dimorphs because they generally appear together. This is not true for the Pennsylvania section according to the appendices in the back of Ellison (1965), and it is certainly not true for the New York section. In addition, their are numerous differences between the two taxa which are elaborated below under the heading of G. boothi. Thus, this hypothesis is highly unlikely and is not treated as valid herein.

Hall and Clarke (1888) figured some specimens that they identified as early ontogenetic stages of a species they referred to as var. calliteles, and a portion of this species is treated herein as B. phyllocaudata. However, these specimens hail from a locality in the Centerfield Limestone, from which only specimens of B. jennyae, new species, are known. Because of this, and because of certain morphological similarities in the pygidia, such as the rounded terminal lappets, these meraspids are referred to B. jennyae.

Hall and Clarke (1888) also reported the presence of *B. phyllocaudata* in the Onondaga Limestone of western New York. No material from this horizon has since been recovered, and this stratigraphic assignment for this taxon remains unsubstantiated, though not impossible, as specimens of the closely related *Bellacartwrightia pleione* (Hall) and *B. jennyae*, new species, are known from the Onondaga Limestone of the Appalachian and Illinois Basins.

Bellacartwrightia pleione (Hall, 1861) Figure 13.1-13.3

Dalmania pleione Hall, 1861: 62; Hall, 1862: 90. Dalmanites pleione (Hall). Hall, 1876: pl. 16, fig. 17.

Dalmanites (Cryphaeus) pleione (Hall). Hall and Clarke, 1888: 41, pl. 16a, fig. 2.

Greenops pleione (Hall). Delo, 1940: 90, pl. 10, fig. 20.

?Dalmanites (Cryphaeus) boothi var. calliteles (Green). Hall and Clarke, 1888: 45, pl. 16a, fig. 9.

?Dalmanites (Cryphaeus) boothi (Green). Hall and Clarke, 1888: 44, pl. 16a, fig. 4.

TYPES: As stated in Delo (1940), the holotype of the species is AMNH 4248 by monotypy, known from the Jeffersonville Limestone (Southwoodian), Falls of the Ohio River, near Louisville, Kentucky. A holotype can be designated because only a single specimen existed in the original type series of Hall (1861). This specimen is figured in Hall and Clarke (1888, pl. 16a, fig. 2) and also herein as figure 13.3.

DIAGNOSIS: Same as for genus, and: lateral margins of pygidial pleural lappets straight; pygidial axis constricts posterior to fifth axial ring; 15 pygidial axial rings; tubercle on medial portion of pygidial and thoracic axial rings; anterior margins of pygidial axial rings, posterior to constriction of pygidial axis, smoothly convex; posterior portion of pygidial pleural segments elevated above anterior portion; laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pleural segments. posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment; terminal pygidial lappet narrow; one row of circular fenestrae on anterior band of thoracic pleural segments; on posterior band, two circular fenestrae close to axis.

MATERIAL: AMNH 4248, 45252, 45253; YPM 33779-33782.

DISCUSSION: This species is known from the following stratigraphic horizon and locality in the Illinois Basin: the Jeffersonville Limestone (Southwoodian), Falls of the Ohio River, near Louisville, Kentucky. It is known from the following horizons and localities in the Appalachian Basin of ENA: ?Onondaga Limestone (Southwoodian), Leroy, New York; Windom Shale, Moscow Formation, borrow pit near Hatch's Lake, at intersection of Bradley Brook and Soule roads, 3.6 miles south of intersection of Bradley Brook Road and New York Route 26, near Eaton, New York, AMNH locality 3031; Deep Run Member (Tioughniogan), Moscow Formation, Jaycox Run, Avon, New York; the Portland Point Member, Moscow Formation, Lansing, New York; the Windom Member, Moscow Formation, Little Beard's Creek, 0.5 mile east of Trailer Park Road, Leicester, Livingston County, New York; Tully Limestone (Taghanic), Carpenter Road, 1 mile from "T" intersection with New York Route 80, south of Sheds, New York, AMNH Locality 3040; ?Frame Member (Tioughniogan), upper Mahantango Formation, roadcut along U.S. Route 22, opposite Huntingdon, Pennsylvania, AMNH locality 3072; ?Moscow Formation, Canandaigua Lake, New York; and at unspecified stratigraphic positions within the Hamilton Group at York and Ludlowville, New York. It is based exclusively on pygidial characteristics. However, there do exist isolated cephala associated with pygidia of B. pleione (YPM 33781, 33782) that are clearly asteropyginid in nature, but these are too poorly preserved to allow character analysis.

Hall and Clarke (1888) suggested that this species was very closely related to the species they treated as D. (C.) boothi var. calliteles, and that species actually appears to represent several species, none of which can be treated as Green's (1837) Cryphaeus calliteles. However, some of Hall and Clarke's (1888) material is treated herein as Bellacartwrightia phyllocaudata and B. jennyae, and these are held to be closely related to B. pleione. Hall and Clarke (1888) also commented that the adult state of B. pleione closely resembled the pygidia of what are treated here as juveniles of B. jennyae. However, H. pleione does not bear such a close resemblance to juveniles of B. jennyae, having a longer and more pronounced terminal pygidial lappet, as well as longer pleural lappets, with the fifth pleural lappet deflected the furthest posteriorly. However, again, Hall and Clarke (1888) do appear to be correct in asserting the close relationship between B. phyllocaudata, B. jennyae, and B. pleione. Bellacar-twrightia pleione could not be incorporated into the phylogenetic analysis and thus it could not be determined whether it shared a sister group relationship with the other species of Bellacartwrightia, but shared characters do suggest that they are narrowly separated branches within a common stem lineage.

Bellacartwrightia whiteleyi,

new species Figures 12.6, 19

TYPES: Holotype AMNH 45313, a complete specimen from the Murder Creek Beds, the Wanakah Shale Member (Tioughniogan), Ludlowville Formation, Hamilton Group, Middle Devonian, Darien, New York; paratypes AMNH 45314, a complete specimen, and 45315, a cephalothorax, both from the same locality as the holotype, and 45293, a small cephalon from a loose concretion somewhere in the upper Wanakah Shale Member, along the shore of Lake Erie, south of 18 Mile Creek, Hamburg, New York, and 45294, from the Wanakah Shale Member, Murder Creek Beds, Lake Erie Shore, north of 18 Mile Creek, Hamburg, New York.

DIAGNOSIS: S2 weakly declined posteriorly at lateral edge; S1 medially developed as smooth curve convex posteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; maximum of eight lenses per dorsoventral file on visual surface; anterior margin of frontal glabellar lobe in dorsal view smoothly rounded; spine on L0; PMI developed as rounded depression; line from posterior to anterior edge of eye forms 5-10° angle with sagittal line; genal spine developed as long flange extending back to sixth or seventh thoracic segment: lateral margin of genal spine at posterior end straight; axial furrows diverge more strongly anterior to S1, diverge less strongly anterior to anterior half of L3; visual surfaces in dorsal view not present within sagittal line drawn tangent to frontal lobe; two transverse rows of circular fenestrae on anterior band of thoracic pleural segments, one transverse row on posterior band; spine on medial portions of pygidial and thoracic axial rings; lateral margins of pygidial pleural lappets

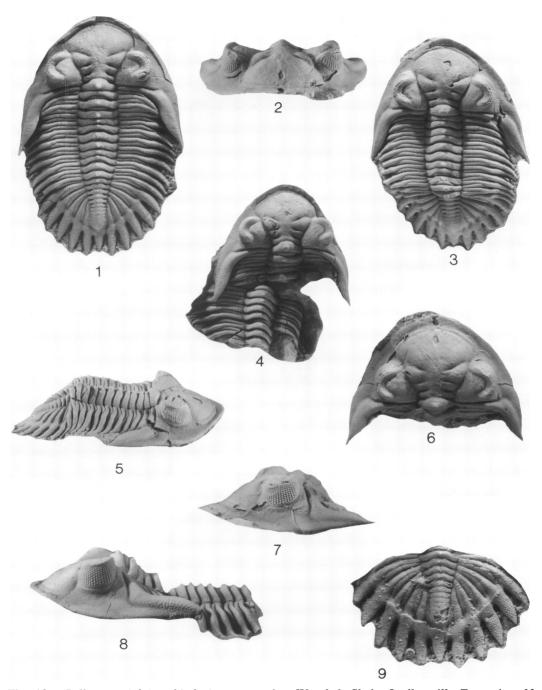


Fig. 19. Bellacartwrightia whiteleyi, new species, Wanakah Shale, Ludlowville Formation, New York. 1. Murder Creek Beds, Darien, AMNH 45314, dorsal view of complete paratype individual, ×2.5. 2, 6, 7. Upper Wanakah Shale, loose concretion along Lake Erie Shore south of 18 Mile Creek, AMNH 45293, anterior, dorsal, and lateral views of paratype cephalon, ×2.5. 3, 5. Murder Creek Beds, Darien, AMNH 45313, dorsal and lateral views of compete holotype individual, ×2. 4, 8. Murder Creek Beds, Darien, AMNH 45315, dorsal and lateral views of complete cephalon and partial thorax, ×2. 9. Murder Creek Bed, Lake Erie Shore, north of 18 Mile Creek, AMNH 45294, dorsal view of paratype pygidium, ×3.

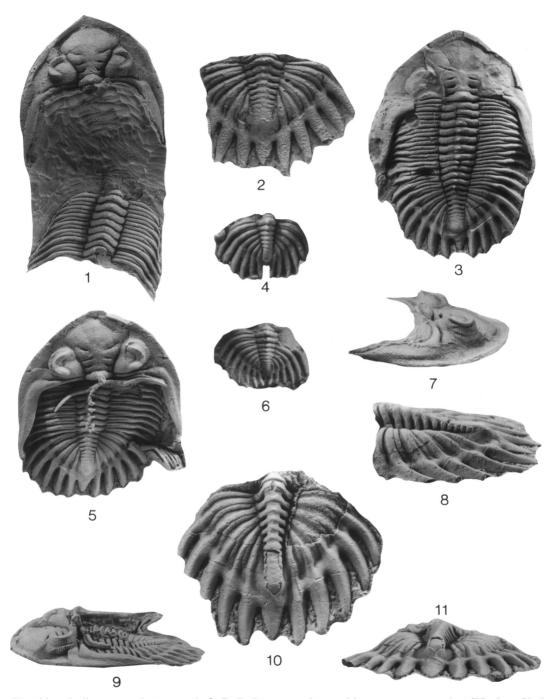


Fig. 20. Bellacartwrightia, spp. 1, 2, 7. Bellacartwrightia calderonae, new species, Windom Shale, Moscow Formation, 2–4 feet below the Taunton Beds, Kashong Glen, west side of Seneca Lake, Bellona, New York. 1. AMNH 45273, dorsal view of disarticulated cephalon and thorax of holotype individual, ×1.5. 2. AMNH 45273, dorsal view of disarticulated pygidium of holotype individual, ×2. 7. AMNH 45273, lateral view of disarticulated cephalon of holotype individual, ×1.5. 3–6, 8–11. Bellacartwrightia jennyae, new species, Centerfield Limestone, Ludlowville Formation, New York. 3. Brown's Creek, York, AMNH 45312, dorsal view of complete holotype individual, ×2.2. 4. Salt Creek Bed, Schaffer

curved; pygidial axis constricts posterior to sixth axial ring; anterior margins of pygidial axial rings posterior to constriction of pygidial axis straight; posterior part of pygidial pleural segment elevated above anterior part; laterally pygidial interpleural furrows arch at same angle as medially; in medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; 14 pygidial axial rings; terminal pygidial lappet narrow.

ETYMOLOGY: Named for T. Whiteley, Rochester, New York who found the type material used to define this species and who generously donated the material to the AMNH.

MATERIAL: AMNH 45293, 45294, 45313–45315.

DESCRIPTION: Same as for B. jennyae, except: cephalic length (sag.) 60 percent of width. Axial furrow nearly straight anterior to S1, diverging forward at about 35°, narrow, shallow; posterior to S1 diverging at 10°; diverging less strongly opposite anterior half of L3 than opposite posterior half. Frontal lobe about 60% of length of glabella. PMI developed as rounded depression. L0 45% of width of frontal lobe. Large Eye Index 0.58. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3. Maximum of eight lenses per dorsoventral file on visual surface. Fixigena with bulge medially. Genal spine developed as long flange extending back to sixth or seventh thoracic segment.

Two transverse rows of circular fenestrae on anterior band of thoracic pleural segments.

Pygidial length about 60% of width without lappets. Axis 25% of pygidial width anteriorly.

DISCUSSION: This species is known from the Murder Creek Bed, Wanakah Shale (Tioughniogan), Ludlowville Formation, Murder Creek, Darien, New York, and the same horizon from the Lake Erie Shore, New York, and below the Murder Creek Bed, Smoke Creek, Windom, New York. This species shows prominent similarity to B. jennyae, and these taxa are closely related, although not sister taxa. Bellacartwrightia jennvae precedes B. whitelevi in the fossil record, and based on phylogenetic topology, it is conceivable that B. jennyae is ancestral to B. whiteleyi, although they do differ in the condition of a number of characters such that transformation between the two species might not be perceived as "easy." Bellacartwrightia whiteleyi could in turn be ancestral to B. calderonae, which follows it in the fossil record. However, this hypothesis would require that B. phyllocaudata follow B. whiteleyi in the fossil record, and this necessitates discarding the questionable occurrences of B. phyllocaudata in the Onondaga Limestone.

In B. whiteleyi, there may be some variation in the number of lenses per dorsoventral file on the visual surface, with most specimens showing a maximum of eight lenses per dorsoventral file on the visual surface, and one specimen preserving a maximum of nine lenses per dorsoventral file on the visual surface. For the purposes of the phylogenetic analysis conducted herein, this taxon was scored as having a maximum of eight lenses per dorsoventral file.

Bellacartwrightia calderonae,

new species Figures 13.4, 20.1, 20.2

TYPES: Holotype AMNH 45273, a complete specimen from the Windom Shale (Tioughniogan), Moscow Formation, Hamilton Group, Middle Devonian, 2–4 feet below the Taunton Beds, Kashong Glen, west side of Seneca Lake, Bellona, New York.

DIAGNOSIS: S2 roughly transverse, weakly convex anteriorly; S1 medially developed as curve convex posteriorly; posterior edge of visual surfaces anterior to transverse line tangent to anterior edge of L0; maximum of

Creek, Centerfield, AMNH 45295, dorsal view of meraspid pygidium, ×9. **5**, **9**. Brown's Creek, York, AMNH 45310, dorsal and lateral views of complete paratype individual, ×2.2. **6**. Brown's Creek, York, AMNH 45296, dorsal view of meraspid pygidium, ×10. **8**, **10**, **11**. Brown's Creek, York, AMNH 45311, lateral, dorsal, and posterior views of pygidium, ×2.6.

eight lenses per dorsoventral file on visual surface; anterior margin of frontal glabellar lobe smoothly rounded; fixigena with distinct bulge medially; L0 with tubercle medially; PMI developed as rounded depression; line from posterior to anterior edge of eye forms 30° angle with sagittal line; genal spines developed as very long flanges extending back to eighth thoracic segment; lateral margin of genal spine at posterior end straight; cephalic axial furrows equally divergent anterior to S0. less strongly divergent anterior to anterior half of L3; visual surfaces in dorsal view present within sagittal line drawn tangent to frontal lobe; tubercle on medial portions of pygidial and thoracic axial rings; lateral margins of pygidial pleural lappets curving; anterior margins of pygidial axial rings posterior to seventh axial ring straight; pygidial axis constricts posterior to seventh axial ring; posterior parts of pygidial pleural segments elevated above anterior parts; laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; 14 pygidial axial rings; terminal pygidial lappet narrow; circular fenestrae on thoracic pleural segments absent.

ETYMOLOGY: Named for Mrs. Iris Calderon, Department of Invertebrates, AMNH.

MATERIAL: AMNH 29215, 45273.

DESCRIPTION: Same as for B. jennyae, except: cephalic length (sag.) 45% of width (tr.). Axial furrows nearly straight anterior to S1, diverging forward at about 35°, narrow, shallow; posterior to S1 diverging at 10°; diverging less strongly opposite anterior half of L3 than opposite posterior half. Frontal lobe about 50% of length of glabella, anterior margin smoothly rounded. PMI developed as rounded depression. LO 45% of width of frontal lobe. Large Eye Index 0.45. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3. S2 weakly declined posteriorly at lateral edges; S1 medially developed as straight line directed anteriorly. Maximum of eight lenses per dorsoventral file on visual surface; posterior edge of eye anterior to transverse line tangent to anterior edge of L0. Fixigena with bulge medially. Line from posterior to anterior edges of eye forms 5-10° angle with sagittal line. Lateral

margin of genal spine at posterior end straight. Tubercle on L0.

Tubercle on thoracic axial rings. Circular fenestrae on thoracic pleural bands and axial rings absent.

Medial tubercle on pygidial axial rings. Axis constricts posterior to seventh axial ring. Laterally pygidial interpleural furrows flex more strongly posteriorly.

DISCUSSION: This species is known from the following locality in the Appalachian Basin of ENA: Windom Shale (Tioughniogan), Moscow Formation, 2-4 feet below Taunton Beds, Kashong Glen, west side of Seneca Lake, Bellona, NY, and from an unspecified stratigraphic horizon at Kashong Glen, on the west side of Seneca Lake. This species is very similar to B. jennyae and B. whiteleyi, but these taxa have diagnosable differences and hail from different stratigraphic horizons, and thus they should be treated as separate species. In particular, the anterior border of B. calderonae is much longer (sag.) than that of the other taxa. The specimen illustrated in figure 13.4 is poorly preserved but appears referable to B. calderonae on the basis of its long, flattened anterior cephalic border and other characters that match those found in the holotype of the species. However, the pygidial pleural lappets may be slightly longer than those found in the holotype.

COLTRANEIA. NEW GENUS

TYPE SPECIES: *Treveropyge celtica* Morzadec, 1969.

Assigned Taxa: None (monotypic).

DIAGNOSIS: S3 faintly sinusoidal; cephalic posterior border furrow longest (sag.) posterior to medial visual surface of eyes; S1 deep curved pit; anterior margin of frontal glabellar lobe transverse; anterior cephalic border transverse medially, medially flexing dorsally; genal spine developed as short triangular projection; maximum of 13 lenses per dorsoventral file on visual surface; PMI developed as rounded depression; line from anterior to posterior edge of eye roughly parallels sagittal line; axial furrows diverge at constant angle; terminal pygidial lappet very reduced; lateral and medial margins of pygidial pleu-

ral lappets curved; pygidial axis constricts posterior to sixth axial ring.

ETYMOLOGY: Named for the jazz saxophonist John Coltrane.

DISCUSSION: This genus contains a single species, Coltraneia celtica (Morzadec, 1969), which had traditionally been assigned to the genus Treveropyge. However, this species differs from the type of the genus Treveropyge in the condition of the following characters: 3, 5, 7, 8, 9, 10, 11, 12, 20, 23, 31, 40, 45, 46, 50, 52, 57, 59, 60, and 63, and therefore cannot be assigned to that taxon. The new genus is sister to a clade including Bradocryphaeus Haas and Mensink, 1969, and Heliopyge Haas and Mensink, 1969, on the basis of the following shared characters: S2 weakly declined posteriorly at lateral edge; S1 crescent shaped, smooth curve convex posteriorly; medially anterior cephalic border pointed in dorsal view, not deflected far beyond lateral margins of cephalon; posterior margin of palpebrum deflected at an angle to transverse line that is significantly larger than at anterior margin; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 5-10°; significant change in cephalic topography where lateral border furrow and librigenae meet; lateral margin of genal spine forms line that departs from sagittal line by about 15-20°, at posterior end straight; interior margin of genal spine evenly deflected laterally; axial furrows less strongly divergent at anterior half of L3 relative to posterior half; no ornament on medial part of pygidial and thoracic axial rings; lateral and medial margins of pygidial lappets curved, distal tips pointed; tops of pygidial pleural segments rounded; anterolateral margins of first five pygidial axial rings evenly convex and not arching strongly anteriorly; longest pygidial lappet longer (exsag.) than length (sag.) of the first three pygidial axial rings. A monotypic genus was erected rather than placing C. celtica in the genus Bradocryphaeus or Heliopyge because the species differed in the condition of several characters diagnostic for those genera. At this point, no additional taxa can be found that fit the diagnosis of the genus Coltraneia.

Coltraneia celtica (Morzadec, 1969)

Treveropyge celtica Morzadec, 1969: 37, pl. 6, fig. 4; Morzadec, 1983: 118, pl. 5, figs. 1–10. ?Treveropyge celtica Morzadec, 1969: 38, pl. 6, fig. 5. Treveropyge sp. Morzadec, 1969: 39, pl. 6, fig. 6.

TYPES: Holotype defined in Morzadec (1983: pl. 5, fig. 1).

DIAGNOSIS: Same as for genus.

DISCUSSION: This species is known from the upper Emsian of the Armorican Massif.

BRADOCRYPHAEUS HAAS AND MENSINK, 1969

TYPE SPECIES: Cryphaeus supradevonicus Frech, 1888.

ASSIGNED TAXA: Neocalmonia (Bradocryphaeus) afghanica Haas and Mensink, 1969. DIAGNOSIS: See Haas and Mensink (1969).

DISCUSSION: The species in this genus are known from the upper Eifelian and Givetian of western Afghanistan and the Givetian and Frasnian of Belgium. On the basis of the diagnosis of Haas and Mensink (1969), as well as the characters of the species Neocalmonia (Bradocryphaeus) afghanica, which they figure, it appears that a range of taxa assigned by several authors to Bradocryphaeus can no longer be placed in that genus. In particular, Neocalmonia (Bradocryphaeus) psilus Haas, 1970, does not have the third pair of pygidial pleural lappets projecting furthest posteriorly, does not have the fifth pair of pygidial pleural lappets merging with the terminal pygidial axis, does not have the narrow terminal pygidial lappet, and does not have the prominent constriction of the pygidial axis posterior to the fifth axial ring characteristic of Bradocryphaeus; however, it does have S3 straight, the lateral edge of S2 weakly declined posteriorly, and the PMI developed as a rounded depression, also characteristic of Bradocryphaeus. At present, it is excluded from Bradocryphaeus until more detailed study of this material is possible. Gandl's (1972) Neocalmonia (Bradocryphaeus) sp. cf. mosana (Richter and Richter, 1926) has the fifth pair of pygidial pleural lappets merging with the terminal pygidial lappet, the narrow terminal pygidial lappet, and the prominent constriction of the pygidial axis posterior to the fifth axial ring; however, the

third pair of pygidial pleural lappets does not project furthest posteriorly. The cephalon of this species is too poorly preserved to permit detailed character comparisons. At present, it is also excluded from Bradocryphaeus until better material is available for study. Gandl (1972) erected a new subgenus, Neocalmonia (Quadratispina), based on the type N. (Q.) quadratispinosa, that bears several similarities to Bradocryphaeus afghanica. In the type of this subgenus, as in B. afghanica, the terminal pygidial lappet is very narrow, the fifth pair of pygidial pleural lappets does merge with the terminal pygidial lappet, and the third pair of pygidial pleural lappets projects furthest posteriorly; however, the pygidial axis constricts posterior to the sixth pygidial axis in the type species of *Quadratis*pina. This subgenus is probably very closely related to Bradocryphaeus; however, because good material could not be obtained, it was not subjected to phylogenetic analysis herein. Its phylogenetic position will be considered at a later date.

Smeenk (1983) figured and assigned several species to Bradocryphaeus. For instance, he assigned the type of Quadratispina, or Neocalmonia (Q.), Q. quadratispinosa, to Bradocryphaeus. Another one of the species he assigned to Bradocryphaeus is treated herein as Hallandclarkeops cantarmoricus. This species is not closely related to Bradocryphaeus afghanica. Instead, H. cantarmoricus is part of a larger clade that is in turn sister to a clade including Bradocryphaeus. "Bradocryphaeus" matallanensis Smeenk, 1983 lacks the narrow terminal pygidial lappet, does not have the fifth pair of pygidial pleural lappets merging with the terminal pygidial lappet, does not have the third pair of pygidial pleural lappets projecting furthest posteriorly, and does not have the prominent constriction of the pygidial axis posterior to the fifth axial ring. For these reasons it is provisionally excluded from Bradocryphaeus. "Bradocryphaeus" sexispiniferus Smeenk, 1983 is also excluded from Bradocryphaeus for similar reasons. In addition, specimens figured by Morzadec that he assigned to Bradocryphaeus, such as "B." fiacrensis (Morzadec in Babin et al., 1977) and "B." feisti Morzadec, 1992, also must be excluded from Bradocryphaeus because they

lack the characteristic morphological features of *Bradocryphaeus* stressed above.

Unfortunately, the type of the genus could not be obtained for the purposes of phylogenetic analysis. A taxon assigned to this species by the authors of the genus, in the publication where the new genus was first presented, was used instead. It is conceivable that some of the taxa that above were excluded from the genus *Bradocryphaeus* might be referable to that genus if the type of the genus was found to differ significantly from *B. afghanica*. However, this is probably unlikely.

Originally, *Bradocryphaeus* was treated as a subgenus of *Neocalmonia*. This taxonomic decision is not acceptable because these two genera are distantly related.

Several authors have commented on the phylogenetic placement of the genus Bradocryphaeus. In the case of several of these authors, doubtless their phylogenetic perspective on Bradocryphaeus is confused because the range of taxa they assign to that genus does not appear to coincide with the conception of the genus based on morphological and phylogenetic evidence presented in Haas and Mensink (1969) and herein. However, a brief overview is instructive and can be found in the discussions of Neocalmonia and Heliopyge.

Bradocryphaeus afghanica Haas and Mensink, 1969

Neocalmonia (Bradocryphaeus) afghanica Haas and Mensink, 1969: 166, pl. 1, figs. 1, 2, pl. 2, fig. 1, text fig. 4.

TYPES: Holotype figured in Haas and Mensink (1969: pl. 1, fig. 2).

DIAGNOSIS: S3 straight; lateral edge of S2 weakly declined posteriorly; maximum of seven lenses per dorsoventral file on visual surface; distance between frontal lobe and facial suture constant from lateral edges to midline; anterior cephalic border medially not deflected anteriorly beyond lateral margins of cephalon; PMI developed as rounded depression; axial furrows diverge more strongly anterior to S1 than posterior to S1; cephalic posterior border furrow straight distally; lateral to eye facial suture, before flexing posteriorly, flexes anteriorly to point op-

posite S2; anterior cephalic border in dorsal view pointed medially; fixigena with distinct bulge medially; posterior margin of palpebrum relative to anterior margin deflected at angles with transverse line that are roughly equivalent; demarcation between librigena and lateral border furrow prominent; prosopon of coarse tubercles; genal spine developed as short triangular projection; lateral margin of genal spine evenly inclined, departs from sagittal line by about 15-20°; terminal pygidial lappet triangular, very narrow; lateral and medial margins of pygidial lappets curved; distal tips of pygidial lappets pointed; posterior pleural field flanking posterior portion of pygidial axis moderately excavated; posterior part of pleural segments elevated above anterior parts; tops of pygidial pleural segments rounded; pygidial pleural furrows long (exsag.); laterally pygidial interpleural furrows arching at same angle as medially; in medial region of adjacent pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; anterolateral margins of first five pygidial axial rings do not arch strongly anteriorly; pygidial lappets long; third pygidial lappet projects furthest posteriorly.

DISCUSSION: This species is known from the upper Eifelian and Givetian of Afghanistan. It was used to epitomize the genus *Bradocryphaeus* in the phylogenetic analysis because it offered the best material of the genus that could be obtained, it matched the original diagnosis of *Bradocryphaeus*, and it was published in the work in which the taxon *Bradocryphaeus* was erected.

HELIOPYGE HAAS AND MENSINK, 1969

TYPE SPECIES: Asteropyge (Comura) helios Richter and Richter, 1926.

ASSIGNED TAXA: Neocalmonia (Heliopyge) caelata Haas and Mensink, 1969, Asteropyge (Asteropyge) hispanica Richter and Richter, 1926, Heliopyge troaonensis Morzadec, 1983, Heliopyge, new species aff. helios (Richter and Richter, 1926), Neocalmonia (Heliopyge) turolensis Gandl, 1972, and Heliopyge porsguenensis Morzadec, 1983.

DIAGNOSIS: S3 straight; S2 weakly de-

clined posteriorly at lateral edge; S1 crescent shaped, medial part developed as smooth curve, convex posteriorly; posterior edge of visual surfaces anterior to transverse line tangent to L0; lateral to eye, facial suture before flexing posteriorly flexes anteriorly to point opposite S2; distance between frontal lobe and facial suture constant from lateral edge to midline; tubercle on L0 present; anterior cephalic border not deflected far anteriorly medially; PMI developed as rounded depression; posterior margin of palpebrum relative to anterior margin deflected at angles with transverse line that are roughly equivalent; line from posterior to anterior edge of eye intersects sagittal line at roughly 0-5° angle: prosopon of fine tubercles; isolated patches of coarse tubercles present on eye ridge, L3, L2, and frontal lobe; genal spine developed as long flange extending back to sixth thoracic segment; medially genal spine developed as flattened ledge, laterally evenly inclined; medial edge of genal spine significantly thickened relative to lateral edge; axial furrows diverge more strongly anterior to S1 than posterior to S1: broad ledge of anterior cephalic border visible anterior to anterolateral positions of frontal lobe; terminal pygidial lappet teardrop shaped, narrow; lateral and medial margins of pygidial lappets curved, lappets long; anterior margins of pygidial axial rings posterior to fifth ring smoothly convex anteriorly; pygidial pleural field flanking posterior portion of axis moderately excavated; tops of pygidial pleural segments rounded; at pygidial margins, surface of pygidium dorsally inflates at contact of lappets and margins; laterally, pygidial interpleural furrows arch at same angle as medially; anterolateral margins of first five pygidial axial rings proceeding from axial furrows to midline do not arch strongly anteriorly; five pairs of pygidial pleural lappets; third pair of pygidial pleural lappets projects furthest posteriorly.

DISCUSSION: Species of this genus are known from the Givetian of Afghanistan, the Frasnian of the Cantabrian Mountains of northern Spain, the Givetian of northeastern Spain, the Frasnian of Belgium, the lower Frasnian of the Massif Armorican, and the Eifelian of the Massif Armorican. An emended diagnosis is given to that provided by

Haas and Mensink (1969) because the diagnosis they gave is inadequate to differentiate the members of this genus from other genera. Originally, Haas and Mensink (1969) considered Heliopyge to be a subgenus of Neocalmonia. Herein, these taxa are treated as two distinct genera because subgeneric classifications were not pursued and because Heliopyge and Neocalmonia are distantly related. Instead, Heliopyge is sister to Paracryphaeus, and the clade they form is in turn sister to Bradocryphaeus. Heliopyge may be closely related to Quadratispina Gandl, 1972; however, the type species of this genus (a subgenus of Neocalmonia according to Gandl [1972]) was not analyzed herein. The genus name is treated as still valid, and the relationships of this genus will be considered at a later time. An autapomorphy of Ouadratispina is the condition of the third pair of pygidial pleural lappets. The lateral and medial margins of this pair of lappets are straight, and the body of the lappets essentially parallels a sagittal line instead of being deflected laterally or medially.

In the past, a wide range of species had been assigned to Heliopyge that can no longer be assigned to it on the basis of their differences from the emended diagnosis presented herein. For example, consider the species ?Heliopyge iberica Haas, 1970, ?Heliopyge asturica Haas, 1970, ?Heliopyge coallajuensis Arbizu, 1979, ?Heliopyge nodosa Morzadec, 1983, and ?Heliopyge zornensis Morzadec, 1983. In none of these taxa does the third pygidial pleural lappet project further posteriorly than the other lappets. It is conceivable that some of these species, if subjected to phylogenetic analysis, would map out as sister to H. turolensis or other valid species of Heliopyge or form part of a stem-lineage of Heliopyge. However, such analysis was not pursued at this time, as it would require the study and incorporation of a ponderous number of taxa, none of which could be obtained. Heliopyge troaonensis Morzadec, 1983, is the oldest species of *He*liopyge. It is known only from pygidia, but it appears to be a valid species of Heliopyge. However, Morzadec's (1983, text fig. 23) illustration of the species may be inaccurate. showing the fourth pygidial pleural lappet projecting further posteriorly than the third

pleural lappet. This does not appear to be the case on the specimens Morzadec (1983) figured in his plates. If Morzadec's (1983) illustration is correct, then H. troaonensis is probably not a valid species of *Heliopyge*. Several authors have commented on the phylogenetic affinities of the genus Heliopyge. As mentioned in the section of this paper dealing with the genus Neocalmonia, Morzadec (1983) thought that Heliopyge was sister to Bradocryphaeus, with that lineage being derived from part of the lineage comprising Comura. Of course, this would make Comura paraphyletic. The paraphyly of Comura was not assessed herein. However, Morzadec (1983) appears to be correct in asserting the close relationship between Bradocryphaeus and Heliopyge, although they are not sister taxa unless Paracryphaeus Gandl, 1972, is lumped with *Heliopyge*.

Haas and Mensink (1969) and Haas (1970) suggested that *Heliopyge* was derived from a paraphyletic *Bradocryphaeus*, which was in turn derived from a paraphyletic *Asteropyge*. The validity of paraphyletic genera is not supported herein, and, in addition, *Asteropyge* appears to be distantly related to *Heliopyge* and *Bradocryphaeus*. *Heliopyge* and *Bradocryphaeus* are closely related, but *Heliopyge* is sister to *Paracryphaeus*.

Arbizu (1979) suggested that *Heliopyge* was derived from *Alcaldops*, and it in turn gave rise to *Bradocryphaeus*. *Alcaldops* was not analyzed in depth herein, but it is treated as closely related to *Tolkienia*. It is not closely related to *Bradocryphaeus* or *Heliopyge*. Similarly, Smeenk (1983) suggested that *Heliopyge* was derived from *Alcaldops*, although he claimed that it was somewhat more distantly related to *Bradocryphaeus*. Thus, the details of the phylogeny he advocated do not appear to be entirely correct.

Heliopyge turolensis was chosen for the purposes of the character analysis conducted herein because it offered the most morphologically complete material available.

Heliopyge turolensis (Gandl, 1972)

Comura (Comura) n. sp. Carls and Gandl, 1978: 463.

Neocalmonia (Heliopyge) turolensis Gandl, 1972: 142, pl. 14, figs. 4-10.

TYPES: Holotype defined in Gandl (1972: pl. 14, fig. 7).

DIAGNOSIS: Anterior cephalic border pointed in dorsal view; anterior margin of frontal glabellar lobe faintly pointed in dorsal view; 11 pygidial axial rings; medial tubercles on pygidial and thoracic axial rings absent.

DISCUSSION: This species is known from the Givetian of northeastern Spain.

PARACRYPHAEUS GANDL, 1972

TYPE SPECIES: Cryphaeus jonesi Oehlert, 1877.

ASSIGNED TAXA: Paracryphaeus hercyniae Gandl, 1972, P. sp. aff. hercyniae, P. prae-jonesi Gandl, 1972, P. alcoleae Gandl, 1972, P. caboi Smeenk, 1983, P. cf. jonesi Smeenk, 1983, and Pseudocryphaeus izensis Morzadec, 1971.

DIAGNOSIS: See Gandl (1972).

DISCUSSION: Only a single species of this genus was considered in the phylogenetic analysis. Gandl (1972) did illustrate specimens of the type of the genus, P. jonesi, and these do show some similarities with the types of P. hercyniae he illustrated. In particular, both have: the long terminal pygidial lappet; the pygidial interpleural furrows that do not flex strongly posteriorly laterally; the pygidial axis constricting posterior to the third axial ring; the anterior margins of the pygidial axial rings posterior to the third axial ring straight; and the anterior margins of the first three pygidial axial rings smoothly convex anteriorly. However, they do differ in: the condition of the pygidial pleural furrows, which are longer (exsag.) in P. jonesi than in P. hercyniae; the condition of the terminal pygidial lappet, which is long and triangular in P. jonesi but long and rectangular in P. hercyniae; the position of the fifth pair of pygidial pleural lappets, which are separated from the terminal pygidial lappet in P. jonesi but connected to the terminal lappet in P. hercyniae; the condition of the dorsal surfaces of the pygidial pleural segments, which are flat in P. jonesi but rounded in P. hercyniae; the condition of the posterior portion of the pleural segment, which is elevated above the anterior portion of the pleural segment in P. jonesi, but of equal elevation in P. hercyniae; and the condition of the pos-

terolateral margins of the pygidial pleural lappets, which are straight in P. jonesi but developed as blunt triangles in P. hercyniae. The cephala of these two taxa also differ. In particular, L1 is very reduced in P. jonesi but not in P. hercyniae; S2 is transverse in P. ionesi, but its lateral margin is directed posteriorly in P. hercyniae; and the anterior margin of the frontal glabellar lobe is faintly pointed in P. hercyniae, whereas it is smoothly rounded in P. jonesi. Based on the similarities presented above, it is conceivable that P. hercyniae and P. jonesi are closely related, and thus P. hercyniae is tentatively retained within Paracryphaeus. However, a phylogenetic analysis incorporating more complete material of P. jonesi might also suggest that these taxa are not sister taxa and thus that P. hercyniae does not belong in the genus Paracryphaeus. For now, these two species are treated as belonging to the same genus.

The phylogenetic position of Paracryphaeus is somewhat counter to that suggested by past authors. In particular, herein Paracryphaeus hercyniae is sister to Heliopyge turolensis Gandl, 1972, whereas Gandl (1972) suggested that *Paracryphaeus* was the root stock from which *Greenops* Delo, 1935, Neometacanthus Richter and Richter, 1948, Rhenops Richter and Richter, 1943, and Kayserops Delo, 1935, evolved (see fig. 5b). Smeenk (1983) argued for a relatively similar phylogenetic position for Paracryphaeus (see fig. 5d). It should be stressed that until material from the type of the genus can be obtained and incorporated into the phylogenetic analysis, the exact position of *Paracry*phaeus must remain ambiguous; however, the phylogenetic analysis of P. hercyniae suggests that it is distantly related to these genera. It is important to point out that there are several interesting convergences between species of Breizhops Morzadec, 1983, and species of *Paracryphaeus*. For instance, B. bigranulosus Morzadec, 1983, and P. hercyniae both have: the longest pygidial lappet very short, with the length (sag.) equal to the length of the first two pygidial axial rings; the anterolateral margins of the first five pygidial axial rings evenly convex; isolated patches of tubercles on L2 and L3; in the medial region of adjacent pygidial pleural

segments, the posterior region of the anterior segment is longer (exsag.) than the anterior part of the posterior segment; the tops of the pygidial pleural segments rounded; the lateral margins of the pygidial lappets curved; the terminal pygidial lappet projecting furthest posteriorly, and subrectangular; no ornament on the thoracic and pygidial axial rings; the interior margin of the genal spine evenly deflected laterally; the raised ridge on the dorsal surface of the genal spine forming a doglegged kink; the lateral margin of the genal spine at its posterior end parallels a sagittal line; the medial edge of the genal spine not significantly thickened; significant change in cephalic topography where the librigena meets the lateral border furrow; L3 flattened dorsally; anterior cephalic border medially not deflected far anteriorly beyond the lateral margins of the cephalon; the anterior cephalic border pointed in dorsal view; the distance between the frontal lobe and the facial suture at the midline relative to laterally increased slightly; S1 crescent shaped, medially developed as a smooth curve convex posteriorly; and S2 weakly declined posteriorly at its lateral edge. However, most of these shared character states are actually the primitive condition of characters, and thus they do not necessarily have relevance for phylogenetic analysis. An approach using parsimony implies that a smaller number of convergences is required to group P. hercyniae with Heliopyge turolensis than with Breizhops bigranulosus.

Important synapomorphies uniting *H. turolensis* and *P. hercyniae* include: the anteriormost part of the palpebrum reaching a point between the medial tips of L3 and the PMI; isolated patches of tubercles present on L2 and L3; a broad ledge of the anterior cephalic border visible anterior to the anterolateral portions of the frontal lobe; and the pygidial pleural furrows moderately incised, short (exsag.). Thus, at this time we suggest that the interesting similarities shared between *Breizhops bigranulosus* and *P. hercyniae* are convergences, whereas the characters uniting *H. turolensis* and *P. hercyniae* denote actual propinquity of descent.

Timm (1981) figured a series of taxa from the Lower Devonian that he assigned to *Par-acryphaeus*. He treated these taxa as evolving out of a paraphyletic Acastella. However, this taxonomic assignment appears to questionable, with Timm's (1981) figured specimens lacking characters that are found in Paracryphaeus but also that define the Asteropyginae. For instance, on those specimens where it is preserved, the genal spine is reduced to a very small nub, and the cephalic border furrow does not extend onto the genal spine and does not bifurcate it. Even in taxa with highly reduced genal spines, such as Pelitlina smeenki, new species, which is the putative sister taxon of the Asteropyginae, the cephalic border furrow extends onto the genal spine, bifurcating it. This synapomorphy characterizes both this species and all of the Asteropyginae (Edgecombe, 1991). In addition, Pelitlina and all the Asteropyginae have broad pygidial pleural lappets bordering the pygidium. In Timm's (1981) Paracryphaeus lata and P. effae, there are no pygidial pleural lappets, only small marginal denticles, as in all species of Acastava and Acastella. Thus, on the basis of these characters, these taxa cannot be treated as belonging to the Asteropyginae or Paracryphaeus, while P. hercyniae certainly possesses these characters and belongs to the ingroup Asteropyginae. Paracryphaeus jonesi, the type of Paracryphaeus, possesses the pygidial characteristics of the Asteropyginae, but the condition of the genal spines could not be determined. If P. jonesi and P. hercyniae are closely related, as predicted by this analysis, then it is most likely that P. jonesi has the cephalic border furrow bisecting the genal spine, as it does in all the other members of the asteropyginines. Thus, it is unlikely that the two aforementioned taxa that Timm (1981) illustrated belong to Paracryphaeus. More likely, they should be assigned to Acastella. Timm's (1981) P. gerrinensis does not have the cephalic posterior border furrow bifurcating the genal spine, but the condition of the pygidium bears mentioning. It does have five pairs of extremely narrow pygidial pleural lappets that do reach considerable length, sharing the condition found in *Pelitlina* and the Asteropyginae. However, it differs from P. hercyniae in the condition of several characters, including: the interpleural furrows weakly arched instead of being arched more strongly posteriorly laterally as they are in P. hercyniae; S3 sinusoidal instead of being convex anteriorly as in P. hercyniae; and the pygidial axis constricting posterior to the fifth axial ring instead of posterior to the third as in P. hercyniae. Thus, this species, like the other species figured in Timm's (1981) paper, probably should not be placed in Paracryphaeus. Overall, Timm's (1981) specimens are very poorly preserved, but it seems likely that they cannot be assigned to Paracryphaeus. At this time it is more likely that they belong to Acastella or some other taxon within a paraphyletic Acastavinae, but better material will need to be examined to ascertain the phylogenetic placement of these species with certainty.

Paracryphaeus hercyniae Gandl, 1972

Paracryphaeus hercyniae Gandl, 1972: 87, pl. 11, figs. 6-10.

Types: Holotype defined in Gandl (1972: pl. 11, fig. 7).

DIAGNOSIS: S3 convex anteriorly; cephalic posterior border furrow shallow distally and longest (sag.) posterior to palpebrum; posterior edge of visual surface posterior to transverse line tangent to L0; distance between frontal lobe and facial suture from lateral margins to midline increases slightly; PMI absent; line from posterior to anterior edge of eye parallels sagittal line; prominent demarcation between librigenae and lateral border furrow; prosopon dominated by coarse tubercles; isolated patches of coarse tubercles present on L2, L3, frontal glabellar lobe, and eye ridge; genal spine developed as moderately long flange extending back to fourth thoracic segment; lateral margin of genal spine forms line that departs from sagittal line by about 15-20°; interior margin of genal spine deflected laterally; posterolateral margins of pygidial pleural lappets developed as blunt triangles; anterior margin of pygidial axial rings posterior to pygidial axial constriction smoothly convex anteriorly; pygidial axis constricts posterior to fourth axial ring; tops of pygidial pleural lappets rounded; pygidial interpleural furrows flex strongly posteriorly laterally; nine pygidial axial rings; margin of terminal pygidial lappet rectangular, transverse posteriorly; terminal pygidial lappet very narrow, half width of posterior portion of axis; pygidial pleural lappets project from terminal lappet.

DISCUSSION: This species is known from the upper Emsian of northern Spain. Morzadec (1983) claimed that the species Paracryphaeus hercyniae Gandl, 1972, was actually closely related to B. lanceolatus, and he suggested that it be assigned to the genus Breizhops. His predictions were tested herein but appear to be incorrect. The phylogenetic analysis suggests that Paracryphaeus hercyniae is distantly related to other species of Breizhops such as B. lanceolatus Morzadec, 1983, B. bigranulosus Morzadec, 1983, and B. chaconae (Arbizu, 1979). Important differences between Paracryphaeus and Breizhops include: in P. hercyniae S3 is convex anteriorly, whereas in all species of Breizhops it is straight, with the anterolateral edge transverse; in P. hercyniae the posterior edge of the visual surfaces is posterior to a transverse line tangent to the anterior edge of L0, whereas in all species of Breizhops the posterior edge of the visual surfaces are anterior to a transverse line tangent to the anterior edge of L0; in P. hercyniae the adult prosopon is dominated by coarse tubercles, whereas in species of *Breizhops* the prosopon is dominated by fine tubercles; in P. hercyniae the genal spine is developed as a moderately long flange extending back to the fourth thoracic segment, whereas in species of Breizhops it is a long flange that extends back to the sixth thoracic segment; in P. hercyniae the lateral margin of the genal spine in its medial portion forms a line that departs from a sagittal line by about 15-20°, whereas in Breizhops the lateral margin of the genal spine parallels a sagittal line; in P. hercyniae the interior margin of the genal spine is deflected laterally at a 20° angle to a sagittal line, whereas in Breizhops the interior margin of the genal spine parallels the lateral margin of the genal spine before deflecting laterally; in P. hercyniae the medial margins of the pygidial lappets are curved, whereas in Breizhops the medial margins of the pygidial lappets are straight; in P. hercyniae the pygidial axis constricts posterior to the fourth axial ring, whereas in Breizhops the pygidial axis constricts posterior to the fifth axial ring: in P. hercyniae the anterior margins of the

pygidial axial rings posterior to the constriction of the pygidial axis are smoothly convex anteriorly, whereas in Breizhops the anterior margins of the pygidial axial rings posterior to the constriction of the pygidial axis are straight; in P. hercyniae there are nine pygidial axial rings, whereas in Breizhops there are 11 pygidial axial rings; in P. hercyniae the margin of the terminal pygidial lappet is rectangular and transverse posteriorly, whereas in Breizhops the terminal pygidial lappet is rectangular and triangular posteriorly; in P. hercyniae the terminal pygidial lappet is very narrow, 0.5 times the width (tr.) of the posterior portion of the axis, whereas in Breizhops the terminal pygidial lappet is narrow, about 1-1.1 times the width (tr.) of the posterior portion of the pygidial axis. Thus, on the basis of these differences, it is hard to support Morzadec's (1983) conclusion that P. hercyniae Gandl, 1972, should be assigned to Breizhops.

Smeenk (1983) assigned a series of cephala and pygidia from different localities, horizons, and formations to G. (G.) chaconae, and he asserted that these were conspecific with Arbizu's (1979) G. chaconae. However, the pygidia appear to belong to a completely different species. This species, Paracryphaeus sp. aff. hercyniae, is nearly identical to P. hercyniae Gandl, 1972, and they share the constriction of the pygidial axis posterior to the fourth pygidial axial ring, the very long pleural lappets, and the long pygidial terminal lappet that has rectangular margins. Although the pygidia Smeenk (1983) figured are poorly preserved, it appears that they may have one more pygidial axial ring than do those of P. hercyniae, and thus until better material can be found they are tentatively assigned to P. sp. aff. hercyniae. These pygidia also differ from those of P. hercyniae in the condition of the posterior margin of the terminal pygidial lappet. The margin is triangular in P. sp. aff. hercyniae but was coded as transverse in P. hercyniae, as that is its condition in some of the paratype material that Gandl (1972) figured, e.g., plate 10, figure 10. However, there does seem to be some variation in this character, with one of the other paratype pygidia having a terminal pygidial lappet that appears to be pointed.

The pygidia of P. sp. aff. hercyniae are

also very similar to those of *B. bigranulosus*, differing principally in being relatively narrower, in having a longer terminal pygidial lappet, in having the pygidial axis constricting posterior to the fourth axial ring instead of posterior to the fifth axial ring, and in having the posterior part of the pleural segments not elevated above the anterior part. However, these similarities must be taken as convergences, because the phylogenetic analysis suggests that *Paracryphaeus* and *Breizhops* are distantly related.

Smeenk (1983) illustrated some interesting trilobites that he assigned to Greenops; however, none of the cephala and pygidia are associated, making it impossible to ascertain whether or not they come from the same species. At least some of the cephala Smeenk (1983) illustrated do appear to be conspecific with B. chaconae, but some of the specimens Smeenk (1983) illustrated show intriguing similarities to P. sp. aff. hercyniae. These are from casts of external molds. In these specimens, the frontal glabellar lobe appears pointed instead of rounded, as it is in B. chaconae. However, in other specimens, the frontal glabellar lobe appears rounded. The frontal glabellar lobe is pointed in P. hercyniae. Other interesting features include the condition of the posterior edge of the visual surfaces, which is posterior to a transverse line tangent to L0 in P. hercyniae, whereas is anterior to such a line in P. sp. aff. hercyniae (and in B. chaconae), and the condition of S3, which is convex anteriorly in P. hercyniae and straight in B. chaconae and in the cephala Smeenk (1983) figured. It is conceivable that the pygidia and cephala Smeenk (1983) illustrated belong to a single species of trilobite, indicating that the cephalic characters largely match B. chaconae but the pygidial characters are closer to P. hercyniae. However, because of the poor state of the material, until complete articulated specimens are available, the pygidia are treated as closely related to P. hercyniae and the cephala are treated as questionably conspecific with B. chaconae.

PILLETINA HAAS, 1970

Type Species: Metacanthina oehlerti praecursor Pillet, 1958.

ASSIGNED TAXA: Metacanthina aequisulcata aequisulcata Gandl, 1972, Metacanthina aequisulcata matutina Gandl, 1972, and Asteropyge (Metacanthina) oehlerti oehlerti Pillet, 1954.

DIAGNOSIS: See Haas (1970).

DISCUSSION: Examination of figures in Pillet (1958) (the author who described and illustrated the species designated the type of Pilletina) of P. praecursor (pl. 1, figs. 2, 2a, 2b; pl. 2, fig. 2) indicates that Metacanthina aequisulcata Gandl, 1972, shares several features of the pygidium in common with P. praecursor, and therefore it is tentatively placed in the genus Pilletina, contra Gandl's (1972) assignment to Metacanthina, but following Haas's (1970) and Smeenk's (1983) assignment. In particular, P. praecursor and P. aequisulcata share the following characters: the lateral and medial margins of the pygidial pleural lappets are curved; the terminal pygidial lappet projects further posteriorly than the other pleural lappets; the pygidial axis constricts posterior to the fifth axial ring; the anterior margins of the pygidial axial rings are straight; posterior to the fifth axial ring, the tops of the pygidial pleural segments are rounded; and the pygidial interpleural furrows arch at the same angle medially and laterally. However, differences between these taxa include the length and depth of the pleural furrows. They are deeper than the interpleural furrows in P. praecursor, but of equal depth in P. aequisulcata, and they are almost as long (exsag.) as the anterior pleural band in P. praecursor but short in P. aequisulcata. In addition, the posterior portions of the pleural segments are more strongly deflected posteriorly in P. praecursor than in P. aequisulcata.

Pillet's (1954) Asteropyge (Metacanthina) oehlerti oehlerti, treated herein as Pilletina oehlerti oehlerti (Pillet, 1954), more closely resembles P. aequisulcata than the type of the genus does. In addition to the characters enumerated above, both taxa also share the form of the pygidial terminal lappet, which has parallel margins anteriorly and is triangular posteriorly, and the posterior pleural segments in both taxa are not directed as strongly posteriorly as they are in P. praecursor. Therefore, if P. oehlerti and P. aequisulcata are correctly assigned to the genus

Pilletina, they are probably more closely related to each other than either is to P. praecursor.

The only species of Pilletina available for character analysis was P. aequisulcata, and the phylogenetic position of the species actually analyzed rather than the genus is what is to be emphasized. Gandl (1972) suggested that Pilletina aequisulcata was the sister taxon of Pseudocryphaeus munieri (which he referred to as Metacanthina munieri). This does not appear to be true, and they can be distinguished by the condition of several characters, including: 6, 7, 8, 9, 10, 15, 19, 22, 23, 28, 29, 31, 32, 41, 44, 50, 51, 52, 57, 58, and 60. Haas (1970) argued for a basal phylogenetic position for Pilletina (see fig. 5a), which he held to be part of a stem lineage that led to several other genera, and this lineage was held to be sister to a lineage that had Pseudocryphaeus as the basal taxon. This is not supported herein, as *Pilletina* appears to be sister to a large clade including Breizhops, Greenops, Rhenops, and Kayserops. It is not closely related at all to Pseudocryphaeus. Morzadec (1983) suggested that Pilletina was a distinct lineage derived from a paraphyletic Pseudocryphaeus (see fig. 5e). Again, this is not supported.

Pilletina aequisulcata (Gandl, 1972)

Metacanthina aequisulcata aequisulcata Gandl, 1972: 109, pl. 8, figs. 6-8, 10, not fig. 9, not pl. 9, fig. 1.

Types: Holotype defined in Gandl (1972: pl. 8, fig. 10).

DIAGNOSIS: Anterior margin of frontal glabella lobe transverse medially; lateral margin of genal spine in medial portion forms line that departs from sagittal line by 15–20°; maximum of seven lenses per dorsoventral file on visual surface; S3 faintly sinusoidal; cephalic posterior border furrow longest (exsag.) posterior to medial visual surface of eyes; distance between frontal lobe and facial suture constant from lateral margin of frontal lobe to midline; anterior cephalic border in dorsal view pointed medially; line from anterior to posterior edge of eye intersects sagittal line at 30° angle; demarcation between librigena lateral to eye and lateral border fur-

row weak; cephalic axial furrows anterior to S1 diverge more strongly; distal tips of pygidial lappets blunt triangles; lateral and medial margins of pygidial pleural lappets curving; pygidial pleural furrows moderately incised, short (sag.); terminal pygidial lappet very broad; pygidial pleural furrows and interpleural furrows equally incised.

DISCUSSION: This species is known from the Siegenian (Pragian?) of northeastern Spain. Gandl (1972) described the species Metacanthina aequisulcata, which he divided into two subspecies, aequisulcata and matutina. These two taxa can be discriminated by several characters, including the form of the pygidial terminal lappet, the number of pygidial axial rings, and the shape of the medial margins of the pygidial pleural lappets. The subspecies aequisulcata is considered to be worthy of specific status and is treated herein as *Pilletina aequisulcata*. Some of the specimens illustrated by Gandl (1972: pl. 8, fig. 9, pl. 9, fig. 1) and assigned to M. aequisulcata aequisulcata do not belong to P. aequisulcata. They differ in the number of pygidial axial rings, 14 vs. 15, and in the shape of the terminal pygidial lappet, which is triangular, and not subrectangular with parallel sides anteriorly. Therefore, these specimens should be assigned to a new species.

This species when first described was assigned to the genus *Metacanthina* by Gandl (1972). However, on the basis of morphological characters of the types of *Pilletina* and *Metacanthina* discussed above, it appears that it should be henceforth assigned to *Pilletina*.

BREIZHOPS MORZADEC, 1983

Type Species: *Breizhops lanceolatus* Morzadec, 1983.

ASSIGNED TAXA: Breizhops bigranulosus Morzadec, 1983, and Greenops (Greenops) chaconae Arbizu, 1979.

DIAGNOSIS: S3 straight, anterolateral edge transverse; S2 weakly declined posteriorly at lateral edge; medial portion of S1 smooth curve directed anteriorly; anterior cephalic border medially does not flex dorsally beyond plane through lateral margins of frontal lobe; anterior cephalic border in dorsal view pointed medially; lateral margins of border

near midline form a 110° angle; anterior cephalic border developed as narrow lip; L3 dorsally flattened; genal spine developed as long flange extending back to sixth thoracic segment; medial edge of genal spine not significantly thickened relative to lateral edge; lateral margin of genal spine parallels sagittal line, interior margin parallels lateral margin before deflecting laterally; terminal pygidial lappet narrow, subrectangular, convex posteriorly; terminal pygidial lappet projects further posteriorly than other lappets; pygidial pleural field flanking posterior part of pygidial axis faintly excavated; fifth pygidial pleural lappet projects further posteriorly than other pleural lappets; all pleural lappets shorter (exsag.) than length (sag.) of first two pygidial axial rings; pleural furrows moderately incised, short (exsag.); at medial region of adjacent pygidial pleural segments, posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; anterolateral margins of first five pygidial axial rings from axial furrows to midline smoothly convex anteriorly.

DISCUSSION: All species in this genus are known from the upper Emsian of Armorica (the Massif Armorican and northern Spain). The genus is sister to a large clade including Philipsmithiana, new genus, Rhenops Richter and Richter, 1943, Greenops Delo, 1935, Kayserops Delo, 1935, Stummiana, new genus, and Deloops, new genus, although Morzadec (1983) suggested that Breizhops was closely related to Asteropyge Hawle and Corda, 1847. Morzadec (1983: text fig. 33) showed *Breizhops* arising out of a lineage of Comura Richter and Richter, 1926, in the upper Emsian, which also gave rise to Asteropyge later in the upper Emsian. The close relationship between Asteropyge and Breizhops is not supported by the phylogenetic analysis conducted herein. In fact, when comparing the types of the two genera, A. punctata (Steininger, 1831) and B. lanceolatus, they are found to differ in the condition of several characters listed in table 1 including: 3, 5, 6, 8, 9, 13, 15, 19, 22, 28, 34, 40, 41, 44, 45, 49, 50, 54, 55, 57, 58, 59, 60, 62, and 63. The close relationship between Comura and Breizhops is also not supported herein. Although species of Comura could not be obtained for the purposes of the phylogenetic analysis, a species of the closely related genus, *Tolkienia*, new genus (discussed above), was coded for character analysis, and was found to be distantly related to *Breizhops*.

Breizhops lanceolatus Morzadec, 1983

Breizhops lanceolatus Morzadec, 1983: 129, pl. 16, figs. 1-5.

TYPES: Holotype defined in Morzadec (1983: pl. 16, fig. 1).

DIAGNOSIS: Cephalic posterior border furrow deep laterally, longest (sag.) posterior to medial visual surfaces of eyes; posterior edge of visual surface anterior to transverse line tangent to L0: maximum of six lenses per dorsoventral file on visual surfaces; facial suture lateral to eye before flexing posteriorly flexes anteriorly to point opposite S1; anterior margin of frontal glabellar lobe faintly pointed in dorsal view; posterior margin of palpebrum relative to anterior margin of fixigena deflected at a significantly larger angle to transverse line; portion of librigena between lateral margin of eye and lateral border furrow not visible in dorsal view; broad ledge of anterior cephalic border visible anterior to anterolateral portions of frontal lobe; distal tips of pygidial lappets at distal end of posterolateral margin inclined at approximately 45° angle relative to sagittal line; pygidial axis constricts posterior to fifth axial ring; laterally, pygidial interpleural furrows arching at same angle as medially; 11 pygidial axial rings.

DISCUSSION: This species is known from the upper Emsian of the Massif Armorican. Morzadec (1983) suggested that it was very closely related to *Paracryphaeus hercyniae* Gandl, 1972, but this conclusion is not supported by the phylogenetic analysis conducted herein, and these taxa appear to be distantly related. The part of Morzadec's (1983: text fig. 33) evolutionary tree that suggested that *B. lanceolatus* is a direct ancestor of *B. bigranulosus* is correct in the exact details only if *B. lanceolatus* is paraphyletic, but the two species are sister taxa.

Breizhops chaconae (Arbizu, 1979)

Greenops (Greenops) chaconae Arbizu, 1979: 73, pl. 2, figs. 3-7.

?Greenops (Greenops) chaconae Arbizu. Smeenk, 1983: 405, pl. 10, figs. 1-3, 7, 8, pl. 25, fig. 5.

TYPES: Holotype defined in Arbizu (1979: pl. 2, fig. 3).

DIAGNOSIS: S3 straight, anterolateral edge flexes posteriorly; posterior edge of visual surface anterior to transverse line tangent to L0; lateral to eye, facial suture before flexing posteriorly flexes anteriorly to point opposite S2; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; distance between frontal lobe and facial suture from lateral edge to midline constant; anteriormost portion of palpebrum opposite PMI; PMI developed as linear incision; demarcation between librigena lateral to eye and lateral border furrow weak, prosopon dominated by fine tubercles; genal spine developed as long flange extending back to sixth thoracic segment; medial edge of genal spine developed as flattened ledge, lateral edge evenly inclined; lateral margin of genal spine parallels sagittal line; medial margins of pygidial lappets straight; pygidial axis constricts posterior to fifth axial ring; anterior margins of pygidial axial rings posterior to fifth axial ring straight; 11 pygidial axial rings; terminal pygidial lappet narrow, with sides parallel anteriorly, triangular posteriorly.

DISCUSSION: This species is known from the upper Emsian of northern Spain. As mentioned above, Smeenk (1983) figured and assigned some pygidia to G. chaconae that differ from those typical of that species (here B. chaconae) and instead appear to be very similar to pygidia of P. hercyniae and are thus referred to here as P. sp. aff. hercyniae. These taxa share the constriction of the pygidial axis posterior to the fourth axial ring as well as the length and shape of the terminal pygidial lappet and the other pleural lappets. They do differ in the number of pygidial axial rings they possess as well as possibly in the shape of the posterior margin of the terminal pygidial lappet. This margin is pointed in P. sp. aff. hercyniae, whereas it is transverse in at least some of the specimens of P. hercyniae. However, at least one pygidium of P. hercyniae illustrated by Gandl (1972: pl. 10, fig. 9) may have the posterior margin of the pygidium pointed, so there may be some variation in this character for

P. hercyniae. It could not be ascertained whether the posterior margin of this pygidium was completely preserved or partially broken. The posterior margin of the terminal pygidial lappet was treated as transverse in the phylogenetic analysis conducted herein.

Breizhops bigranulosus Morzadec, 1983

Pseudocryphaeus sp. Morzadec, 1969: 43, pl. 8, fig. 9.

Breizhops bigranulosus Morzadec, 1983: 130, pl. 17, figs. 1-11.

Types: Holotype defined in Morzadec (1983: pl. 17, fig. 2).

DIAGNOSIS: S3 straight, anterolateral edge flexes posteriorly; posterior edge of visual surfaces anterior to transverse line tangent to L0: maximum of four lenses in a dorsoventral file on the visual surface; lateral to eye, facial suture before flexing posteriorly flexes anteriorly to point opposite S1; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; tubercle on L0 present; anterior cephalic border not deflected far anteriorly beyond lateral margins of cephalon; line from posterior to anterior edge of eye forms roughly 45° angle with sagittal line; demarcation between librigenae and lateral border furrow prominent; prosopon dominated by fine tubercles; isolated patches of coarse tubercles present on L2, L3, frontal glabellar lobe, and eye ridge; genal spine developed as long flange extending back to sixth thoracic segment; lateral margin of genal spine forms a line that departs from sagittal line by about 15-20°; cephalic axial furrows anterior to S1 diverge at same angle as posterior to S1; axial furrows at anterior and posterior halves of L3 equally divergent; portion of librigena between lateral margin of eye and lateral border furrow not visible; interior margin of genal spine parallels lateral margin before deflecting laterally; terminal pygidial lappet with sides parallel anteriorly, triangular posteriorly; distal tips of pygidial lappets at distal end of straight posterolateral margin inclined at approximately 45° angle relative to sagittal line; pygidial axis constricts posterior to fifth axial ring; posterior portion of pleural segment elevated above anterior portion of pleural segment; tops of

pleural segments rounded; pygidial interpleural furrows arch at same angle laterally as medially; 11 pygidial axial rings; terminal pygidial lappet narrow.

DISCUSSION: This species is known from the upper Emsian of the Massif Armorican. As discussed above, Morzadec (1983) considered this species to be a direct descendant of *B. lanceolatus*. Phylogenetic analysis indicates that these species are closely related. However, Morzadec (1983) also recognized the close affinity between *P. hercyniae* Gandl, 1972, and *B. bigranulosus*, but this conclusion is not supported by the phylogenetic analysis conducted herein. (See the section on *Paracryphaeus hercyniae* for more elaboration.)

PHILIPSMITHIANA, NEW GENUS

TYPE SPECIES: Philipsmithiana hyfinkeli, new species.

Assigned Taxa: Philipsmithiana burtandmimiae, new species.

DIAGNOSIS: Anterior cephalic border pointed in dorsal view, developed as narrow lip; line from posterior to anterior edge of eye intersects sagittal line at 30° angle; axial furrows at anterior half of L3 relative to posterior half less strongly divergent; posterior edge of visual surface anterior to transverse line tangent to anterior edge of L0; eye rests on vertical wall of librigena; immediately before cephalic lateral border furrow meets posterior border furrow, posterior border furrow flexes anteriorly; ornamentation on L0 absent; plane between medial and lateral edges of genal spine medially flattened ledge, laterally evenly inclined; medial edge of genal spine not significantly thickened, lateral edge parallels sagittal line; interior margin parallels lateral margin before deflecting laterally; isolated patches of tubercles on L2; medial edge of genal spine not significantly thickened relative to lateral edge; lateral margin of genal spine at posterior end straight; circular fenestrae absent; pleural furrows almost as long (sag.) as anterior pleural band; 15 or 16 pygidial axial rings; prosopon of fine tubercles; pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; terminal pygidial lappet broad, margin rectangular, convex posteriorly, does not project as far posteriorly as longest pleural lappets.

ETYMOLOGY: Named for BSL's grandfather, Philip Smith, who passed away during the course of this study.

DISCUSSION: This new genus is erected to accommodate two new species that bear a close phylogenetic relationship to certain Armorican and Eastern North American taxa traditionally assigned to Greenops; however, they lack certain derived features found in these taxa while also possessing certain derived characters absent in these taxa, and for this reason they are assigned to a separate genus that is sister to a clade including Greenops, Rhenops, and Neometacanthus. Phylogenetically informative differences between members of Philipsmithiana and Greenops include the maximum number of lenses in a dorsoventral file on the visual surface, six in Greenops, seven or nine in Philipsmithiana; the condition of the anterior cephalic border, rounded in Greenops, pointed in Philipsmithiana; the genal spine, developed as a moderately long flange extending back to the fifth thoracic segment in Greenops, extending back to the sixth or seventh thoracic segment in Philipsmithiana; and the tops of the pygidial pleural segments, flat in Greenops, rounded in Philipsmithiana.

The species *P. burtandmimiae* and *P. hy-finkeli* appear to be sister taxa on the basis of the shared possession of identical states for the following characters: 13, 25, 30, 35, 45, 65, and 66.

Philipsmithiana hyfinkeli,

new species

Figures 6.1–6.5, 7.7, 7.9, 21.2, 21.5–21.9, 22.1, 22.2, 22.5, 22.6, 22.8, 22.9, 22.11

TYPES: Holotype AMNH 45259, a complete specimen from the Middle Devonian of Morocco, near Mt. Issomour, exact locality and stratigraphic position unknown; paratypes AMNH 45260, 45264, and 45265, nearly complete specimens from the same locality as the holotype.

DIAGNOSIS: S3 convex anteriorly; maximum of nine lenses per dorsoventral file on visual surfaces; lateral to eye, before facial suture flexes posteriorly, suture flexes anteriorly to point between distal tips of S1 and

S2; distance between frontal lobe and facial suture constant from midline to lateral edges of frontal lobe; fixigena with distinct bulge medially such that it parallels dorsal plane; anterior cephalic border medially not deflected far anteriorly; PMI developed as linear incision; L3 lies in flat plane; isolated patches of coarse tubercles on L3; visual surfaces in dorsal view present within sagittal line drawn tangent to lateral edge of frontal lobe; terminal pygidial lappet subrectangular.

ETYMOLOGY: The species was named for Hy Finkel, BSL's uncle.

MATERIAL: AMNH 45259-45265.

DESCRIPTION: Cephalic length (sag.) 50% of width. Axial furrow nearly straight anterior to S1, diverging forward at about 30°, narrow, shallow; posterior to S1 diverging at 10°. Cranidial anterior border short exsagittally and medially, lying in constant dorsoventral plane; pointed medially in dorsal view. Preglabellar furrow moderately incised and narrow. Cranidial anterior border lengthens slightly medially, developed as narrow lip. Facial suture anterior to eyes flexes laterally, then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length equal to width across frontal lobe; frontal lobe ellipsoid, about 60% of length of glabella; L1-L3 flat (sag.); frontal lobe at level of posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI shallow linear incision. S3 moderately incised, convex anteriorly, branches diverging at about 130°. Sagittal region of L3-L2 nearly flat in dorsal view, lateral lobes weakly declined abaxially. L3 wedge-shaped, lengthening distally, outer edge posteriorly flexes laterally, anterior end flexing less strongly laterally, with isolated patches of coarse tubercles. S2 roughly transverse, weakly convex anteriorly, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial portion developed as smooth curve directed anteriorly, contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially. depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly medially, deep. L0 55% of width of frontal lobe, moderately arched (tr.), longest

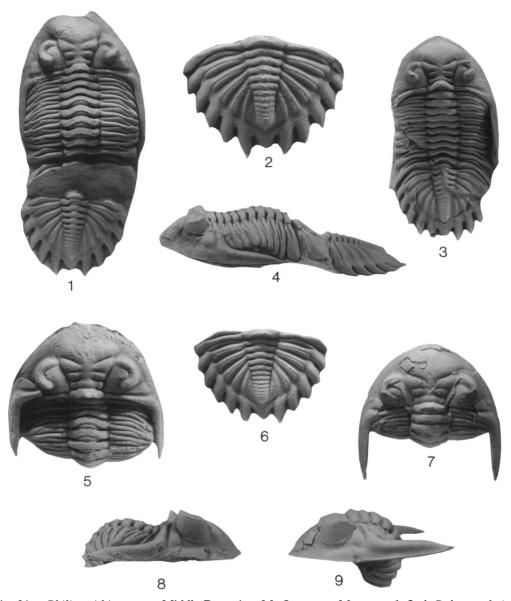


Fig. 21. Philipsmithiana spp., Middle Devonian, Mt. Issomour, Morocco. 1, 3, 4. P. burtandmimiae, new species. 1, 4. AMNH 45266, dorsal and lateral views of nearly complete holotype individual, ×1.2. 3. AMNH 45267, dorsal views of nearly complete paratype individual, ×2.1. 2, 5–9. P. hyfinkeli, new species. 2. AMNH 45264, dorsal view of pygidium of complete paratype individual, ×2. 5, 8. AMNH 45264, dorsal and lateral views of cephalon and portion of thorax of complete paratype individual, ×2. 6. AMNH 45265, dorsal view of pygidium of complete paratype individual, ×2. 7, 9. AMNH 45265, dorsal and lateral views of cephalon and portion of thorax of complete paratype individual, ×2.

sagittally. Large Eye Index 0.45. Anterior edge of eye adjacent to axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite S1, anterior to transverse line tangent to L0; eye raised slightly above fron-

tal lobe; maximum of nine lenses in a dorsoventral file on visual surface; isolated patches of coarse tubercles on eye ridge. Fixigena with distinct bulge roughly paralleling dorsoventral plane. Postocular fixigenal field

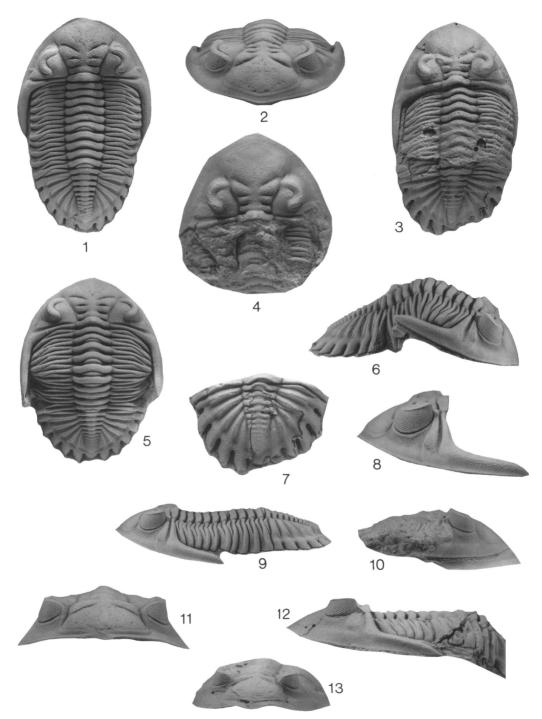


Fig. 22. Philipsmithiana spp., Middle Devonian, Mt. Issomour, Morocco. 1, 2, 5, 6, 8, 9, 11. P. hyfinkeli, new species. 1, 9, 11. AMNH 45260, dorsal, lateral, and anterior views of complete paratype individual, ×1.3. 8. AMNH 45260, lateral view of cephalon of paratype individual, ×2.1. 2, 5, 6. AMNH 45259, anterior, dorsal, and lateral views of complete holotype individual, ×1.7. 3, 4, 7, 10, 12, 13. P. burtandmimiae, new species. 3, 12, 13. AMNH 45271, dorsal, lateral, and anterior views of complete individual, ×1.6. 7. AMNH 45271, dorsal view of pygidium, ×2. 4, 10. AMNH 45272, dorsal and lateral views of cephalon, ×1.6.

30% of length of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, weakly flexing anteriorly distally, anterior margin nearly straight, of relatively constant length, longest posterior to medial visual surfaces of eyes, deep distally. Genal spine long flange extending back to sixth thoracic segment; cephalic lateral border furrow bisects spine; medial edge of genal spine flattened, lateral edge evenly inclined; lateral margin of genal spine parallels sagittal line, at posterior end recurves; interior margin of genal spine parallels lateral margin before deflecting laterally. Lateral border furrow narrow, shallow; lateral border weakly widens posteriorly. Anterior margin of cephalic doublure bluntly pointed. Prosopon of fine tubercles.

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Thorax of 11 segments. Axial ring about 25% of width of thorax; rings of equal length (exsag.) at distal ends, and medially flexed forward. Ring furrow of equal length, moderately incised, flexing forward medially and distally. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; scattered tubercles around medial portion of axial ring; articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.); outer part of pleurae more strongly flexed downward, straight in posterior segments. Posterior band long (exsag.); anterior margin straight, deflected at constant, small angle posteriorly, length (exsag.) shortens as length of pleural furrow increases medially; after approximate midpoint, length (exsag.) of pleural furrow decreases laterally, length of posterior band increases, becomes planar; posterior surface declined posterodorsally; anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially transverse, deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally.

Pygidium broadly triangular in outline excluding marginal lappets; length about 75% of width excluding lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 20° anterior to fifth pygidial axial ring, roughly parallel posterior to

fifth pygidial axial ring. Axis 25% of pygidial width anteriorly, with 15 rings; rings of nearly equal length distally and sagittally, longest distally; anterior and posterior edges of axial rings anterior to fifth axial ring from midline to distal edge weakly arch anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets straight, medial margins straight, distal tips at distal end of posterolateral margin inclined at approximately 45° angle relative to sagittal line. Axial terminus subrectangular, convex posteriorly, projecting not as far posteriorly as fifth pair of pleural lappets, broad, 1.5 times width of posterior portion of pygidial axis; postaxial region 20% of pygidial length (sag.). Pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; anterior and posterior pleural segments of equal elevation; tops of segments rounded; pleural furrows deeper than interpleural furrows, moderately incised, long (exsag.). Laterally pygidial interpleural furrows arching more strongly than medially. In medial region of adjacent pleural segments, posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known from a series of specimens from an unspecified limestone in the Middle Devonian of Morocco. It bears a strong resemblance to *Greenops boothi* from Eastern North America, but as discussed above, character analysis suggests that the taxa belong to two separate subclades that are relatively closely related but that are not sister taxa. The discovery of the new Moroccan taxa indicates that possibly a diverse new asteropyginid fauna may be recovered from these localities.

Philipsmithiana burtandmimiae, new species

Figures 7.2–7.6, 7.8, 21.1, 21.3, 21.4, 22.3, 22.4, 22.7, 22.10, 22.12, 22.13

TYPES: Holotype AMNH 45266, a complete specimen from the Middle Devonian of Morocco, near Mt. Issomour, exact locality and stratigraphic horizon unknown; paratype AMNH 45267, a nearly complete specimen from the same locality as the holotype.

DIAGNOSIS: S3 straight; anterolateral edges flexes posteriorly; maximum of seven lenses

per dorsoventral file on visual surface; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; distance between frontal lobe and facial suture increases slightly from lateral edges to midline; visual surfaces present within sagittal line drawn tangent to frontal lobe; fixigena evenly sloping from axial furrow to internal margin of eye; anterior cephalic border medially deflected anteriorly beyond lateral margins of cephalon; isolated patches of coarse tubercles on L3 absent; PMI developed as rounded depression; anterior and posterior margins of palpebrum deflected at equal but opposite angles relative to transverse line; L3 distinctly swollen; terminal pygidial lappet triangular; lateral margins of pygidial lappets curved, distal tips pointed; pygidial lappets long, length (exsag.) from proximal to distal tip approximately equals length of first three (sag.) pygidial axial rings; pygidial interpleural furrows arch at same angle laterally as medially.

ETYMOLOGY: Named for Burt and Mimi Lieberman, BSL's parents.

MATERIAL: AMNH 45266-45272.

Description: Same as for *P. hyfinkeli* except: Cephalic length (sag.) 55% of width. Distance between frontal glabellar lobe and facial suture increases slightly from lateral edges of frontal lobe to midline; anterior cephalic border medially deflected anteriorly beyond facial suture by length (sag.) of L1; S3 straight, anterolateral edge flexes posteriorly; PMI developed as rounded depression; maximum of seven lenses per dorsoventral file on visual surfaces; fixigena evenly sloping from axial furrow to internal margin of eye; posterior and anterior margins of palpebrum deflected at equal but opposite angles relative to transverse line.

Thoracic axial ring about 30% of width of thorax.

Length of pygidium about 70% of width excluding lappets; axis 30% of pygidial width anteriorly; lateral margins of pygidial lappets curved, distal tips of lappets at distal end of straight posterolateral margin inclined at 45° angle relative to sagittal line; terminal pygidial lappet triangular; pygidial interpleural furrows arch at same angle laterally and medially; pygidial lappets long (exsag.),

length (sag.) of first three axial rings; post-axial region 25% of pygidial length (sag.).

DISCUSSION: This species is also known from an unspecified limestone of Middle Devonian age in Morocco.

MODELLOPS. NEW GENUS

Type Species: *Greenops struvei* Morzadec, 1969.

Assigned Taxa: None (monotypic).

DIAGNOSIS: S3 straight, anterolateral edge flexes posteriorly; S2 roughly transverse, weakly convex anteriorly; posterior and anterior margins of palpebrum deflected at angles to transverse line that are roughly equivalent; posterior edge of visual surface anterior to transverse line tangent to anterior edge of L0: maximum of eight lenses per dorsoventral file on eye surfaces; immediately before cephalic lateral border furrow meets posterior border furrow, posterior border furrow flexes anteriorly; axial furrows diverge more strongly anterior to S1, less strongly anterior to anterior half of L3; facial suture lateral to eye, before flexing posteriorly, flexes anteriorly to point opposite S2; eye rests on sloping librigenal field; anterior cephalic border rounded in dorsal view, developed as narrow lip; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; posterior and anterior margins of fixigena deflected at equal but opposite angles relative to transverse line; ornamentation on L0 absent; lateral margin of genal spine parallels sagittal line, at posterior end straight; interior margin of genal spine deflected laterally at approximately 20° angle to sagittal line; terminal pygidial lappet subrectangular, convex posteriorly, broad, does not project as far posteriorly as lappets that project furthest posteriorly; lateral margins of pygidial lappets curved, medial margins straight; anterior margins of pygidial axial rings posterior to fifth ring smoothly convex anteriorly; tops of pygidial pleural segments rounded; pygidial pleural field flanking posterior portion of pygidial axis moderately excavated; in medial region of adjacent pleural segments, posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment; pleural furrows developed as long (exsag.) excavated space; laterally pygidial interpleural furrows arching at same angle as medially; 14 pygidial axial rings.

ETYMOLOGY: Named for Andrew Modell, Department of Invertebrates, who took the photographs used in this publication.

DISCUSSION: This monotypic genus is erected to accommodate a species sister to a clade including Asteropyge, Neometacanthus, Greenops, and Kayserops, along with several other genera. The lone species assigned to this genus differs from Philipsmithiana in the condition of several characters important to the phylogenetic analysis conducted herein, and thus the species could not be included within that genus without making it paraphyletic.

Modellops struvei (Morzadec, 1969)

Greenops? struvei Morzadec, 1969: 40, pl. 7, figs. 1-5, pl. 8, figs. 1-8.
Greenops struvei Morzadec. Morzadec, 1983: 122, pl. 8, figs. 1-11.

TYPES: Holotype defined in Morzadec (1983: pl. 8, fig. 1).

DIAGNOSIS: Same as for genus.

DISCUSSION: This species is known from the upper Emsian of the Massif Armorican. Morzadec (1969) originally assigned this species to the genus Greenops with some question because of differences he perceived between it and the type of the genus, G. boothi. In particular, he saw differences in the length of the genal spines and in the number of pygidial axial rings. However, Morzadec (1983) suggested that other species assigned to Greenops, such as G. chaconae Arbizu, 1979 (discussed above and here assigned to Breizhops Morzadec, 1983), had genal spines similar to those found in Modellops struvei. He suggested that because of this and the condition of the anterior margins of the pygidial axial rings, which laterally are straight and medially are strongly arched anteriorly, the species should be assigned to Greenops. Morzadec (1983) also suggested that it must be one of the oldest species of the genus Greenops, and he treated it as a direct ancestor of G. osismorum Morzadec. 1983 (here Stummiana osismorum). As discussed above, there are several differences between species assigned herein to Philipsmithiana and Greenops boothi, and the

same is true for the taxon assigned to Modellops. However, M. struvei does converge on G. boothi in certain aspects of its morphology. In particular, the lateral margin of the genal spine at its posterior end is straight, and the anterolateral margins of the first five pygidial axial rings laterally are straight and medially arch strongly anteriorly, but the bulk of the character evidence suggests the separation of M. struvei from Greenops. Therefore, Morzadec's (1983) contention that M. struvei is an ancestor of any species of Greenops is not upheld herein. In addition, M. struvei cannot be a direct ancestor of Stummiana osismorum as these taxa are separated by several nodes comprising different genera in the phylogeny given in figure 4.

NEOMETACANTHUS RICHTER AND RICHTER, 1948

TYPE SPECIES: *Phacops stellifer* Burmeister, 1843.

ASSIGNED TAXA: Neometacanthus sp. aff. stellifer (Burmeister, 1843), and possibly Greenops (Neometacanthus) new species Hickerson, 1992, Dalmanites (Cryphaeus) barrisi Hall and Clarke, 1888, and Asteropyge fitzpatricki Walter, 1923.

DIAGNOSIS: S3 straight; anterolateral edge flexes posteriorly; cephalic posterior border furrow longest (sag.) posterior to palpebrum, shallow and transverse laterally; S2 nearly transverse, weakly convex anteriorly; maximum of seven lenses per dorsoventral file on visual surface; anterior cephalic border pointed medially; tubercle on L0; line from posterior to anterior edge of eye roughly parallel to sagittal line; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; prosopon of fine tubercles; genal spine developed as long flange extending back at least to sixth thoracic segment; medial edge of genal spine significantly thickened relative to lateral edge; plane between medial and lateral edges of genal spine evenly inclined; cephalic axial furrows diverge more strongly anterior to S1; interior margin of genal spine parallels lateral margin before deflecting laterally; terminal pygidial lappet subrectangular anteriorly; convex posteriorly, not projecting as far posteriorly as fifth pygidial pleural lappet; lateral and medial margins of pygidial lappets curved; tops of pygidial pleural segments rounded; pleural furrows moderately incised, short (exsag.); laterally pygidial interpleural furrows flex more strongly posteriorly; posterior region of anterior pleural segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This generic name is applied to two species with surety, but may also contain three additional species. The genus Neometacanthus, based on the phylogeny presented herein, can no longer be treated as a monotypic subgenus of a larger genus Greenops (or vice versa) as certain authors had suggested in the past, e.g., Haas (1970), Gandl (1972), Smeenk (1983), and Morzadec (1983). Several authors have suggested that Neometacanthus was closely related to Greenops, and this conclusion is partially supported by the analysis conducted herein. However, the genera are not sister taxa. Instead, Neometacanthus appears to be sister to a larger clade including Rhenops Richter and Richter, 1943, and Asteropyge Hawle and Corda, 1847, on the basis of the following character states: 5(1), 22(2), 36(1), 55(1), 57(1), and 63(1). This clade is in turn sister to a clade containing Greenops Delo, 1935, Kayserops Delo, 1935, and some other new genera.

Kielan (1954) described a new species that she referred to as Asteropyge (Neometacanthus) czarnockii. Although the specimens she figured are poorly preserved (Kielan, 1954: pl. 7, figs. 6-11), they do share several characters of the pygidium with the type of the genus Asteropyge, A. punctata (Steininger, 1831), and it is therefore provisionally placed in that genus. In particular, they share the following characters: the fourth pair of pygidial pleural lappets projects further posteriorly than the fifth pair of pygidial pleural lappets; laterally the pygidial interpleural furrows arch more strongly posteriorly; the tops of the pygidial pleural segments are rounded; the pygidial terminal lappet is greatly reduced, the lateral and medial margins of the pygidial pleural lappets are straight; the distal tips of the pygidial pleural lappets are pointed; the pygidial axis constricts posterior to the fifth axial ring; the anterior margins of

the pygidial axial rings posterior to the fifth axial ring are smoothly convex anteriorly; and the pleural furrows are moderately incised and short (exsag.). However, it is important to state that some of these characters are also found in N. stellifer, and the two genera Asteropyge and Neometacanthus appear to be closely related. Because Asteropyge czarnockii shares more characters with Asteropyge punctata than with Neometacanthus stellifer, it is tentatively assigned to Asteropyge. Neometacanthus could still be viewed as a subgenus of Asteropyge, but that would also necessitate lumping the genus Rhenops within Asteropyge. As there appear to be good diagnostic characters for *Rhenops*, discussed below, this is not done, and Neometacanthus is treated as a separate genus. Asteropyge czarnockii differs from N. stellifer in the condition of several characters. In particular, the pygidial pleural lappets are relatively much longer in Kielan's (1954) species than they are in N. stellifer: the fourth pair of pygidial pleural lappets project further posteriorly than the fifth pair in A. czarnockii, whereas in N. stellifer the fifth pair of pygidial pleural lappets projects further posteriorly.

Smeenk (1983: pl. 27, fig. 2) figured a pygidium that he assigned to *Greenops* (*Neometacanthus*) sp.; however, it is too poorly preserved to assign with surety to any genus.

Authors such as Morzadec (1992) and Hickerson (1992) have remarked on the presence of the genus Neometacanthus in ENA. On the basis of the phylogenetic analysis performed herein, the sole species in North America traditionally assigned to Neometacanthus, what has traditionally been referred to as Greenops (Neometacanthus) calliteles (Green, 1837), is distantly related both to the type of Neometacanthus and to the type of the genus Greenops. Instead, that species is assigned to a new genus, Bellacartwrightia, discussed in greater detail above under the appropriate heading. However, for various reasons discussed above, the specific name calliteles can no longer be assigned to this taxon, and it is instead referred to as Bellacartwrightia phyllocaudata, new species. That species differs from N. sp. aff. stellifer in the condition of several characters, including the following listed in table 1: 1, 2, 5, 6, 7, 8, 15, 16, 23, 25, 31, 34, 38, 39, 40, 43, 46, 47, 48, 52, 56, 57, 58, 59, 62, 65, and 66

In addition, Ludvigsen (1979: fig. 57d) figured a slab showing two trilobites that he assigned to Neometacanthus calliteles (Green). These taxa are not conspecific with what is treated herein as Bellacartwrightia phyllocaudata. Instead, they belong to a new species, Bellacartwrightia jennyae, which can be distinguished both from the other species of Greenops that occur in ENA, which are discussed below, and from B. phyllocaudata on the basis of the structure of the pygidial pleural lappets, the number of pygidial axial rings, the presence of tubercles on L0 and on the thoracic and pygidial axial rings, and the condition of the genal spines, which in Bellacartwrightia jennyae have a large flattened shelf dorsally and project very far posteriorly (back to the eighth thoracic seg-

There are, however, a series of taxa from the Middle-Upper Devonian (following the stratigraphic perspective of Hickerson [1992]) of Iowa that may belong to the genus Neometacanthus. Greenops (Neometacanthus) new species Hickerson, 1992, could not be obtained because it was under study by Hickerson. The illustrations of this species given in Hickerson (1992) are too poor to assess its relationship with N. stellifer. Thus, it was impossible to ascertain whether or not this new species should be assigned to the genus Neometacanthus. It is questionably placed in that genus (but no longer treated as a subgenus of *Greenops*) until the actual type material can be examined. Hickerson (1992) also suggested that Neometacanthus barrisi (Hall and Clarke, 1888) belonged in the genus Neometacanthus. The holotype of this species is lost and no other type material is known (Delo, 1940). Again, the illustrations in Hall and Clarke (1888) and Delo (1940) are too crude to merit comment on the possible generic affinities of this asteropyginid taxon. It does appear to lack the diagnostic characters of Greenops and is not retained in that genus. However, the pygidium does have the long pleural lappets that curve laterally and medially and the well-developed terminal pygidial lappet characteristic of Neometacanthus, and it is also tentatively placed in that genus. Finally, Asteropyge fitzpatricki Walter, 1923, is another poorly preserved taxon from Iowa that was under study by Hickerson and could not be obtained in this analysis. The figures in Walter (1923) are too poor to ascertain with surety the generic affinities of this species, but like N. barrisi it has the long pygidial lappets curving laterally and medially as well as the well-developed terminal pygidial lappet. This taxon is also questionably placed in the genus Neometacanthus.

Gandl (1972) suggested that the taxa G. traversensis Stumm, 1953, and G. aequituberculatus Stumm, 1953, and a pygidium that he referred to as G. (Neometacanthus?) sp. M should be assigned to the subgenus Neometacanthus. Greenops aequituberculatus is not treated as valid herein, and instead appears to comprise a series of taxa. Greenops traversensis Stumm, 1953, and G. alpenensis Stumm, 1953, are synonymized herein and are assigned to the genus Kayserops. They are discussed in greater detail below, under the heading for that genus. The species Kayserops traversensis differs from Neometacanthus sp. aff. stellifer in the condition of several characters, including: 2, 3, 7, 8, 10, 19, 22, 24, 25, 32, 36, 40, 43, 48, 55, 56, 58, 59, 60, 61, and 62, and they are closely related. Gandl's (1972) G. (N.?) sp. M is known only from a pygidium, and thus is too incompletely preserved to ascertain its exact phylogenetic position. However, when compared with N. stellifer, it is found to differ in the condition of several characters, including: the terminal pygidial lappet, which is pointed in Gandl's species and subrectangular in N. stellifer; the pygidial lappets, which project further posteriorly than the terminal pygidial lappet in N. stellifer whereas the terminal lappet projects further posteriorly in Gandl's species; and the pygidial pleural field flanking the pygidial axis, which is faintly excavated in N. stellifer, whereas it is prominently excavated in Gandl's species. They do share similar margins of the pygidial pleural lappets, and the condition of the interpleural furrows. At this time it is probably safe to say that Gandl's sp. M is not closely related to Neometacanthus.

Neometacanthus sp. aff. stellifer (Burmeister, 1843)

Phacops stellifer Burmeister, 1843: 115, pl. 4, fig. 8.

Neometacanthus (Neometacanthus) stellifer (Burmeister). Pillet, 1961: 104, pl. 3, figs. 1-6. Greenops (Neometacanthus) stellifer (Burmeister). Struve, 1959: O480, fig. 380, 1.

DIAGNOSIS: Eleven pygidial axial rings. MATERIAL: YPM 6642.

DISCUSSION: This species is known from the Couvinian (mostly Eifelian) of Gees Gerolstein, the Eifel District, Germany; the Eifelian of the Armorican Massif in Spain and Portugal; possibly the Middle Devonian of Romania; and the Givetian of Morocco. Pillet (1961) commented on how poorly known this species is considering that it is a "classic" taxon in the history of Devonian trilobitology and on how the poor figures given by past authors do not permit discussion of the affinities of their specimens. His assessment is correct, making it difficult to verify the synonymies given by 19th century authors for lost or unavailable specimens assigned to N. stellifer. Pillet (1961) illustrated taxa from several different stratigraphic horizons and geographic localities that he assigned to N. stellifer. First it is necessary to verify that Pillet's (1961) suite of specimens do represent a single species, as he illustrated pygidia from the type locality of this species, from the Eifelian of Germany, as well as from the Givetian of Morocco. The pygidia he illustrated from the different localities do seem to be very similar, sharing the long (exsag.) and narrow (tr.) pygidial pleural lappets curving laterally and medially, the narrow terminal pygidial lappet that is convex posteriorly, the nodes on the pygidial axial rings, and the wide dished space between adjacent pleural lappets. However, the pygidia from Morocco that he figured do not have such a broad distance along the pygidial margin between the lappets as is found in pygidia from the type locality, and, in addition, the pygidial interpleural furrows flex more strongly posteriorly laterally in the Moroccan pygidia. Thus, it appears that there may be some slight differences between the German and Moroccan pygidia that Pillet (1961) assigned to the taxon N. stellifer; however, for the purposes of

this analysis, these pygidia are treated as tentatively conspecific, because the slight differences manifested between pygidia hailing from different localities would have no effect on this taxon's phylogenetic position as they do not include characters employed in phylogenetic analysis. In addition, as a wide range of specimens could not be seen, it was impossible to ascertain whether these differences represent variation within a species or differences between two distinct species. As the types of N. stellifer could not be obtained, it could not be determined with absolute certainty whether or not the specimens Pillet (1961) illustrated are conspecific with N. stellifer; therefore, they cannot be assigned to that species. However, they do appear to be very similar to illustrated specimens of N. stellifer, and they are therefore assigned to N. sp. aff. stellifer (Burmeister) until the types of that species can be obtained.

ASTEROPYGE HAWLE AND CORDA, 1847

TYPE SPECIES: Olenus punctatus Steininger, 1831.

DIAGNOSIS: See Struve (1959).

Assigned Taxa: Asteropyge (Neometacanthus) czarnockii Kielan, 1954.

DISCUSSION: Species in this genus are known from the Givetian of Poland, from the Eifelian of the Eifel District in Germany, and possibly from the Middle Devonian of Dasburg. There exist species that in the past had been assigned to Asteropyge that are not assigned to Asteropyge herein because they differ from the type species in the condition of a few characters diagnostic of the genus. These species were poorly preserved or were not available for study. Although they are not assigned to this genus at this time, the possibility exists that if incorporated into a phylogenetic analysis, they might appear in a monophyletic clade including the type of the genus Asteropyge and excluding all of the other genera analyzed herein.

As discussed above under the heading of *Neometacanthus*, Kielan (1954) figured and described the species *Asteropyge* (*Neometacanthus*) *czarnockii* from the Givetian of Poland. Although poorly preserved, this species

does appear to possess the characteristics diagnostic of the genus Asteropyge. In particular, it has the long (exsag.) pygidial pleural lappets curving laterally and medially, and it has a very reduced or absent terminal pygidial lappet, though the field that would have been occupied by this lappet is narrower transversely than it is in A. punctata (Steininger, 1831). In addition, L3 is swollen and the fourth pygidial pleural lappet projects further posteriorly than the other pleural lappets. The genus Asteropyge is very closely related to Neometacanthus, but Neometacanthus can no longer be considered a subgenus of Asteropyge unless Asteropyge were expanded to include not only Neometacanthus but also Rhenops. This is not done herein, and thus these two genera will be treated as two independent and distinct entities.

Walter (1923) figured and discussed two taxa from the Middle-Upper Devonian (following the stratigraphy of Hickerson [1992]) that he assigned to Asteropyge barrisi (Hall and Clarke, 1888) and A. fitzpatricki Walter, 1923. Unfortunately the types of species were under study by Hickerson and could not be obtained for this analysis, and the figures of Walter's (1923) specimens do not permit a detailed analysis because of their poor reproduction. Both taxa do have the long (exsag.) pygidial pleural lappets curving laterally and medially that are characteristic of Asteropyge. However, in these taxa the terminal pygidial lappet is well developed and narrow, unlike its condition in the type of Asteropyge. It is conceivable that these taxa are derived from a lineage of Asteropyge that convergently reacquired a narrow terminal pygidial lappet, or these taxa could belong to stem lineage Asteropyge and thus have retained the terminal pygidial lappet, which was later lost in more derived members of Asteropyge such as A. punctata. Until this material can be obtained and studied, assignment of a generic name to these specimens remains difficult, and at present they are questionably placed in the genus Neometacanthus, as discussed above under that generic subheading.

Morzadec and Arbizu (1978) described a new species *Kayserops cantarmoricus* that Morzadec (1983) later assigned to the genus *Asteropyge*. This assignment is not supported

by the phylogenetic analysis conducted herein as this species differs from A. punctata in the condition of several characters, including: 2, 3, 5, 6, 10, 11, 13, 14, 17, 18, 19, 23, 24, 27, 28, 29, 31, 32, 37, 40, 42, 46, 47, 48, 49, 52, 54, 57, 59, 60, 61, 62, and 66. It is henceforth assigned to the genus Halland-clarkeops, new genus, which is discussed in greater detail above. Hallandclarkeops is distantly related to Asteropyge.

In several cases, taxa that share numerous similarities with A. punctata have been assigned to the genus Heliopyge. However, there exist many differences between true Heliopyge, such as the type H. helios (Richter and Richter, 1926) and H. turolensis Gandl, 1972, and the type of Asteropyge. The type of Asteropyge does not appear to be closely related to Heliopyge in the cladogram shown in figure 4, and they differ in the condition of a significant number of character states (see tables 1, 2).

Several authors have commented on the phylogenetic relationships of the genus Asteropyge. In particular, Morzadec (1983) suggested that it was closely related to Breizhops and Comura (see fig. 5E). The close relationship between Asteropyge and Breizhops is not supported herein, as they are separated by several nodes on the cladogram given in figure 4. In regard to Comura, the type of Asteropyge is separated from a representative species of a genus closely related to Comura, such as Tolkienia granulispina (Morzadec, 1983), by several nodes on the cladogram in figure 4, and differs from it in the condition of many characters. Both Haas (1970) and Smeenk (1983) suggested that Asteropyge was closely related to *Pilletina* (see fig. 5A, D). Again, these taxa are separated by several nodes on the cladogram given in figure 4, and any similarities between these taxa must be treated as either homoplasy or symplesiomorphy. Haas (1970) also suggested that Asteropyge gave rise to the genus Neocalmonia (see fig. 5a). The type species of the genera are separated by several nodes on the single most parsimonious cladogram of the Asteropyginae and differ in the condition of several characters, including: 2, 4, 6, 7, 10, 13, 19, 22, 23, 24, 25, 27, 28, 29, 31, 32, 37, 40, 42, 43, 45, 48, 50, 54, 55, 57, 59, 61, and 64. However, they do share some similarities in their pygidial morphology. In particular, the terminal pygidial lappet is very reduced in both taxa. In addition, in A. punctata the fourth pygidial lappet projects further posteriorly than the other lappets. In N. quadricosta Pillet, 1969, the third pleural lappet projects furthest posteriorly. However, N. quadricosta only has four pleural lappets as opposed to five in A. punctata. Depending on how the pygidial pleural segments were lost developmentally and phylogenetically (following the phylogeny in fig. 4 that indicates Neocalmonia is a derived asteropyginid), the pleural lappets that project furthest posteriorly in the two taxa could be serial homologues, implying the same character state for these two taxa. Neocalmonia quadricosta was scored as having the fourth pygidial lappet projecting furthest posteriorly, assuming that its third lappet is serially homologous to the fourth lappet of all the other asteropyginines with five pleural lappets.

Asteropyge punctata (Steininger, 1831) Figures 7.1, 11

Olenus punctatus Steininger, 1831: 356, pl. 21, fig. 7.

Asteropyge punctata (Steininger). Struve, 1959: 0478, figs. 378, 379, 9a, b.

DIAGNOSIS: Broad ledge of anterior cephalic border visible anterior to anterolateral portions of frontal lobe; S3 straight; maximum of seven lenses per dorsoventral file on visual surface: distance between frontal lobe and facial suture from anterolateral edges to midline constant; immediately before cephalic posterior border furrow contacts lateral border furrow, posterior border furrow flexes anteriorly; anterior cephalic border rounded medially; tubercle on L0 absent; anterior cephalic border deflects anteriorly medially; demarcation between librigena lateral to eye and lateral border furrow weak; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; anterior margin of frontal lobe smoothly rounded in dorsal view; posterior margin of fixigena relative to anterior margin deflected at an angle to transverse line that is significantly larger; plane between medial and lateral edges of genal spine evenly inclined; lateral margin of genal spine parallels sagittal line; interior margin of genal spine parallels lateral margin before deflecting laterally; lateral and medial margins of pygidial lappets straight; pygidial pleural field flanking posterior portion of pygidial axis faint; anterior and posterior portions of pygidial pleural segments of equal elevation; fourth pygidial pleural lappet projects further posteriorly than other lappets; in medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; 15 pygidial axial rings; terminal pygidial lappet broad anteriorly; terminal pygidial lappet very reduced, developed as blunt triangle; anterolateral margins of first five pygidial axial rings when proceeding from axial furrows to midline are evenly convex and weakly arched anteriorly.

Material: AMNH 18767, 18768, 18770; YPM 6643.

DISCUSSION: This species is known from the Calceola beds, Gees, Gerolstein, the Eifel District, Germany, and Steininger (1831) also reported it from the graywacke of Dasburg. This species is another case of a "classic" asteropyginid taxon that is poorly known. We hope the additional illustrations of this material are helpful.

RHENOPS RICHTER AND RICHTER, 1943

TYPE SPECIES: Cryphaeus anserinus Richter (1916).

DIAGNOSIS: See Struve (1959).

ASSIGNED TAXA: Cryphaeus lethaeae Kayser, 1889, Rhenops redonesianus Morzadec, 1981, Rhenops babini Morzadec, 1983, Rhenops circumapodemus Smeenk, 1983, Asteropyge (Rhenops) index Richter and Richter, 1943, and Rhenops new species.

DISCUSSION: Species in this genus are known from the upper Emsian of northern Spain, the Armorican Massif, and Germany; the Eifelian of Morocco; and the Frasnian or Famennian of Venezuela. *Rhenops* is the only asteropyginid genus known to occur in Gondwana. The species from Venezuela, *Rhenops* new species, is known from rocks that may be either Frasnian or Famennian in age, though this assignment could be questionable (Carvalho, personal commun.). If recovered from the Famennian, then it would

be the youngest asteropyginid known, as all species of this subfamily were thought to have died out at the end of the Frasnian, directly following the Kellwasser anoxic event (Feist, 1991; Morzadec, 1992).

The type species of the genus was not available for analysis because the Senckenberg Museum, where the types are housed, would not allow loan of the original specimens. Instead, a species that appears to be closely related to it on the basis of illustrated material, R. babini, was used to code character information. The chief differences between that taxon and R. anserina are the condition of the terminal pygidial lappet, which projects further than the pleural lappets in R. anserina but not in R. babini, and the anterior cephalic border, which appears pointed in R. babini but it is rounded in R. anserina. The other species listed above were assigned to Rhenops because they possess the characters diagnostic of the genus. However, the possibility exists that if these taxa were subjected to phylogenetic analysis they might not map out as sister to R. babini.

Morzadec (1983) suggested that this genus was derived from the Pseudocryphaeus lineage and is sister to a clade containing Kayserops Delo, 1935, Neometacanthus Richter and Richter, 1948, and Greenops Delo, 1935 (see fig. 5e). Some of these evolutionary conclusions are supported by the analysis conducted herein, as Rhenops does appear to be closely related to Neometacanthus and it. along with Asteropyge, forms a clade sister to Neometacanthus. However, Rhenops is distantly related to both Pseudocryphaeus and Kayserops. Haas's (1970) phylogeny of the asteropyginines depicted essentially the same relationships as Morzadec's (1983) did, except he treated Kayserops as distantly related to Pseudocryphaeus and Rhenops (see fig. 5a). His phylogenetic perspective is thus also not supported, except with respect to the close affinity between Neometacanthus and Rhenops. Smeenk (1983) suggested that Rhenops was derived from the Paracryphaeus lineage (see fig. 5d). Again, this conclusion is not supported by the phylogenetic topology for the asteropyginines promulgated herein.

Rhenops babini Morzadec, 1983

Rhenops babini Morzadec, 1983: 121, pl. 7, figs. 1-7.

TYPES: Holotype defined in Morzadec (1983: pl. 7, fig. 1).

DIAGNOSIS: S3 faintly sinusoidal; lateral to eye, facial suture before flexing posteriorly flexes anterior to point opposite S1; maximum of eight lenses per dorsoventral file on visual surface; PMI developed on external mold as linear incision; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 0-5°; posterior edge of visual surface posterior to transverse line tangent to L0; demarcation between librigena and lateral border furrow prominent; genal spine developed as long flange extending back to sixth thoracic segment; medially, genal spine developed as flattened ledge, laterally evenly inclined; terminal pygidial lappet triangular; distal tips of pygidial lappets developed as blunt triangles; pygidial pleural fields flanking posterior portion of pygidial axis prominently excavated; when proceeding from axial furrows to midline, anterolateral margins of first five pygidial axial rings first straight, then arch strongly anteriorly.

DISCUSSION: This species is known from the upper Emsian of the Armorican Massif. It does appear to be somewhat closely related to Greenops, as Eldredge (1993) suggested, as it belongs to a larger clade including Asteropyge and Neometacanthus that is sister to a clade including Greenops and other North American asteropyginines such as species of Kayserops Delo, 1935, and Stummiana, new genus. Prominent similarities between species of Rhenops and Greenops include aspects of their pygidial morphology. In particular, Rhenops babini and many species of Greenops both have the distal tips of the pygidial lappets developed as blunt triangles, and both also have the anterior margins of the first five pygidial axial rings first straight and then flexing strongly anteriorly when proceeding from the axial furrows to the midline.

Rhenops, new species Figure 15

DIAGNOSIS: Distance between frontal lobe and facial suture increases slightly from lat-

eral edge of glabella to midline; librigena strongly arched; S3 faintly sinusoidal; S2 transverse, convex anteriorly; maximum of seven lenses per dorsoventral file on visual surface; anterior cephalic border pointed medially; PMI developed as linear incision; genal spine developed as long flange extending back to sixth or seventh thoracic segment; medial edge of genal spine significantly thickened relative to lateral edge; lateral margin of genal spine at posterior end recurves posteriorly; interior margin of genal spine parallels lateral margin before deflecting laterally; broad ledge of anterior cephalic border visible anterior to anterolateral portions of frontal lobe; medial tubercles on pygidial and thoracic axial rings present; terminal pygidial lappet triangular; pygidial pleural field flanking posterior portion of pygidial axis moderately excavated; lateral and medial margins of pygidial pleural lappets straight; distal tips of pygidial lappets at distal end of straight posterolateral margin inclined at 45° angle relative to sagittal line; pygidial lappets long; 14 pygidial axial rings.

MATERIAL: One internal mold of cephalon and first three thoracic axial rings missing left genal spine, three external molds of cephala, and one external mold of a pygidium, all housed in the Department of Geology, University of Rio de Janeiro, Brazil.

DISCUSSION: This species is known from strata in Venezuela that have been provisionally assigned to the Frasnian or Famennian (Carvalho, personal commun.). It is the only species of Asteropyginae known from Gondwanaland, and if the strata do turn out to be of Famennian age it is the only species of Asteropyginae to survive into the Famennian. Access to the specimens illustrated herein was provided by M. de. G. Carvalho (Department of Geology, University of Rio de Janeiro), and we thank her. This is the only species of Asteropyginae known from Gondwanaland. Oliver (1977) and Eldredge and Ormiston (1979) recognized the close relationship that northern Gondwanaland had both to ENA and to northern Africa/Armorica, and Oliver (1977) treated ENA and northern Gondwana as a single biogeographic unit, the Appohimchi Subprovince. However, their analyses were largely confined to biogeographic patterns in the Middle Devonian, and Eldredge and Ormiston (1979) did not discuss Upper Devonian trilobites. This species is part of a clade containing taxa from Armorica, suggesting that the patterns Oliver (1977) and Eldredge and Ormiston (1979) recognized for the Middle Devonian may also hold for the Upper Devonian. At least one other trilobite species from northern Gondwana (Colombia), *Monodechenella boteroi* (Caster and Richter *in* Richter and Richter, 1950), is sister to a clade of ENA species (see Lieberman, 1994).

This species is only informally diagnosed and illustrated herein. It will be fully described in a forthcoming publication by M. de G. Carvalho.

GREENOPS DELO, 1935

Type Species: Cryphaeus boothi Green, 1837.

ASSIGNED TAXA: Greenops chilmanae Stumm, 1965, G. widderensis, new species, G. barberi, new species, G. grabaui, new species, Greenops new species from the Kashong Shale, and Greenops sp. B.

DIAGNOSIS: L3 dorsally flattened; eye socle rests on vertical wall of librigena; maximum of six lenses in dorsoventral file on visual surfaces; facial suture lateral to eye, before flexing posteriorly, flexes anteriorly to point between distal tips of S1 and S2; genal spine developed as moderately long flange extending back to fifth thoracic segment; demarcation between librigena lateral to eye and lateral border furrow weak; lateral margin of genal spine parallels sagittal line; medial edge of genal spine not significantly thickened; faint or prominent raised ridge on dorsal surface of genal spine at its posterior flexure at juncture of posterior border furrow and cephalic border furrow is smoothly curved, convex distally; cephalic axial furrows equally divergent opposite margins of L3; relatively sparse circular fenestrae on posterior band of thoracic pleural segment close to thoracic axis; lateral margins of pygidial lappets curved; anterior margins of pygidial axial rings posterior to constriction of axis smoothly convex anteriorly; 11 pygidial axial rings; pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; fifth pair of pygidial pleural lappets

project furthest posteriorly; tops of pygidial pleural segments flat; pleural furrows moderately incised.

DISCUSSION: Central to the entire Greenops problem is the issue of what the actual type of the genus is. Delo (1935) erected and diagnosed the genus Greenops, and he designated Cryphaeus boothi Green as the type. However, because the original types of Green's (1837) species are lost, this taxonomic action did not serve to clear the waters. Delo (1935) figured two specimens as representative of his Greenops boothi. Unfortunately, he figured two different species. One is a new species that will be figured in an upcoming paper (Lieberman and Kloc, in prep.). It cannot be considered a valid type or neotype for the species because it hails from a very different stratigraphic horizon and geographic locale than Green's (1837) species, and because it differs significantly from Green's (1837) original material. The other (Delo [1935: fig. 32] equals Hall and Clarke [1888: pl. 16a, fig. 7]) is actually *Bel*lacartwrightia phyllocaudata, new species, which some earlier authors treated as G. calliteles. Delo's (1935) actions compounded the problems initiated by Hall and Clarke (1888), who figured a range of species from New York state that they assigned to Dalmanites (Cryphaeus) boothi. The only solution that can stabilize the taxonomic problems associated with this genus is to designate a neotype for Greenops boothi from the type locality of Green (1837) in Pennsylvania. This is done in this paper. In addition, the phylogenetic relationships of several species within the genus Greenops were evaluated using the new type concept.

A range of specimens traditionally assigned to G. boothi from New York state are discussed herein. These include some of the specimens Hall and Clarke (1888) assigned to Dalmanites (Cryphaeus) boothi. Most of these are described herein and assigned to G. barberi, new species, and G. grabaui, new species. However, there are several other new species from the Hamilton Group of New York state probably referable to Greenops that were recognized in this study. These new taxa shall be described in an upcoming paper along with asteropyginines from the Tully Limestone (Lieberman and Kloc, in prep.).

One, from the Kashong Shale, is figured herein (fig. 8.3). The purpose of the analysis presented herein was to come up with a suitable type for the genus *Greenops* and to evaluate the relative phylogenetic position of that genus within the family Asteropyginae.

In addition to designating the new genus *Greenops*, Delo (1935) placed *Greenops* in the subfamily he created, the Asteropyginae, which he in turn placed in the Dalmanitidae. However, as discussed above, the Asteropyginae can no longer be placed in the Dalmanitidae and instead should be assigned to the Acastidae (Edgecombe, 1991, 1993; Ramsköld and Edgecombe, 1993).

The genus Greenops surely represents one of the most important genera of asteropyginines, from both a phylogenetic and a biogeographic perspective. Several authors have commented on the phylogenetic position of Greenops, e.g., Haas (1970), Gandl (1972), Arbizu (1979), Smeenk (1983), and Morzadec (1983, 1992). In addition, species that past paleontologists have assigned to Greenops in the literature have been recovered from sediments in both ENA and Armorica. As discussed below, Greenops is henceforth restricted to a clade of ENA asteropyginines. Ludvigsen (1979) commented that the ancestry of Greenops lies with Early Devonian trilobites of Europe and North Africa, and this assertion appears to be correct, as discussed above and below. As discussed above, paleontologists have since the time of Delo (1935) assigned the Middle Devonian Laurentian assemblage of asteropyginines to Greenops and Greenops (Neometacanthus). However, on the basis of the phylogeny presented in figure 4, the validity of assigning all of these species to that genus is questioned. Some were recognized to instead belong to a genus distantly related to *Greenops*. referred to as Bellacartwrightia, new genus.

Many asteropyginine taxa from ENA have been figured in Hall and Clarke (1888), Delo (1940), Stumm (1953, 1965), Eldredge (1971a), Ludvigsen (1979), and Speyer (1988). Of these species, only *Greenops boothi* (Green, 1837), G. chilmanae Stumm, 1965, G. widderensis, new species, G. grabaui, new species, and G. barberi, new species, can henceforth be considered to belong in the genus Greenops. All other ENA spe-

cies, e.g., G. alpenensis Stumm, 1953, G. traversensis Stumm, 1953, G. aequituberculatus Stumm, 1953, G. arkonensis Stumm, 1953, G. pleione (Hall, 1861), G. boothi var. calliteles (Green, 1837), G. sp. A Stumm, 1953, and G. (Neometacanthus) new species Hickerson, 1992, are either synonyms of other taxa or represent species whose evolutionary histories are quite distinct from the genus Greenops, when based on a phylogenetic perspective incorporating the type species of the genus. In addition, some of the specimens figured by the authors and assigned to G. boothi are actually incorrectly identified and represent new and/or very phylogenetically distinct taxa. Other asteropyginine species from ENA that Delo (1940) assigned to Greenops, such as G. barrisi (Hall and Clarke, 1888), G. comis (Hall and Clarke, 1888), and G. kindlei Delo, 1940, could not be obtained or were too poorly preserved to be incorporated in the phylogenetic analysis. However, when possible, the evolutionary relationships of these species are discussed below under the subheadings of their probable congeners. None of these species can be retained within the genera Greenops or Neometacanthus, except possibly N. barrisi (Hall and Clarke) (discussed under the heading of Neometacanthus).

Thus, an upshot of this analysis is that what were traditionally treated as species of the genus *Greenops* in ENA actually belong to a range of phylogenetically disjunct lineages. This, in combination with the biogeographic distributions of genera in the Asteropyginae, indicates that there must have been several independent invasions of the Asteropyginae from Armorica into ENA. For instance, the species *G. grabaui*, *G. chilmanae*, *G. barberi*, and *G. widderensis* are sister to a clade including taxa from ENA and Armorica.

Stumm (1953, etc.) presented systematic analyses based on idealized views of repeated patterns of allopatric differentiation in several different clades of trilobite taxa, in which different but closely related species of each clade of Middle Devonian trilobites are found in the different tectonic basins within ENA. These views largely do appear to be upheld in the case of the genus *Greenops*. Indeed, for one of the few instances where

Stumm (1953) claimed that a species occurred in both the Appalachian and Michigan Basins, in this case, Greenops boothi, it appears that he was incorrect, and that species is restricted to the Appalachian Basin. The specimens from the Michigan Basin of southwestern Ontario that he assigned to G. boothi instead belong to G. widderensis. Greenops chilmanae and G. arkonensis, both originally described by Stumm (1953, 1965), were treated as restricted to the Traverse Group of the Michigan Basin. Herein, G. arkonensis is assigned to the new genus Stummiana and is not part of a clade of ENA Greenops; it is instead sister to a species from the Armorican Massif, while G. chilmanae is still treated as a Michigan Basin endemic.

In further support of Stumm's (1953) emphasis on allopatric differentiation in Middle Devonian trilobites of ENA, the two new taxa assigned herein to Greenops, G. barberi and G. widderensis, are known exclusively from the Appalachian and Michigan Basins, respectively. When the biogeographic states of taxa assigned to Greenops are mapped onto the phylogeny given in figure 4, a taxon from the Appalachian Basin is sister to a species endemic to the Michigan Basin. In turn, these are sister to a taxon from the Michigan Basin, implying a close relationship between the faunas of the Appalachian and Michigan Basins, also supported by Lieberman (1994) and Lieberman and Eldredge (1996).

Lieberman (1994) and Lieberman and Eldredge (1996) also concluded that the Illinois Basin was an area outside the Appalachian and Michigan Basin sister area grouping. In concordance with this, asteropyginines known from Hamilton Group equivalents (late Eifelian-Givetian), and possibly from the Frasnian of the Illinois Basin, do not appear to belong to the genus Greenops, and more likely belong to the genus Neometacanthus, which is phylogenetically distantly related to Greenops. (See discussion under that generic name and under Asteropyge.) Thus, on the whole the asteropyginines of the Michigan and Appalachian Basins are much more closely related to one another than either is to those in the Illinois Basin. Only a single species in the Asteropyginae considered herein, Bellacartwrightia pleione (Hall,

1861), occurs in both the Middle Devonian of the Illinois and Appalachian Basins. This in turn can be construed as suggesting either that different clades of Armorican taxa invaded different basins of ENA, or that there was some ecological preference that confined these clades to a single basin.

All of the species of *Greenops* are known from the Givetian portion of the Hamilton and Traverse Groups. New species probably assignable to Greenops to be described in Lieberman and Kloc (in prep.) hail from several horizons in the Givetian of the Appalachian Basin, including the Hamilton Group and the overlying Tully Limestone from the Taghanic stage. The genus Greenops is sister to a clade including one species from the late Emsian, and the clade they comprise is in turn sister to a clade including another species from the late Emsian. Therefore, the lineage leading to Greenops had probably differentiated by the late Emsian and invaded ENA some time between the late Emsian and the Givetian. Thus, there could be some gap in the history of this clade that is missing from the fossil record.

In addition to the ENA taxa discussed, certain Armorican taxa traditionally assigned to Greenops also must be excluded from that genus. In particular, Modellops struvei (Morzadec, 1969), discussed above, and Breizhops chaconae (Arbizu, 1979), also discussed above, must be excluded from Greenops because of their possession of certain characters and absence of others. Gandl's (1972: 140, pl. 8, fig. 5) Greenops (Neometacanthus?) sp. M. aff. aequituberculatus was discussed above under the heading of Neometacanthus and was found to differ from the type of that genus in the condition of several characters. Gandl's species can be distinguished from Greenops in the condition of the following characters, suggestive that it probably should not be assigned to that genus either: the pygidial pleural field flanking the posterior part of the axis is well excavated, and the anterior margins of the first five pygidial axial rings proceeding from the axial furrows to the midline are not first straight and then strongly arched anteriorly but rather are evenly convex anteriorly. However, the distal tips of the pleural lappets are blunt triangles, and this character is

found in one species of Greenops, G. grabaui; however, it is also found in Pilletina aequisulcata (Gandl, 1972). As Gandl's species does not possess some of the diagnostic characters of Greenops, it is excluded from that genus. Until more complete material of Gandl's (1972) species can be obtained, its phylogenetic position must remain ambiguous.

Smeenk (1983: pl. 11, figs. 1-6) illustrated and described a new species from the lower Frasnian of northern Spain that he referred to as G. (G.) ultimus. He claimed that it is the youngest species of *Greenops* known, and he recognized that it differed from other species of *Greenops* in the condition of several characters. He suggested that these deviations from the typical Greenops morphotype may represent adaptations by the genus in a doomed attempt to survive. He figured pygidia and cephala and treated one of the pygidia as the holotype. None of the pygidia and cephala are articulated to a common thorax, making it difficult to ascertain whether they do indeed belong to the same species. Although the material is poorly preserved and could not be obtained for phylogenetic analysis, some comment is worthwhile on the affinities of this material, for if it does indeed belong to Greenops, this would extend the stratigraphic range of that genus up into the lower Frasnian, whereas it had usually been thought to go extinct at the end of the Givetian. The pygidia do have the following traits, which are common to some species of Greenops: flattened pygidial pleural lobes; the same condition of the anterior margins of the five anteriormost pygidial axial rings; posterior flexion of the pygidial interpleural furrows laterally; curving lateral margins of the pygidial pleural lappets; anterior and posterior pleural segments of equal elevation; and pleural furrows moderately incised and short (exsag.). However, Smeenk's illustrated specimens do differ from Greenops in the condition of the anterior margins of the pygidial axial rings posterior to the constriction of the fifth axial ring; in addition, in Smeenk's (1983) G. ultimus, the terminal pygidial lappet is teardrop shaped, and this condition is not known in any other species of Greenops. Smeenk's (1983) cephalon differs from Greenops in the condition of the following salient characters: the lateral margin of the genal spine recurves medially posteriorly, and the distance between the frontal glabellar lobe and the facial suture is constant from the lateral edge of the frontal lobe to the midline; however, it resembles Greenops in some of the other characters considered in this study. This cursory analysis suggests that Smeenk's (1983) taxonomic assignment of his new species to Greenops may not be correct; however, this species must be coded into the data matrix and run on a parsimony algorithm to ascertain its phylogenetic and taxonomic position with confidence. Because of the poor state of the material and because it was not available for analysis, this cannot be done at this time but it will be pursued at a later date. Thus, contra Smeenk (1983), the stratigraphic range of the genus Greenops is at this point not extended into the lower Frasnian. Another taxon formerly assigned to Greenops that must be excluded is G. osismorum Morzadec, 1983, here Stummiana osismorum. Although this species does resemble Greenops in the condition of the facial suture lateral to the eve and in having the tops of the pygidial pleural segments flat, it differs in the condition of the following characters: in S. osismorum the posterior border furrow flexes laterally immediately before it meets the cephalic lateral border furrow, whereas in Greenops it is straight laterally; there are a maximum of seven lenses in a dorsoventral file on the visual surface in S. osismorum, whereas there are no more than six in Greenops; the genal spine extends back to the fourth thoracic segment in S. osismorum, in Greenops it extends back to the fifth segment; the genal spine is evenly inclined from its medial to lateral edges in S. osismorum, whereas in Greenops the medial edge is thickened; in S. osismorum the eye socle rests on a sloping librigenal field, whereas in Greenops it sits on a vertical wall; and in S. osismorum the lateral margin of the genal spine at its posterior end recurves posteriorly, but in *Greenops* it is straight. On the basis of these and other characters. Stummiana osismorum from the Armorican Massif is instead treated as sister to S. arkonensis (Stumm, 1953).

Stummiana arkonensis from ENA, also formerly assigned to Greenops, must be ex-

cluded from a monophyletic Greenops. It differs from Greenops in most of the characters listed above for the comparison with S. osismorum, although for some of these characters the states had to be coded as unknown for S. arkonensis. It does differ from S. osismorum in having the condition of the eye socle as in Greenops. In both, the eye socle rests on a vertical wall of the librigena. Stummiana arkonensis is another example of an asteropyginid from ENA traditionally assigned to Greenops that must be excluded on the basis of phylogenetic evidence.

Additionally, taxa such as Greenops (Greenops) sp. cf. boothi in Gandl (1972), here Armorigreenops leoi, new species, and A. sp. cf. leoi must be excluded from Greenops. Although this species bears strong superficial similarity to G. boothi, it differs in the condition of several characters utilized in the phylogenetic analysis presented herein. In particular, in A. leoi and its closely related congener, the facial suture lateral to the eye flexes anteriorly to a point opposite S1, rather than to a point between S1 and S2 as in Greenops; the genal spine extends back to the sixth or seventh thoracic segment rather than back to the fifth thoracic segment as in Greenops; the lateral margin of the genal spine at its posterior end recurves posteriorly rather than being straight as in Greenops; and the eye socle rests on a sloping librigenal field rather than on a vertical wall as in Greenops.

What was formerly *Greenops chaconae* Arbizu, 1979, also treated as a species of *Greenops* by Smeenk (1983), must be excluded from *Greenops* and is instead assigned to *Breizhops* Morzadec, 1983. The taxon differs from all species of *Greenops* in the condition of the following characters, all defining nodes of the phylogeny in figure 4: 6, 8, 26, 35, 41, 45, and 50.

Finally, one of the taxa traditionally assigned to *Greenops boothi* var. *calliteles* (here *Bellacartwrightia phyllocaudata*) plays an important part in the confusion surrounding the definition and recognition of the genus *Greenops*. A problem similar to that of *Greenops boothi*, involving lost types for this species, has engendered taxonomic confusion leading to such ramifications as a host of different species being assigned to "calliteles."

To resolve the problems fully with this species would require designation of a neotype, as was done here for G. boothi. This will be done in Lieberman and Kloc (in prep.), and it involves preparation of material from the type locality of this species in Pennsylvania. However, much of the confusion can be erased by recognizing that species closely related and/or traditionally assigned to calliteles are very distantly related to true Greenops, comprising a genus that lies in the other one of the major clades within the Asteropyginae. They differ in the condition of many of the character states given in table 1. and they were only grouped together by earlier paleontologists because of their geographic and temporal proximity. These species and this taxonomic problem are discussed extensively under the heading of Bellacartwrightia, new genus. The upshot of the phylogeny presented herein is that these taxa could not have shared a common ancestor more recently than the Siegenian (?Pragian).

Several authors have discussed the evolutionary relationships of the genus Greenops with other taxa in the Asteropyginae. Haas (1970), Gandl (1972), Arbizu (1979), Smeenk (1983), and Morzadec (1983) treated the genus as sister to *Neometacanthus* and/or treated Neometacanthus as a subgenus of Greenops (see fig. 5a-e). The phylogenetic analysis conducted herein does not suggest a close relationship between Neometacanthus and Greenops. Because of the topology of phylogenetic relationships presented in figure 4, Neometacanthus cannot be treated as a valid subgenus of *Greenops* without erecting additional subgenera and treating Greenops as a relatively broad genus. This approach is not pursued herein.

Gandl (1972) and Arbizu (1979) also suggested that *Greenops* was closely related to *Kayserops*. The type of the genus, *Kayserops kochi* (Kayser, 1883), could not be obtained for the purposes of phylogenetic analysis. However, *Kayserops inflatus* Morzadec, 1983, was analyzed and coded for characters herein, and it seems to satisfy the diagnostic characters of *Kayserops* given by Delo (1935). This species is discussed in greater detail below. However, it is clear that *Greenops* is sister to a larger clade containing *Kayserops*, but the two genera are not themselves

sister taxa. In particular, important differences between the two genera include: in Kayserops S3 is convex anteriorly, whereas in Greenops it is straight and transverse anterolaterally; in Kayserops the PMI is absent, whereas it is a rounded depression in Greenops; in Kayserops the medial edge of the genal spine is significantly thickened, whereas in Greenops it is not; in Kayserops there is a prominent tubercle developed on the medial portion of the pygidial and thoracic axial rings, whereas in Greenops there is none; in Kayserops the posterior part of the pygidial pleural segment is elevated above the anterior part, whereas in Greenops they are of equal elevation; and in Kayserops the tops of the pygidial pleural segment are rounded, whereas in Greenops they are flat.

Greenops boothi (Green, 1837)
Figure 23

Cryphaeus boothi Green, 1837: 343. ?Greenops boothi (Green). Ellison, 1965: 160, pl. 18, fig. 8

Types: Neotype YPM 35807, a complete specimen from an exposure of 30 ft of the lower Frame Shale Member, Mahantango Formation, in a road cut 450 ft long on the west side of Route 26 (Standing Stone Road) and about 600 ft northeast of the intersection of Church Street and Second Street. The neotype was collected from a zone of large concretions and was preserved in a large concretion 4 in. thick and 20 in. long. The outcrop is below Riverview Cemetery and across from the armory in the Huntingdon 7.5-minute quadrangle, on the east side of Huntingdon, Huntingdon County, Pennsylvania.

Future taxonomic work in the Asteropyginae depends on the designation of a stable neotype for the type of the genus *Greenops*, *G. boothi* (Green, 1837). However, in order to do this, several problems need to be successfully addressed. First, we must focus on how the species has been treated historically. The type of the species described and figured by Green (1837: 343, text figure) from Huntingdon, Pennsylvania has been lost for many years (Delo, 1940).

Hall and Clarke (1888) redescribed and illustrated *Dalmanites* (Cryphaeus) boothi and

D. (C.) boothi var. calliteles using material from New York. They did not illustrate or study Green's (1837) specimen. Hall and Clarke (1888) suggested that the specimens they assigned to Dalmanites (Cryphaeus) boothi, all from New York, were identical to Green's (1837) Cryphaeus boothi, though from a different locality. Delo (1940) supported their contention.

However, the conclusions of each of these authors appear to be incorrect. When we examined the material that Hall and Clarke (1888) used to redescribe D. (C.) boothi. largely housed at the NYSM, although there is also material at the AMNH, it appears that Hall and Clarke (1888) assigned several different species to D. (C.) boothi. The specimens they used to redescribe D. (C.) boothi were illustrated on plates 16 and 16a of Hall and Clarke (1888). These are referable to the following species: Greenops new species from the Kashong Shale of the Hamilton Group (Hall and Clarke, 1888: pl. 16, figs. 1, 2); G. grabaui, new species (Hall and Clarke, 1888: pl. 16, fig. 4); Bellacartwrightia new species from the Windom Shale of the Hamilton Group (Hall and Clarke, 1888: pl. 16a, figs. 4, 6); and Bellacartwrightia phyllocaudata, new species (Hall and Clarke, 1888: pl. 16a, figs. 5, 7). None are conspecific with a species of Greenops recovered from what is inferred to be Green's (1837) type locality in Pennsylvania (rationale for determination of type locality given below).

For their redescription of D. (C.) boothi var. calliteles, Hall and Clarke (1888) used specimens referable to the following species: Greenops new species from the Kashong Shale (Hall and Clarke, 1888: pl. 16, figs. 9, 14, 15); Bellacartwrightia jennyae, new species (Hall and Clarke, 1888: pl. 16, figs. 7, 8, 10-13, 19-22); and Bellacartwrightia phyllocaudata, new species (Hall and Clarke. 1888: pl. 16, figs. 16, 17). We have not yet recovered a specimen that can undoubtedly be referred to Bellacartwrightia calliteles (proposed generic assignment for that species) from Green's (1837) type locality in Pennsylvania, but it is unlikely that any of the species in New York are conspecific with it. (See sections on Bellacartwrightia, B. jennyae, and B. phyllocaudata for further discussion.)





2



3

Fig. 23. Greenops boothi (Green, 1837), lower Frame Member, Mahantango Formation, Huntingdon, Pennsylvania. 1, 2. YPM 35807, dorsal and lateral views of cephalon and part of thorax of complete neotype individual, ×2.5. 3. YPM 35807, dorsal view of pygidium and partial thorax of complete neotype individual, ×2.5.

Thus, there are two problems evident in Hall and Clarke's (1888) analysis that have engendered significant taxonomic confusion. First, each species and/or variety was actually described using several separate species, and second, there is substantial overlap in the taxa used to define the different species/varieties. This has obviously created significant problems in terms of coming up with a series of characteristics diagnostic for *G. boothi*, and, since that species is the type of the genus, this has created confusion surrounding the genus *Greenops*.

Delo (1935) proposed the name Greenops, and he designated Cryphaeus boothi Green, 1837 as the type species of the genus. Delo (1935: 415, figs. 32, 33) illustrated a pygidium and small cephalon that he assigned to his new genus Greenops and designated the type of the genus. From examination of material in the NYSM, the pygidium is that illustrated in Hall and Clarke (1888) as plate 16a, figure 7, which is NYSM 4287, and it is actually a specimen of Bellacartwrightia phyllocaudata distinguished from G. boothi and G. grabaui by its possession of a greater number of pygidial axial rings, its possession of spines on the axial rings, and its pointed terminal pygidial lappet. The small cephalon Delo (1935) illustrated as figure 33 is equivalent to Hall and Clarke's (1888) plate 16, figure 3, which is NYSM 4282. This species actually belongs to a new species of Greenops from the Kashong that will be discussed in a forthcoming work. Thus, the material on which Delo (1935) based the type species of his genus Greenops does not appear to represent true G. boothi, throwing considerable confusion around the species and the genus Greenops, and necessitating that they be properly designated here.

Delo (1940) redescribed G. boothi, and he also treated "calliteles" as a variety of that species. The illustrations in Delo (1940) are taken from Hall and Clarke (1888), and he used Greenops new species from the Kashong Shale (Delo, 1940: pl. 10, figs. 13, 15). Delo (1940) basically reinforced the decisions made in Delo (1935). In this publication he presented a redescription of G. boothi based on new material because the original types had been lost or destroyed by fire. Delo (1940) figured the material he used to redes-

cribe G. boothi. His plate 10, figure 14, is identical to the pygidium illustrated as figure 32 in Delo (1935), which is a specimen of B. phyllocaudata. Delo's (1940) plate 10, figure 15 is a dorsal view of a complete cephalothorax. His illustration is equivalent to Hall and Clarke's (1888) plate 16, figures 1 and 2, which are NYSM 4281. This specimen has characters differing from true G. boothi. In particular, it has axial nodes on the thoracic axial rings and L0, there are a maximum of seven lenses in a dorsoventral file on the visual surface, S3 is sinusoidal, and the terminal pygidial lappet is pointed posteriorly. These characters are the same as those found in a new species of Greenops from the Kashong, and thus the material Delo (1940) used to redescribe Greenops boothi represents two species, neither of which can be treated as true G. boothi. Delo's (1940) figure 17, which he also treated as G. boothi, appears to represent the pygidium of another new species from the Windom Shale of the Moscow Formation, and this species is most likely referable to the genus Bellacartwrightia, new genus. It shall also be described in a forthcoming paper (Lieberman and Kloc, in prep.).

Later, Stumm (1953) used what is treated here as G. widderensis, new species, from the Hamilton Group of southwestern Ontario to describe G. boothi. As elaborated in the discussion sections for this species and G. widderensis, these two taxa cannot be treated as conspecific. Additional misidentifications of G. boothi since the publication of Stumm (1953) are elaborated in the discussion section for this species below.

From this analysis it is apparent that there is significant confusion in the literature over the identity of *Greenops boothi*, the type of the genus *Greenops*. It is essential to establish nomenclatorial stability for the genus and the species, which are considered "important" taxa within the subfamily Asteropyginae. In the case of *Greenops boothi*, no holotype, lectotype, or syntype exists. To establish such stability it is necessary that we designate a neotype for *G. boothi*. This work is clearly "revisory" in nature, as it is a critical study of the Asteropyginae, particularly those in ENA. In addition, there has been significant confusion in the identification and

recognition of different species of Asteropyginae in the Middle Devonian of ENA, partially engendered by the absence of types for most of the species studied in this family and by the fact that the Eastern North American exponents of this subfamily have not been thoroughly revised in over 50 years nor have they been illustrated as photographs in taxonomic studies. Thus, this case meets all of the requirements for the erection of a neotype. Following Article 75 of the Code of Zoological Nomenclature, the taxonomic strategy employed in this case is the designation of a neotype for the purposes of nomenclatorial stability. A series of diagnostic characters requisite for the identification of this species is presented below in the diagnosis of Greenops boothi.

The Code of Zoological Nomenclature states that if the types are lost, attempts should be made to designate the neotype from the type locality. Green's (1837) original type locality was in the vicinity of Huntingdon, Pennsylvania. Any attempt to select a neotype from Hall and Clarke's (1888) material in geographically distant New York State would be unacceptable without assiduously searching for material in the Huntingdon region, even though so many workers have referred to the New York material in subsequent analyses of *G. boothi*.

To try to reconstruct the exact position of Green's (1837) locality, we referred to Ellison's (1965) excellent analysis of the Devonian stratigraphy of Pennsylvania. Asteropyginines have only been recovered in Pennsylvania from the Mahantango Formation (Middle Devonian, Tioughniogan), which is partially coeval with the Hamilton Group of New York. Ellison (1965) described the Mahantango Formation as consisting of five members with a total thickness of 959 ft. To determine the stratigraphic position of the type locality of G. boothi, we must first turn to Green (1837). Green (1837: p. 341) stated that "this magnificent trilobite was found near the town of Huntingdon" and that "all were found in the same locality." Although Green's (1837) site description is vague and of little help regarding where to begin searching, he did report something about the rock type G. boothi was recovered from. He referred to the rock as a ferruginous clay

slate filled with other petrifications, and stated that this stratum rests on a coarse-grained sandstone filled with organic remains. It is highly likely that this description corresponds to the fossiliferous Frame Shale Member of the Mahantango Formation, which overlies the fossiliferous Donation Siltstone Member.

Ellison (1965: p. 162) suggested that in the Huntingdon, Pennsylvania, area G. boothi and Bellacartwrightia calliteles have only been found in the upper part of the Frame Shale Member. Ellison (1965) added that the two taxa occur side by side and that Green (1837) must have collected them in that area. These conclusions appear to be partly erroneous. First, Green did not collect specimens of G. boothi or Bellacartwrightia calliteles. The specimens of these taxa were given to Green by collectors. In fact, Green (1837) named the species G. boothi in honor of the collector, Professor James C. Booth. He received a pygidium of B. calliteles from Dr. R. M. S. Jackson. Green (1837) gave no indication that he ever visited the collecting site. In addition, nowhere did Green (1837) state that the taxa were collected together. contra Ellison (1965). In fact, Green (1837) gave no detailed locality information for B. calliteles, and stated that it occurred in a soft ferruginous slate that had sparkling particles of iron pyrite sparsely disseminated throughout. This rock description is different from the one he gave for G. boothi. Thus, there is no indication from Green's (1837) descriptions to support Ellison's (1965) assertion that the trilobites were collected side by side.

In the summer of 1995, GJK visited Huntingdon, Pennsylvania, to collect Asteropyginae in the Mahantango Formation. It soon became apparent, as in New York, that there are many different species of asteropyginines occurring at different levels. At least three species of Greenops and two species of Bellacartwrightia were collected in that formation. This certainly makes it difficult to select an appropriate neotype for G. boothi. However, using Green's (1837) statements, Ellison's (1965) fieldwork, and the results from GJK's collections in Huntingdon, we conclude that the most likely type horizon for G. boothi is the Frame Member of the Mahantango Formation. This is based on the following: (1) the lithology of the Frame Member fits Green's (1837) description of the rock type in which G. boothi was collected; (2) Green (1837) stated that G. boothi occurs in the ferruginous clay slate that reposes on a coarse-grained sandstone, and this description corresponds to the Frame Member, which sits on the Donation Member; (3) Green (1837) stated that the rock is filled with petrifications, and the Frame Member is quite fossiliferous; and (4) the site where specimens were collected appears to have been quite fossiliferous, attracting several collectors who found G. boothi, according to Green (1837). This suggests that the site most likely occurs at an interval in the fossil record where G. boothi is common. Ellison (1965: 54) stated that G. boothi occurs at five different horizons in the Frame Member, and it is far more abundant in this member than in any of the other members of the Mahantango Formation.

However, even within the Frame Member of the Mahantango Formation, two different species referable to *Greenops* were collected. One species occurs in Ellison's (1965: 247) section 16 in the 7-12 ft interval below the top of the Frame. The other species occurs at a lower level and was collected from concretions in a road cut along Route 26. This creates a new problem. Green's (1837) description of Greenops boothi is not sufficiently detailed to determine which species should be chosen as the neotype of G. boothi. However, one of the two species of Greenops that occurs in the Frame Member occurs in the more shaly intervals of that member and is always thoroughly distorted by tectonic processes. If we are to erect a neotype for G. boothi for the purposes of providing nomenclatorial stability, it is essential that we be able to provide a series of characters that are diagnostic of that species and that allow it to be distinguished from other taxa easily. In the case of the deformed specimens this was not possible. However, the other species of Greenops in the Frame Member is known from a single specimen preserved in a limestone concretion. When it was prepared, it was found to be of exceptional quality, showing in excellent detail the features needed for a suitable description and a list of diagnostic characters. Thus, this specimen,

YPM 35807, is designated the neotype for *Greenops boothi* because: (1) it fits Green's (1837) description; (2) it is from the type locality in Huntingdon, Pennsylvania; (3) although the specimen is from a limestone concretion, the concretion occurs in rocks fitting the lithology described by Green (1837); and (4) the exceptional quality of the specimen will avoid confusion over diagnostic characters.

DIAGNOSIS: S2 roughly transverse, weakly converse anteriorly; S1 crescent shaped, medial portion developed as smooth curve convex posteriorly; posterior edge of visual surfaces posterior to transverse line tangent to anterior edge of L0; distance between frontal lobe and facial suture at midline relative to laterally increases slightly; medially anterior cephalic border in dorsal view pointed; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 30°; plane from medial to lateral edges of genal spine evenly inclined; axial furrows anterior to S1 relative to furrows posterior to S1 relative to sagittal line diverge more strongly; visual surface in dorsal view not present within sagittal line drawn tangent to lateral margins of frontal lobe; interior margin of genal spine relative to sagittal line deflected laterally 20°; terminal pygidial lappet broad anteriorly, subrectangular, convex posteriorly; distal tips of pygidial lappets developed as blunt triangles; laterally pygidial interpleural furrows flex more strongly posteriorly; anterior margins of first five pygidial axial rings going from axial furrows to midline straight, then strongly arched anteriorly medially; pygidial lappets long; two transverse rows of circular fenestrae on anterior band of thoracic pleural segments.

MATERIAL: YPM 35807.

DESCRIPTION: Cephalic length (sag.) 45% of width. Axial furrow diverges more strongly anterior to S1 than posterior to S1, diverges less strongly anterior to anterior half of L3, diverging forward at about 25°, narrow, moderately incised. Cranidial anterior border developed as moderate lip deflected moderately anterior to facial suture. Preglabellar furrow shallow and narrow. Length (sag.) of cranidial anterior border increases slightly from lateral to medial edges of frontal lobe. Facial suture anterior to eyes flexes

laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length equal to 100% of width across frontal lobe; frontal lobe ellipsoid, about 55% of length of glabella; L1-L3 flat (sag.); frontal lobe at level of posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI rounded depression. S3 shallow, straight, anterolateral edge transverse, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening distally, outer edge forms smaller angle with sagittal line at anterior end than at posterior end. S2 weakly convex anteriorly, roughly transverse, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial portion developed as smooth curve convex anteriorly, laterally contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly medially, deep. L0 55% of width of frontal lobe, moderately arched (tr.), longest sagittally, bearing single tubercle medially. Large Eye Index 0.50. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite S0, posterior to transverse line tangent to L0; eye raised above frontal lobe; maximum of six lenses per dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 30% of length of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, anterior margin nearly straight. Genal spine developed as moderately long flange extending back to fifth thoracic segment, cephalic border furrow bisects spine; genal spine evenly inclined from medial to lateral edge; medial edge of genal spine not significantly thickened relative to lateral edge; lateral margin of genal spine parallels sagittal line, straight at posterior end; interior margin of genal spine deflecting laterally. Raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border

furrow forms smooth curve convex distally. Lateral border furrow narrow, shallow; lateral border weakly widens posteriorly. Anterior margin of cephalic doublure pointed. Prosopon of fine tubercles.

Thorax of 11 segments. Axial ring about 25% of width of thorax, of equal length (exsag., sag.), at distal ends and medially flexed forward. Ring furrow of equal length, moderately incised, flexing forward medially and distally. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; at lateral margin of axial rings, circular fenestrae present. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.), outer part of pleurae more strongly flexed downward, straight in posterior segments. Posterior band long (exsag.), anterior margin straight, deflected at constant small angle posteriorly; length shortens as length of pleural furrow increases medially; after approximate midpoint, length of pleural furrow decreases laterally; length of posterior band increases, becomes planar; posterior surface declined posterodorsally, with few circular fenestrae near axis; anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially transverse; deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally, with two transverse rows of circular fenestrae.

Pygidium broadly triangular in outline excluding marginal lappets, length about 50% of width without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 25° angle anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Axis 30% of pygidial width anteriorly, with 11 rings; rings of nearly equal length distally and sagittally, longest sagittally; anterior edge of axial rings posterior to fifth axial ring from midline to distal edge convex anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips developed as blunt triangles. Axial terminus subrectangular, convex posteriorly, broad, does not project as far posteriorly as other lappets; postaxial region

25% of pygidial length (sag.). Pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; anterior and posterior bands of pleural segment equally elevated; tops of segments flat; pleural furrows deeper than interpleural furrows, short (exsag.). Laterally pygidial interpleural furrows flex more strongly posteriorly. In medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is only known from the type locality. Hall and Clarke (1888) assigned a series of species to their Dalmanites (Cryphaeus) boothi. However, the bulk of the taxa they illustrated are different from true G. boothi in having: axial nodes on the thoracic and pygidial axial rings and L0; a maximum of seven dorsoventral files; terminal pygidial lappet subrectangular vet pointed posteriorly; and S3 sinusoidal. It is interesting to note that these characteristics are not actually as they are depicted in Hall and Clarke's (1888), plates which more closely resemble true G. boothi and G. grabaui, new species. The specimens they illustrated represent a new species of asteropyginid from the Kashong Shale of the Hamilton Group, which shall be described in a forthcoming work (Lieberman and Kloc, in prep.). Only one of the species in the NYSM's numbered type collections for Dalmanites (Cryphaeus) boothi represents G. grabaui. None represent G. boothi as defined using a neotype from Green's (1837) type locality.

Delo's (1935, 1940) papers as they bear on the problem of *G. boothi* are discussed above in the section on the designation of a neotype, which is necessary for nomenclatorial stability. Shimer and Shrock's (1944: 647, pl. 274, figs. 11, 12) illustrations of material they assigned to *G. boothi*, being based on Delo (1940), are thus also not correctly designated as *G. boothi*.

Many of the taxa that Eldredge (1971a), Ludvigsen (1979), and Babcock and Speyer (1987) figured as G. boothi or assigned to the genus Greenops can no longer be considered to belong to either of these taxa. In particular, Eldredge's (1971a: pl. 14, fig. 4) material belongs to Bellacartwrightia jennyae, new species, on the basis of the structure of the py-

gidial pleural lappets, the number of pygidial axial rings, the presence of tubercles on L0 and on the thoracic and pygidial axial rings, and the genal spines that have a large flattened shelf dorsally and project very far posteriorly (back to the eighth thoracic segment).

Ludvigsen (1979: fig. 55a, b) illustrated two specimens from Hamilton Group aged strata in the Michigan Basin from southwestern Ontario that he assigned to G. boothi. Ludvigsen's (1979) illustrated taxa appear to be conspecific with the taxon from southwestern Ontario that Stumm (1953: pl. 7, figs. 1-3) assigned to G. boothi. These taxa differ from G. boothi in the condition of a few of their character states, and on the basis of these consistent character differences they must be assigned to G. widderensis, new species. Greenops widderensis can be differentiated from G. boothi by the following characters: in G. widderensis the distance between the frontal lobe and the facial suture at the midline, relative to the distance at its lateral margin, is constant, and in G. boothi the distance increases slightly; in G. widderensis there is no prominent ornamentation on the medial portion of L0, and in G. boothi there is a single tubercle; in G. widderensis a line from the posterior to the anterior edge of the eye forms an angle to a sagittal line that is approximately 10°, and in G. boothi the angle is approximately 30°; in G. widderensis medially the genal spine is developed as a flattened ledge and laterally it is evenly inclined, and in G. boothi the genal spine is evenly inclined from its medial to lateral edges; in G. widderensis the interior margin of the genal spine parallels the lateral margin before deflecting laterally, and in G. boothi the interior margin of the genal spine is deflected evenly laterally; in G. widderensis the distal tips of the pygidial lappets are pointed, and in G. boothi they are developed as blunt triangles; and in G. widderensis there is one transverse row of circular fenestrae on the anterior band of the thoracic pleural segments, and in G. boothi there are two transverse rows of circular fenestrae.

Greenops boothi and G. widderensis are probably closely related, but the exact phylogenetic position of G. boothi relative to the other species of Greenops was not deter-

mined. This shall be done in a forthcoming publication (Lieberman and Kloc, in prep.).

Ludvigsen (1979: fig. 54b) illustrated a reconstruction of *G. boothi*. His reconstruction actually combines characters of both *G. grabaui* and *G. widderensis*, with the pygidium resembling those more typical of *G. grabaui* (having a broad terminal lappet convex posteriorly) and the head conforming more closely to that typical of *G. widderensis*.

Hall and Clarke (1888) and other authors traditionally treated one of the divergent asteropyginine morphotypes in the Hamilton Group of ENA as a variety of G. boothi. They referred to this species as Dalmanites (Cryphaeus) boothi var. calliteles. However, this species is in fact distantly related to G. boothi and is instead assigned to a new genus. These taxa differ in the condition of several characters, including: 1, 2, 4, 5, 6, 7, 14, 15, 16, 23, 25, 26, 28, 31, 35, 38, 39, 46, 47, 50, 52, 55, 56, 57, 60, 61, 62, 65, and 66, and they can no longer be considered as separate varieties of a single species. They belong to divergent evolutionary lineages that last shared a common ancestor sometime in the Early Devonian (?Pragian), significantly predating the Middle Devonian (largely Givetian) Hamilton Group. Hall and Clarke (1888) originally separated the two taxa based on characters of the pygidia that they claimed were correlated with differences in other features. They recognized that these differences were evident in early ontogenetic stages, and they claimed that they were "genetic," meaning real rather than ecophenotypic differences. Although the differences do appear to be real, that is, the taxa are diagnosably distinct, the propinquity of descent suggested by Hall and Clarke (1888) is probably an artifact of their systematic method, which united the taxa because they were recovered from similar-aged sediments in similar localities.

Hall and Clarke (1888) erroneously assigned a range of taxa to both Dalmanites (Cryphaeus) boothi and D. (C.) boothi var. calliteles; however, they recognized the range of variation inherent in their samples of asteropyginine taxa, which they largely allocated to two distinct taxa. They claimed that the bulk of variation from what they perceived as the type of Cryphaeus boothi was

in characters of the pygidial fimbria. In addition, they stated that no variation has taken place in this feature without correlative variation in certain other respects. They suggested that in many individuals, particularly the specimens they recovered from the limestones of the Hamilton Group and the underlying Onondaga Limestone, the lobes of the pygidial pleural segments are narrow, lanceolate, and elevated, rather than flat, broad, and contiguous, as they typically are in G. boothi. They recognized that the pygidia with the narrow, rounded, and elevated pygidial pleural segments are identical to those described by Green (1837) as Cryphaeus calliteles, whereas the types with the flat, broad, and contiguous pygidial pleural segments are conspecific with the form described by Green (1837) as C. boothi. Both species were first described using specimens from the same Hamilton Group locality in Huntingdon, Pennsylvania, However, these types were later lost, and Hall and Clarke's (1888) taxa were based on specimens from Hamilton Group aged sediments in New York, not Pennsylvania. There are also probably two more new species in the set of types they figured that will be discussed in a forthcoming work (Lieberman and Kloc, in prep.). Not counting the two new species to be described in a forthcoming paper, this range of variation that they figured and discussed can actually be assigned to three distinct taxa instead of the two they called Dalmanites (Cryphaeus) boothi and D. (C.) boothi var. calliteles. The three taxa are G. grabaui, Bellacartwrightia phyllocaudata, and Bellacartwrightia iennyae.

Struve (1959) illustrated a cephalon and pygidium that he assigned to *Greenops* (*Greenops*) boothi. The cephalon bears a strong resemblance to *G. boothi*. However, it is based on a specimen from Ontario, Canada. *Greenops widderensis*, new species, a close relative of *G. boothi*, is the only species of *Greenops* described from Ontario, though *Stummiana arkonensis* (Stumm, 1953) also occurs in the Middle Devonian of Ontario. The cephalon as drawn bears the diagnostic characteristics of *Greenops* rather than *Stummiana*, and it is presumed that the cephalon illustrated belongs to *G. widderensis* rather than *G. boothi*. The pygidium, figure 381, 1b,

hails from Naples, New York, and has 11 axial rings, with a broad rectangular pygidial terminal lappet that is convex posteriorly, the distal tips of the lappets developed as blunt triangles, and the pleural lappets relatively short. All these morphological features are characteristic of G. grabaui, and because the specimen is from New York it is treated as that taxon rather than G. boothi, which appears to be restricted to Pennsylvania. The assertion that the locality this pygidium hails from is Naples, New York, appears dubious. Only Upper Devonian strata are known from this locality, and no species of asteropyginines are known from the Upper Devonian of ENA. Thus, this pygidium and the locality data are problematic.

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Speyer (1988) figured two specimens from the Kashong Shale of the Hamilton Group that he referred to as Greenops boothi and Greenops sp. These taxa differ from G. boothi in the condition of several characters, and they represent a new species that will be described in a forthcoming paper (Lieberman and Kloc, in prep.). Speyer (1988) also illustrated specimens from New York (figs. 1c, d, 9c) that, because of their state of preservation, must be ambiguously assigned to either G. grabaui, new species, or G. barberi, new species, rather than G. boothi. Speyer's (1988) figures 7a, 8c, d, which he assigned to G. boothi, must instead be assigned to G. barberi because of the condition of the pygidial pleural lappets, the pygidial axis, and the genal spines.

Babcock and Speyer (1987) illustrated specimens (figs. 4.1-4.5) from the Ledyard Shale of New York that they assigned to G. boothi. Because of the state of preservation of these taxa, it could not be ascertained whether these specimens should be assigned to G. grabaui or G. barberi. They are unlikely to represent true G. boothi.

Ellison (1965: pl. 18, fig. 8) illustrated a specimen he assigned to Greenops from the upper shale member of the Mahantango Formation at Girty's Notch, Pennsylvania. This specimen is only preserved as an internal mold, and it is impossible to determine for sure which species of Greenops it belongs to. It is questionably assigned to G. boothi.

Burns (1991) illustrated specimens from the Mahantango Formation (Erian/Givetian) of West Virginia and Virginia that he referred to as G. boothi. However, because of the nature of his illustrations it could not be determined whether the taxa he discussed belong to G. boothi, some other species of Greenops, Bellacartwrightia phyllocaudata, or some other species of Bellacartwrightia. As only B. phyllocaudata, B. calliteles, and G. boothi are known from localities further south in the Appalachian Basin, probably one of these is depicted.

Other authors have suggested that Greenops boothi, or a form nearly identical to it, is also known from Armorica. Keyser (1878) suggested that the species that had been referred to as Cryphaeus boothi and C. calliteles were in fact synonymous with other European asteropyginid taxa such as Cryphaeus stellifer. Cryphaeus stellifer is discussed above as Neometacanthus stellifer, and it does differ in the condition of several characters from G. boothi. The two cannot be considered synonyms, and they are not close evolutionarily. Moreover, Hall and Clarke (1888) gave several other cogent arguments why Keyser (1878) could not be correct, and now his contention of synonymy between the Armorican and ENA asteropyginine taxa he discussed appears untenable. In more recent times, other authors have advanced different arguments along similar lines. In particular, Gandl (1972) referred to a series of specimens as Greenops sp. cf. boothi, and these were also discussed by Smeenk (1983). These specimens belong to Armorigreenops leoi, new species, which is closely related to the ENA Greenops clade, but it is not sister to G. boothi. It differs from that taxon in the condition of several characters that are discussed below. The specimen Arbizu (1979) referred to as G. sp. cf. boothi belongs to a different species, Deloops sp. aff. haasi, and is also discussed below.

Haas (1970: 4, fig. 10) figured a pygidium housed at Universität Munster that he referred to as G. (G.) boothi. This specimen has similarities with species of *Greenops*. In particular, the pygidial interpleural furrows flex more strongly posteriorly laterally, the lateral margins of the pygidial pleural lappets are curved, the fifth pair of pygidial pleural lappets projects further posteriorly than the terminal pygidial lappet, and the tops of the

pygidial pleural segments are flattened. It also bears some of the characters found in some, though not all, of the species in the genus Greenops. In Haas's (1970) pygidium, the anterior parts of the first five pygidial axial rings are not straight and then strongly arching anteriorly when proceeding from the axial furrows to the midline. Rather, they are gently convex anteriorly. In addition, it differs from all species of Greenops in the condition of the anterior part of the pygidial axial rings posterior to the fifth axial ring, which are straight instead of gently convex. This pygidium can be further distinguished from G. boothi by the possession of 12 axial rings and by the condition of the distal tips of the pygidial pleural lappets, which are at the distal end of a straight posterolateral margin inclined at a 45° angle relative to a sagittal line in Haas's (1970) pygidium but developed as blunt triangles in G. boothi, though this character does not diagnose all species of Greenops (no species of Greenops have the pygidial lappets as they are in Haas's pygidium). Without a cephalon present, it is difficult to ascertain the precise phylogenetic position of this specimen. However, the pygidium does possess some of the diagnostic characters found in some species of Philipsmithiana, new genus. In particular, Haas's (1970) pygidium has the distal tips of the pygidial pleural lappets at the distal end of a straight posterolateral margin of the pygidial pleural lappets, as well as the short (exsag.) pygidial lappets, and the condition of the anterior margins of the pygidial axial rings that characterizes all species of Philipsmithiana. Without further material it is not possible to determine if this species should be assigned to Philipsmithiana or Greenops. However, Haas's (1970) plate 4, figure 11, from the Wanakah Shale of the Ludlowville Formation does appear referable to a close relative of G. boothi, G. grabaui, new species.

Morzadec (1983) presented an evolutionary tree (his fig. 33) that depicts the phylogenetic relationships of some of the species of *Greenops*. Morzadec (1983) suggested that *Greenops boothi* is sister to a clade containing what are here *Deloops haasi* and *Stummiana osismorum*. He was correct in asserting the close relationship of these taxa,

although the precise topology of his phylogenetic scheme is not supported herein. Presumably, he assumed monophyly of the ENA Greenops, though he did not discuss this point. A questionable aspect of his evolutionary tree is his placement of G. struvei as a direct ancestor of G. osismorum (here Stummiana osismorum). As discussed above, Morzadec's (1983) G. struvei is assigned herein to a new genus, Modellops. This genus appears to be distantly related to Greenops, and any similarities the genera share are the result of primitive retentions in Greenops, or convergences.

Greenops chilmanae Stumm, 1965

Greenops chilmanae Stumm, 1965: 165, pl. 1, fig. 9; Stumm, 1967: 114, pl. 2, figs. 4, 5; Kesling et al., 1974: pl. 12, fig. 29; Kesling and Chilman, 1975: 154, pl. 31, figs. 5-7.

Greenops aequituberculatus Stumm, 1953: 131, pl. 8, fig. 9.

TYPES: Holotype defined in Stumm (1965: pl. 1, fig. 9).

DIAGNOSIS: Anterior cephalic border rounded; fixigenae evenly sloping from axial furrows to internal margins of eye; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 0-5°; before cephalic lateral border furrow meets posterior border furrow, posterior border furrow flexes anteriorly; tubercle present on L0; cephalic axial furrows diverge at same angle anterior and posterior to S1; one transverse row of circular fenestrae on anterior band of thoracic pleural segments; terminal pygidial lappet triangular, does not project as far posteriorly as pygidial pleural lappets flanking it; lateral margins of pygidial lappets curved, medial margins straight; distal tips of lappets pointed; longest pygidial lappet longer than sagittal length of first three pygidial axial rings.

DISCUSSION: This species is known from the following latest Eifelian and/or Givetian localities in the Traverse Group of the Michigan Basin in ENA: Gravel Point Formation (Cazenovian), Longispina emmetensis zone, Lake Michigan at Gravel Point, 1.5–2 miles west of Charlevoix, Michigan; Silica Shale (Cazenovian), north quarry of the Medusa Portland Cement Company at Silica, 1.5

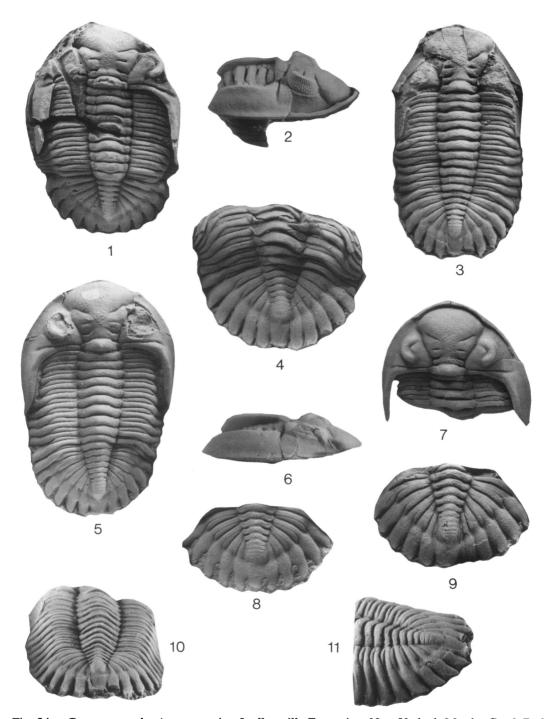


Fig. 24. *Greenops grabaui*, new species, Ludlowville Formation, New York. 1. Murder Creek Beds, Wanakah Shale, Darien, AMNH 45307, dorsal view of complete specimen, ×2.4. 2, 7. AMNH 45290, lateral and dorsal views of cephalon and portion of thorax of complete individual, ×4. 8. AMNH 45290, dorsal view of pygidium of complete individual, ×4. 3, 10. Murder Creek Beds, Wanakah Shale, Darien, AMNH 45309, dorsal and posterior views of complete individual, ×3. 11. Murder Creek Beds, Wanakah Shale, Darien, AMNH 45309, lateral view of pygidium of complete individual, ×3. 4. AMNH 45291,

miles southwest of Sylvania, Lucas County, Ohio, AMNH locality 3063. No specimens are known from post-Cazenovian strata.

As discussed above, G. aequituberculatus encompassed a series of species and can no longer be considered valid. One of the specimens Stumm (1953) designated as a paratype of G. aequituberculatus is actually a pygidium of G. chilmanae. Stumm (1965) suggested that G. chilmanae was most closely related to G. boothi (Green). These taxa are certainly very closely related, but at this time the precise phylogenetic position of Greenops boothi was not examined, so this proposition could not be tested. Instead, G. chilmanae is sister to G. barberi, new species, and G. widderensis, new species, is sister to the clade they comprise. However, considering that Stumm (1965) was unaware at the time he wrote the paper of the two new species of Greenops described herein, and he had not used a concept of G. boothi based on material from Green's (1937) type locality, his conclusion is not too far off base. The sister group relationship between G. chilmanae and G. barberi is supported by their shared possession of the following characters: S2 weakly declined posteriorly at lateral edge; visual surfaces in dorsal view present within sagittal lines drawn tangent to the margins of the frontal lobe; and terminal pygidial lappet triangular and narrow.

Greenops grabaui, new species Figures 8.6, 9.3–9.7, 10.1–10.5, 24, 25.1–25.5, 25.7, 25.8

Dalmanites (Cryphaeus) boothi (Green). Hall and Clarke, 1888: 42, pl. 16, fig. 4, pl. 16a, fig. 3. Greenops boothi (Green). Babcock and Speyer, 1987: figs. 4.1-4.5; Speyer, 1988: figs. 1c, d, 8c, d, 9c.

?Greenops boothi (Green). Eldredge, 1971a: pl. 14, fig. 6.

?Greenops (Greenops) boothi (Green). Struve, 1959: O480, fig. 381, 1b; Haas, 1970: 113, pl. 4, fig. 11.

?Dalmanites (Cryphaeus) boothi (Green). Hall and Clarke, 1888: pl. 16a, fig. 3.

Types: Holotype BMS E25857, a complete specimen from the Murder Creek bed of the Wanakah Shale (Tioughniogan), Ludlowville Formation, Hamilton Group, along the shore of Lake Erie, north of 18 Mile Creek, Hamburg, New York. Paratypes AMNH 45228 and 45229, complete specimens from the lower Ledyard Shale (Tioughniogan), Ludlowville Formation, Hamilton Group, Jaycox Run Creek, upstream from Nations Road, Geneseo Township, Livingston County, New York, AMNH locality 3041; and AMNH 45308, a complete specimen from the Murder Creek Beds, Wanakah Shale (Tioughniogan), Ludlowville Formation, Hamilton Group, Darien, New York.

DIAGNOSIS: Anterior cephalic border rounded: distance between frontal lobe and facial suture at midline relative to distance between frontal lobe and facial suture at lateral margins of frontal lobe equivalent; anteriormost part of palpebrum between medial tip of L3 and PMI; fixigena evenly sloping from axial furrow to internal margin of eve: cephalic axial furrows anterior to S1 diverge more strongly than furrows posterior to S1; S2 weakly convex anteriorly; line from posterior to anterior edge of eye forms 30° angle to sagittal line; posterior edge of visual surface posterior to transverse line tangent to L0; lateral to eye, facial suture before flexing posteriorly flexes anterior to point opposite distal tip of S2; posterior margin of fixigena relative to anterior margin of fixigena deflected at an angle to transverse line that is roughly equivalent; tubercle on L0 absent; interior margin of genal spine parallels lateral margin before deflecting laterally; medial tubercles on pygidial and thoracic axial rings absent; single pair of circular fenestrae on thoracic axial rings flanking medial part of axis: one transverse row of circular fenestrae on anterior band of thoracic pleural seg-

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dorsal view of pygidium and portion of thorax of complete individual, $\times 3$. 6. AMNH 45291, lateral view of cephalon of complete individual, $\times 3$. 5. Murder Creek Beds, Wanakah Shale, Darien, AMNH 45308, dorsal view of complete paratype individual, $\times 4$. 9. AMNH 45292, dorsal view of pygidium, $\times 3$.

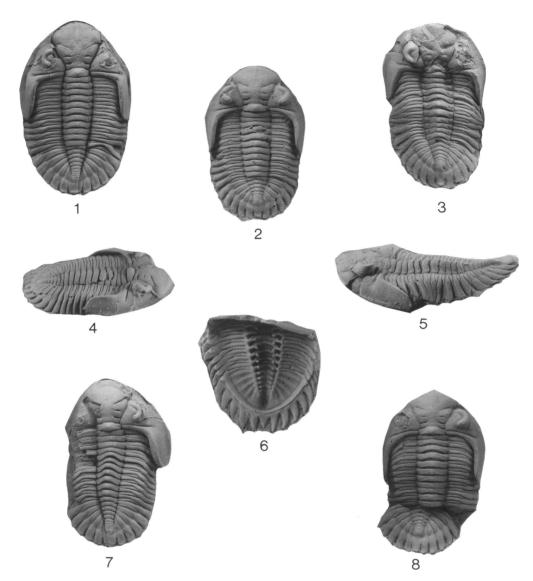


Fig. 25. Greenops spp. 1–5, 7, 8. Greenops grabaui, new species, New York. 1. Ledyard Shale, Ludlowville Formation, Jaycox Run Creek, upstream of Nations Road, Geneseo Township, Livingston County, AMNH 45228, dorsal view of complete paratype individual, ×1.8. 2, 4. Murder Creek Bed, Wanakah Shale, Ludlowville Formation, along Lake Erie shore, north of 18 Mile Creek, Hamburg, BMS E25857, dorsal and lateral views of complete holotype individual, ×2. 3, 5. ?Ludlowville Formation, Canandaigua Lake, NYSM 4281, dorsal and lateral views of complete individual, ×1.7. 7. Ledyard Shale, Ludlowville Formation, Jaycox Run Creek, upstream of Nations Road, Geneseo Township, Livingston County, AMNH 45229, dorsal view of complete paratype individual, ×2. 8. Murder Creek Bed, Wanakah Shale, Ludlowville Formation, along Lake Erie shore, north of 18 Mile Creek, Hamburg, BMS E25856, dorsal view of complete individual, ×2. 6. Greenops barberi, new species, Smoke Creek Bed, Windom Shale, Moscow Formation, Penn Dixie Quarry at intersection between Big Tree and Bayview Roads, BMS E25855, ventral view of pygidium and portion of thorax, ×2.

ments; terminal pygidial lappet subrectangular, broad, 1.5 times width (tr.) of posteriormost part of pygidial axis; medial margins of pygidial lappets straight; distal tips of pygidial lappets developed as blunt triangles; pygidial pleural lappets relatively short (exsag.); anterior and posterior pleural bands of pygidial segments of equal elevation; tops of pygidial pleural segments flat; in medial region of adjacent pleural segments posterior band of anterior segment equal in length (exsag.) to anterior band of posterior segment.

ETYMOLOGY: Named for A. Grabau, who made many important contributions to stratigraphic methodology in general, and the stratigraphy of the trilobite beds of the Wanakah Shale in particular, which is the type locality of the species.

MATERIAL: AMNH 45228, 45229, 45254, 45274, 45275, 45290–45292, 45307–45309, 45317; BMS E25856, E25857; NYSM 4283; YPM 6603, 6665, 33764, 33783–33785, 33787, 33855–33858, 35740, ?33778, ?33786–?33788, ?35742.

DESCRIPTION: Cephalic length (sag.) 45% width. Axial furrow diverges more strongly anterior to S1 than posterior to S1, diverges less strongly anterior to anterior half of L3, diverging forward at about 25°, narrow, moderately incised. Cranidial anterior border developed as moderate lip deflected moderately anterior to facial suture. Preglabellar furrow shallow and narrow. Length (exsag., sag.) of cranidial anterior border constant from lateral to medial edges of frontal lobe. Facial suture anterior to eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length (sag.) equal to 100% of width (tr.) across frontal lobe; frontal lobe ellipsoid, about 55% of length of glabella; L1-L3 flat (sag.); frontal lobe at same elevation as posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI a rounded depression. S3 shallow, straight, anterolateral edge transverse, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening (exsag.) distally, outer edge forms smaller angle with sagittal line at anterior end than at posterior end. S2 weakly convex anteriorly, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial part developed as smooth curve convex anteriorly, laterally contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 convex anteriorly medially, deep. L0 55% of width (tr.) of frontal lobe, moderately arched (tr.), longest sagittally. Large Eye Index 0.50. Anterior edge of eve contacts axial furrow opposite anterodistal corner of L3: posterior edge of eye opposite distal tip of S0, posterior to transverse line tangent to L0; eye raised above frontal lobe; maximum of six lenses per dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 30% of length (exsag.) of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse. Genal spine developed as moderately long flange extending back to fifth thoracic segment, cephalic border furrow bisects spine; medial part of genal spine flattened, lateral part evenly inclined; medial part of genal spine not significantly thickened relative to lateral part; lateral margin of genal spine parallels sagittal line, straight at posterior end; interior margin of genal spine parallels lateral margin before deflecting laterally. Raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border furrow forms smooth curve convex distally. Lateral border furrow narrow, shallow, lateral border weakly widens (tr.) posteriorly. Anterior margin of cephalic doublure rounded. Prosopon of fine tubercles.

Thorax of 11 segments. Axial ring about 25% of width of thorax, of equal length (exsag., sag.), at distal ends and medially flexing forward. Ring furrow of equal length (exsag., sag.), moderately incised, flexing forward medially and distally. Anterior margin of ring well defined, inclined posterodorsally, most elevated near posterior edge; at lateral margin of axial rings, circular fenestrae present. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal;

anterior segments rather evenly convex (tr.), outer part of pleurae more strongly flexed downward, straighter in posterior segments. Posterior band long (exsag.), anterior margin straight, deflected at constant small angle relative to transverse line posteriorly; length (exsag.) shortens as length of pleural furrow increases medially; posterior surface declined posterodorsally, with few circular fenestrae near axis. Anterior pleural band short (exsag.), medially transverse; deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally, with one transverse row of circular fenestrae.

Pygidium broadly triangular in outline excluding marginal lappets, length about 50% of width without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 25° angle anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Axis 30% of pygidial width anteriorly, with 11 rings; rings of nearly equal length distally and medially, but longest medially; anterior edge of axial rings posterior to fifth axial ring from midline to distal edge convex anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips developed as blunt triangles. Axial terminus subrectangular, convex posteriorly, broad, does not project as far posteriorly as other lappets; postaxial region 25% of pygidial length (sag.). Pygidial pleural field flanking posterior part of pygidial axis faintly excavated; anterior and posterior bands of pleural segment equally elevated; tops of segments flat; pleural furrows deeper than interpleural furrows, short (exsag.). Laterally pygidial interpleural furrows flex more strongly posteriorly. In medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known from the following Givetian stratigraphic horizons and localities in the Hamilton Group of the Appalachian Basin in ENA: lower Ludlow-ville Formation including the Stone Mill Limestone (Tioughniogan), Robert's Road Quarry, north of New York Route 26 on Robert's Road, West Eaton, New York, AMNH locality 3032; Centerfield Lime-

stone, Ludlowville Formation, Brown's Creek, York, New York; Ledvard Shale, Ludlowville Formation, cut along tracks of Delaware and Western Railroad, 3 miles west of East Bethany, New York, AMNH locality 3045; lower Ledyard Shale, 11 Mile Creek, Darien, New York; Wanakah Shale, Ludlowville Formation, Cazenovia Creek, near Transit Road, Erie County, New York, AMNH locality 3048; Ledvard Shale, Spring Creek, Alden, New York; Wanakah Shale, Ludlowville Formation, Bethany, New York; trilobite beds, Wanakah Shale, Smoke Creek. Windom, New York: Wanakah Shale, Murder Creek Beds, Darien, New York; Ludlowville Formation, Seymour Pond, New York; ?Frame Member (Tioughniogan), upper Mahantango Formation, road cut along U.S. Route 22 opposite Huntingdon, Pennsylvania; ?upper Mahantango Formation (Tioughniogan), Cumberland, Maryland, James Hall AMNH locality 10; and ?Mahantango Formation (Tioughniogan), Koshong Creek, Maryland.

This species is very common in the Ludlowville Formation. In the Taghanic Tully Limestone, poorly preserved specimens are known that appear to be referable to *Green*ops. They are probably closely related to *G.* grabaui, but until better material is recovered, this species will not be described.

Struve (1959: fig. 381, 1b) illustrated a pygidium from the Hamilton Group of Naples, New York, that he assigned to Greenops (Greenops) boothi. The pygidium has 11 axial rings with a broad rectangular pygidial terminal lappet that is convex posteriorly, and the distal tips of the lappets are developed as blunt triangles. It is difficult to tell from a drawing, but these morphological features are characteristic of G. grabaui, and the pygidium, being from New York, is questionably treated as belonging to that species rather than G. boothi, which is not known to occur in New York. However, Naples, New York, is an exclusively Upper Devonian locality, and no species belonging to the Asteropyginae are known from the Upper Devonian of ENA. Thus, the veracity of this locality must be treated as dubious.

Several modern authors (e.g., Eldredge, 1971a; Ludvigsen, 1979; Babcock and Speyer, 1987) have assigned a range of astero-

pyginine specimens in the Appalachian Basin of ENA to G. boothi. First, on the basis of character analysis performed herein it appears that G. boothi is not the sole representative of the Asteropyginae in the Appalachian Basin. In addition, Greenops boothi appears to be absent from the portion of the Appalachian Basin preserved in modern-day New York. There are, however, at least two other species of Greenops known from the Appalachian Basin of New York: G. barberi, new species, and G. grabaui, new species. Greenops grabaui can be discriminated from G. boothi by the condition of the following: in G. grabaui the distance between the frontal lobe and the facial suture at the midline, relative to laterally, is constant, whereas in G. boothi this distance increases slightly; in G. grabaui medially the anterior cephalic border in dorsal view is rounded, whereas in G. boothi it is pointed; in G. grabaui there is no prominent ornamentation on the medial portion of L0, whereas in G. boothi there is a single tubercle; in G. grabaui the medial edge of the genal spine is flattened, and laterally it is evenly inclined, whereas in G. boothi the genal spine is evenly inclined from its medial to lateral edges; in G. grabaui the interior margin of the genal spine parallels the lateral margin before deflecting laterally, whereas in G. boothi the interior margin of the genal spine is deflected laterally; in G. grabaui the pygidial lappets are relatively short (exsag.), whereas in G. boothi they are relatively long; and in G. grabaui there is one transverse row of circular fenestrae on the anterior band of the thoracic pleural segments, whereas in G. boothi there are two rows.

Speyer (1988) illustrated specimens from New York (figs. 1c, d, 8c, d, 9c) that are assigned herein to G. grabaui, new species, rather than G. boothi.

Babcock and Speyer (1987) illustrated specimens (figs. 4.1–4.5) from the Ledyard Shale of New York that they assigned to G. boothi. Because of the state of preservation of these taxa, it could not be ascertained whether these specimens should be assigned to G. grabaui or to G. barberi. They are unlikely to represent true G. boothi.

Greenops grabaui, although clearly diagnosable herein, does show some intraspecific

variation in at least one feature. That is the terminal pygidial lappet, which can show some variation in breadth from sample to sample.

Smeenk (1983: pl. 10, figs. 9, 10) figured two preservationally deformed pygidia that he referred to as Greenops (Greenops) sp. Although poorly preserved, these specimens do possess some of the characters found in various species of the genus Greenops. These characters include: the form of the anterior margin of the first five pygidial axial rings, which from axial furrow to midpoint are first straight and then arch strongly anteriorly; the shape of the pygidial interpleural furrows, which laterally flex more strongly posteriorly: the lateral margins of the pygidial pleural lappets are curved; the pygidial axis constricts posterior to the fifth axial ring; the anterior margins of the pygidial axial rings posterior to the constriction of the fifth ring are smoothly convex anteriorly; the pygidial pleural field flanking the posterior portion of the pygidial axis is faintly excavated; and the pleural furrows are moderately incised and short (exsag.). Awaiting the recovery of cephalothoraxes, which of course contain phylogenetically relevant characters, these pygidia are assigned to the genus Greenops. In addition to these characters bespeaking the affinity of these pygidia with the genus Greenops, these pygidia share common character states with G. grabaui, and they may be closely related to that species. These characters include the following: the terminal pygidial lappet is broad, 1.5 times the width (tr.) of the posterior part of the pygidial axis; it is rectangular and convex posteriorly; the medial margins of the pygidial lappets curve laterally; the distal tips of the pygidial lappets are developed as blunt triangles; the anterior and posterior bands of pleural segments are of equal elevation; the tops of the pygidial pleural segments are flat; when comparing adjacent pleural segments the posterior region of the anterior segment is equal in length (exsag.) to the anterior part of the posterior segment; and the longest pygidial lappet is short (exsag.), equal to the length (sag.) of the first two pygidial axial rings. The pygidia Smeenk figured do differ from those of G. grabaui in the possession of 12 pygidial axial rings (versus 11 in G.

grabaui). These specimens are tentatively referred to as G. sp. B, and are treated as very closely related to G. grabaui. However, until better as well as more completely preserved specimens are recovered, this phylogenetic hypothesis cannot be tested.

This potentially close relationship between G. grabaui and G. sp. B raises the possibility of an additional instance of a sister group relationship between asteropyginines in ENA and Armorica. As discussed above and below, there are at least four instances of invasion from Armorica into ENA by the asteropyginines, with several lineages of Armorican taxa sister to one or more clades of ENA taxa. However, if G. sp. B is sister to G. grabaui, then it would imply that there may have been some invasion from ENA back into Armorica. It would also imply that the five species of Greenops in ENA (including Greenops boothi) would not be monophyletic to the exclusion of Armorican Greenops. Greenops sp. B was not incorporated into the phylogenetic analysis conducted herein because of the dearth of material and because of its poor state of preservation. Tentatively, the five species of *Greenops* in ENA are treated as monophyletic until additional material of G. sp. B becomes available, allowing additional testing of this hypothesis.

Greenops barberi, new species Figures 8.1, 8.2, 8.4, 25.6, 26.1–26.7

Greenops boothi (Green). Speyer, 1987: fig. 7g.

Types: Holotype PRI 41947, a complete specimen from the Smoke Creek Bed, Windom Shale (Tioughniogan), Moscow Formation, Hamilton Group, Buffalo Creek, Erie County, New York; paratypes PRI 41948, a pygidium from the same locality as the holotype, PRI 41949, a cephalothorax from the same locality as the holotype, PRI 41951, a nearly complete specimen from the same locality as the holotype, and AMNH 45277 and 45278, two pygidia from the same locality as the holotype.

DIAGNOSIS: Anterior cephalic border rounded; S2 weakly declined posteriorly at lateral edge; medial part of S1 developed as straight line directed anteriorly; posterior edge of visual surface anterior to transverse line tangent to anterior edge of L0; immediately before cephalic lateral border furrow meets posterior border furrow, posterior border furrow straight; distance between frontal lobe and facial suture at midline greater than distance between frontal lobe and facial suture at lateral margins of frontal lobe; ornamentation absent on LO; line from posterior to anterior edge of eye forms 5-10° angle to sagittal line; isolated patches of tubercles on L3 absent; plane from medial to lateral edges of genal spine evenly inclined; axial furrows anterior to S1 relative to furrows posterior to S1 diverge more strongly relative to sagittal line; visual surfaces in dorsal view present within sagittal lines drawn tangent to frontal lobe; interior margin of genal spine parallels lateral margin before deflecting laterally; one transverse row of circular fenestrae on anterior band of thoracic pleural segments; terminal pygidial lappet developed as narrow (tr.) sharply pointed triangle; distal tips of pygidial lappets pointed; pygidial lappets long (exsag.); laterally pygidial interpleural furrows arch at same angle as medially; anterolateral margins of first five pygidial axial rings going from axial furrows to midline straight, then strongly arched anteriorly medially.

ETYMOLOGY: Named for Fred Barber, who contributed the specimens used to define this species.

MATERIAL: AMNH 45256, 45257, 45277–45279; BMS E25855; PRI 41947–41951; YPM ?33778, ?33786, ?33788, ?33855, ?33856.

DESCRIPTION: Cephalic length (sag.) 50% of width. Axial furrow diverges more strongly anterior to S1 than posterior to S1, diverges less strongly anterior to anterior half of L3, diverging forward at about 25°, narrow, moderately incised. Cranidial anterior border developed as moderate lip deflected moderately anterior to facial suture. Preglabellar furrow shallow and narrow. Length (exsag., sag.) of cranidial anterior border increases slightly from lateral to medial edges of frontal lobe. Facial suture anterior to eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length (sag.) equal to 100% of width (tr.) across frontal

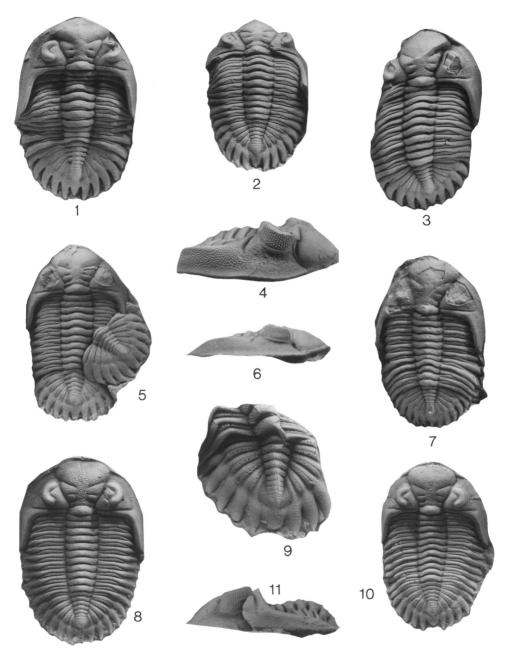


Fig. 26. Greenops spp. 1–7. Greenops barberi, new species, Smoke Creek Bed, Windom Shale, Moscow Formation, Buffalo Creek, New York. 1. AMNH 45277, dorsal view of complete paratype individual, ×2.3. 2, 4. AMNH 45278, dorsal and lateral views of complete paratype individual, ×2 and ×3. 3. PRI 41951, dorsal view of nearly complete paratype individual, ×2.3. 5. PRI 41947, dorsal view of complete holotype individual and PRI 41948, dorsal view of paratype pygidium, ×2. 6. PRI 41949, lateral view of paratype cephalon and partial thorax, ×2.5. 7. AMNH 45279, dorsal view of complete individual, ×4. 8–11. Greenops widderensis, new species, Widder Formation (Tioughniogan), Traverse Group, Hungry Hollow, Arkona, Ontario. 8. AMNH 45276, dorsal view of complete paratype individual, ×2.1. 9. AMNH 45316, dorsal view of pygidium and partial thorax of partially complete individual, ×2. 11. AMNH 45316, lateral view of cephalon of partially complete individual, ×2.5. 10. AMNH 45258, dorsal view of complete holotype individual, ×2.1.

lobe; frontal lobe ellipsoid, about 55% length (sag.) of glabella; L1-L3 flat (sag.); frontal lobe at same elevation as that of posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI a rounded depression. S3 shallow, straight, anterolateral edge transverse, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening distally, outer edge forms smaller angle with sagittal edge at anterior end than at posterior end. S2 weakly declined posteriorly at lateral edge, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial part developed as straight line directed anteriorly, laterally contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly medially, deep. L0 50% of width of frontal lobe, moderately arched (tr.), longest sagittally. Large Eye Index 0.50. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite L1 distal edge of, anterior to transverse line tangent to L0; eye raised above frontal lobe; maximum of six lenses per dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 25% of length (exsag.) of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, anterior margin nearly straight. Genal spine developed as moderately long flange extending back to fifth thoracic segment, cephalic border furrow bisects spine; lateral and medial edges of genal spine evenly inclined; medial edge of genal spine not significantly thickened relative to lateral edge; lateral margin of genal spine parallels sagittal line, straight at posterior end; interior margin of genal spine evenly deflected laterally. Raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border furrow forms smooth curve convex distally. Lateral border furrow narrow, shallow, lateral border weakly widens posteriorly. Anterior margin of cephalic doublure rounded. Prosopon of fine tubercles.

Thorax of 11 segments. Axial ring about 25% of width (tr.) of thorax, of equal length (exsag., sag.), at distal ends and medially flexed forward. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; at lateral margin of axial rings, circular fenestrae present. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.), outer part of pleurae more strongly flexed downward, more straight in posterior segments. Posterior band long (exsag.), anterior margin straight, deflected at constant, small angle posteriorly relative to transverse line; length of posterior band increases laterally, becomes planar; posterior surface declined posterodorsally, with few circular fenestrae near axis; anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially transverse; deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally, with one transverse row of circular fenestrae.

Pygidium broadly triangular in outline excluding marginal lappets, length about 60% of width without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 25° angle anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Axis 30% of pygidial width (tr.) anteriorly, with 11 rings; rings of nearly equal length (exsag., sag.) distally and medially, longest medially; anterior edge of axial rings posterior to fifth axial ring from midline to distal edge convex anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips pointed. Axial terminus triangular, does not project as far posteriorly as other lappets; postaxial region 15% of pygidial length (sag.). Pygidial pleural field flanking posterior part of pygidial axis faintly excavated; anterior and posterior bands of pleural segment equally elevated; tops of segments flat; pleural furrows deeper than interpleural furrows, short (exsag.). Laterally pygidial interpleural furrows arch at same angle as medially. In medial region of adjacent pleural segments, posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known from the following localities in the Givetian of the Appalachian Basin: Wanakah Shale, Ludlow-ville Formation (Tioughniogan), loose concretion, Spring Brook concretionary bed, Cazenovia Creek, Spring Brook, New York; Windom Shale, Moscow Formation (Tioughniogan), Smoke Creek Bed, Buffalo Creek, Erie Co., New York; upper Windom Shale, Big Tree road, near Hamburg, New York; Windom Shale, Moscow Formation, Hatch's Lake, New York.

There appears to be some variation in this taxon. In particular, some specimens have S2 roughly transverse and weakly convex anteriorly, whereas others have S2 weakly declined posteriorly at its lateral edge; some specimens have, in dorsal view, the visual surfaces present within sagittal lines drawn tangent to the lateral edges of the frontal, whereas in others the visual surfaces are not present within such lines; in some specimens the medial margins of the pygidial lappets are curved, whereas in others they are straight; and, finally, in some the anterolateral margins of the first five pygidial axial rings going from the axial furrows to the midline are evenly convex and do not arch strongly anteriorly, whereas in others these margins were first straight and then strongly arched anteriorly medially. The species was scored in phylogenetic analysis as having the derived state for all these characters. At this time, this variation does not appear to have any stratigraphic consistency.

Greenops widderensis, new species

Figures 9.1, 9.2, 26.8-26.11

Greenops boothi (Green). Stumm, 1953: 130, pl. 7, figs. 1-3; Ludvigsen, 1979: 80, figs. 54b, 55a, b.

Greenops (Greenops) boothi (Green). Struve, 1959: O480, fig. 381, 1a.

?Greenops (Greenops?) collitelus (Green). Struve, 1959: O480, fig. 379, 5.

TYPES: Holotype AMNH 45258, a complete specimen from the Widder Formation (Tioughniogan), Hamilton Group, Hungry Hollow, Arkona, Ontario; paratype AMNH 45276, from the same locality as the holotype.

DIAGNOSIS: Anterior cephalic border rounded; S2 roughly transverse, weakly convex anteriorly; medial part of S1 developed as smooth curve convex posteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; immediately before cephalic lateral border furrow meets posterior border furrow, posterior border furrow straight; distance between frontal lobe and facial suture at midline relative to laterally is constant; ornamentation on L0 absent; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 5-10°; isolated patches of tubercles present on L3; medially genal spine flattened ledge, laterally evenly inclined; axial furrows anterior to S1 relative to posterior diverge more strongly; visual surfaces in dorsal view not present within sagittal line drawn tangent to frontal lobe; interior margin of genal spine parallels lateral margin before deflecting laterally; one transverse row of circular fenestrae on anterior band of thoracic pleural segments; terminal pygidial lappet broad, subrectangular, convex posteriorly; distal tips of pygidial lappets pointed; pygidial lappets long (exsag.); laterally pygidial interpleural furrows flex more strongly posteriorly; anterior margins of first five pygidial axial rings going from axial furrows to midline evenly convex and do not arch strongly anteriorly.

ETYMOLOGY: The name of this species is derived from the locality where the specimens were collected, the Widder Formation of the Hamilton Group, Hungry Hollow, Arkona, Ontario.

MATERIAL: AMNH 45258, 45276, 45316; ROM 1731H; YPM 7039, 35738, 35739.

DESCRIPTION: Cephalic length (sag.) 50% of width (tr.). Axial furrow diverges more strongly anterior to S1 than posterior to S1, diverges less strongly anterior to anterior half of L3, diverging forward at about 25°, narrow, moderately incised. Cranidial anterior border developed as moderate lip deflected moderately anterior to facial suture. Preglabellar furrow shallow and narrow. Length (exsag., sag.) of cranidial anterior border constant from lateral to medial edges of frontal lobe. Facial suture anterior to eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of fa-

cial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length (sag.) equal to 100% of width (tr.) across frontal lobe; frontal lobe ellipsoid, about 55% length (sag.) of glabella; L1-L3 flat (sag.); frontal lobe at same elevation as posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI a rounded depression. S3 shallow, straight, anterolateral edge transverse, equally incised medially and distally, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening distally, outer edge forms smaller angle with sagittal edge at anterior end than at posterior end. S2 weakly declined posteriorly at lateral edge, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial part developed as smooth curve convex anteriorly, laterally contacting axial furrow. L1 short (exsag.), flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 convex anteriorly medially, deep. L0 50% width (tr.) of frontal lobe, moderately arched (tr.), longest sagittally. Large Eye Index 0.50. Anterior edge of eye contacts axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite distal tip of S0, posterior to transverse line tangent to L0; eye raised above frontal lobe; maximum of six lenses per dorsoventral file on visual surface. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 25% of length (exsag.) of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, anterior margin nearly straight. Genal spine developed as moderately long flange extending back to fifth thoracic segment, cephalic border furrow bisects spine; medial edge of genal spine flattened, lateral edge evenly inclined; medial edge of genal spine not significantly thickened relative to lateral edge; lateral margin of genal spine parallels sagittal line, straight at posterior end; interior margin of genal spine parallels lateral margin before deflecting laterally, evenly deflected laterally. Raised ridge on dorsal surface of genal spine at juncture of posterior border furrow and lateral border furrow forms smooth curve convex distally.

Lateral border furrow narrow, shallow, lateral border weakly widens posteriorly. Anterior margin of cephalic doublure rounded. Prosopon of fine tubercles.

Thorax of 11 segments. Axial ring about 25% of width (tr.) of thorax, of equal length (exsag., sag.), at distal ends and medially flexed forward. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; at lateral margin of axial rings, circular fenestrae present. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.), outer part of pleurae more strongly flexed downward, straight in posterior segments. Posterior band long (exsag.), anterior margin straight, deflected at constant, small angle posteriorly relative to transverse line; length (exag.) of posterior band increases laterally, band becomes planar laterally; posterior surface declined posterodorsally, with few circular fenestrae near axis; anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially transverse; deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally, with one transverse row of circular fenestrae.

Pygidium broadly triangular in outline excluding marginal lappets, length about 50% of width without lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 25° angle anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Axis 30% of pygidial width anteriorly, with 11 rings; rings of nearly equal length (exsag., sag.) distally and medially, longest medially; anterior edge of axial rings posterior to fifth axial ring from midline to distal edge convex anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips pointed. Axial terminus subrectangular, convex posteriorly, broad, does not project as far posteriorly as other lappets; postaxial region 15% of pygidial length (sag.). Pygidial pleural field flanking posterior part of pygidial axis faintly excavated; anterior and posterior bands of pleural segment equally elevated; tops of segments flat; pleural furrows deeper than interpleural furrows, short (exsag.). Laterally pygidial interpleural furrows flex more strongly posteriorly. In medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known from the Widder Formation (Tioughniogan) of the Hamilton Group, Hungry Hollow, Arkona, Ontario, Canada. Struve (1959) showed an illustration of a cephalon from the Middle Devonian of Ontario, Canada, that he assigned to G. (G.) boothi. The specimen appears similar to G. boothi, but it is also very similar to G. widderensis. Because G. widderensis is the only species of Greenops described from Ontario, and because the specimen illustrated does not appear to be referable to Stummiana arkonensis (Stumm, 1953), also from the Hamilton Group of Ontario, Struve's (1959) specimen is assigned to G. widderensis. Struve (1959) also figured a hypostome from Ontario that he assigned to Greenops (Greenops?) calliteles (Green). It is conceivable that this hypostome belongs to Greenops widderensis, one of only two species of asteropyginines described from Ontario. The hypostome could also belong to Stummiana arkonensis (Stumm, 1953), and thus this hypostome is questionably assigned to either of these taxa. Recovery of articulated specimens with hypostomes for each of these species would make it possible to resolve this issue.

As discussed above under the heading of G. boothi, Stumm (1953) illustrated three specimens from the Widder Formation and possibly the Hungry Hollow Formation of the Michigan Basin in southwestern Ontario. These specimens do bear some resemblance to G. boothi, G. grabaui, and G. barberi. However, they belong to a new species that is restricted to the Michigan Basin, whereas G. boothi, G. grabaui, and G. barberi are restricted to the Appalachian Basin. Greenops widderensis can be discriminated from G. grabaui by the following characters: in G. widderensis a line from the posterior to the anterior edge of the eye forms an angle to a sagittal line that is approximately 10°, whereas in G. grabaui the angle is approximately 30°; in G. widderensis the distal tips of the pygidial lappets are pointed, whereas in G.

grabaui they are developed as blunt triangles; in G. widderensis the anterolateral margins of the first five pygidial axial rings are evenly convex, whereas in G. grabaui, when proceeding from the axial furrow to the midline, the anterior margin is straight, then arches anteriorly; and in G. widderensis the longest pygidial pleural lappet is the length (exsag.) of the length (sag.) of the first three pygidial axial rings, whereas in G. grabaui it is the length (sag.) of the first two pygidial axial rings. Characters for distinguishing G. barberi from G. widderensis include the condition of the terminal lappet, which in G. widderensis is subrectangular, broad anteriorly, and convex posteriorly, whereas in G. barberi it is a sharply pointed triangle, narrow anteriorly; and the condition of the genal spine, which in G. widderensis is flattened medially, is evenly inclined in G. barberi. Other characters for distinguishing G. widderensis from G. barberi are given under the heading of G. barberi.

Ludvigsen (1979) figured a cephalon and a complete individual from the Hamilton Group, Thedford, Ontario, Canada, that he assigned to G. boothi. As discussed above under the heading of G. boothi, these taxa must instead be treated as G. widderensis. In particular, they differ from G. boothi in the characters enumerated above in the section on G. boothi. For the same reasons, Ludvigsen's (1979: fig. 54b) reconstruction of G. boothi must instead be treated as a reconstruction of G. widderensis.

DELOOPS, NEW GENUS

Type Species: Greenops haasi Morzadec, 1983.

ASSIGNED TAXA: Greenops (Greenops) cf. boothi Arbizu, 1979: pl. 13, figs. 1, 2.

DIAGNOSIS: S3 convex anteriorly; S2 weakly declined posteriorly at lateral edge; S1 crescent shaped, medial part developed as smooth curve convex posteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; maximum of seven lenses in dorsoventral file on visual surface; facial suture lateral to eye, before flexing posteriorly, flexes anteriorly to point opposite S1; anterior margin of frontal glabellar lobe smoothly rounded; anterior ce-

phalic border pointed, developed as narrow lip; PMI absent; posterior and anterior margins of palpebrum deflected at angle to transverse line that is roughly equivalent; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 30°; distance between facial suture and frontal lobe constant from lateral edges of lobe to midline; tubercle on L0; cephalic axial furrows diverge more strongly anterior to S1 than posterior to S1; fixigenae with distinct bulge medially; genal spine extending back to fourth thoracic segment; medially genal spine developed as flattened ledge, laterally as an evenly inclined plane; medial edge of genal spine not significantly thickened, lateral margin parallels sagittal line, recurves at posterior end, interior margin of genal spine deflected laterally; tops of pygidial pleural segments flat; lateral margins of pygidial lappets curved, medial margins straight, distal tips blunt triangles; anterior margins of pygidial axial rings posterior to sixth axial ring smoothly convex anteriorly; 14 pygidial axial rings; pygidial axis constricts posterior to sixth axial ring; anterior and posterior parts of pygidial pleural segments of equal elevation; tops of pygidial pleural segments flat; pleural furrows moderately incised, short (exsag.); laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment; terminal pygidial lappet subrectangular, broad (tr.), projects further posteriorly than pleural lappets.

ETYMOLOGY: Named by combining *Delo* for the paleontologist who did some of the important early work revising and describing asteropyginines of North America and Europe with *Ops* for eyes. This name follows the etymological style of Delo (1935), who established several new genera and named these new taxa by combining the names of important paleontologists with the suffix *-ops*.

DISCUSSION: This genus contains two species, each of which had originally been treated as closely related to species of the genus *Greenops*. However, placing these taxa in the genus *Greenops* would have required the expansion of that genus into a particularly large and cumbersome taxonomic unit, and this

decision was not pursued. This genus represents one of a series of Armorican forms that bear a strong resemblance and have some phylogenetic affinity with the core species of the genus *Greenops* from North America, but none of the species of *Deloops* have a sister group relationship with *Greenops* to the exclusion of all other genera. Thus, these two species can no longer be treated as belonging to the genus *Greenops*, but their close affinity to taxa in that genus, and to those taxa in the genus *Stummiana*, which also contains a North American species, is important to point out.

Arbizu (1979) figured a cephalothorax and a pygidium from the Givetian of Spain that he assigned to *Greenops* (*Greenops*) cf. boothi. This species is nearly identical to Deloops haasi (Morzadec, 1983). The only significant difference between the two taxa is that the terminal pygidial lappet is relatively broader in D. haasi. Therefore, Arbizu's (1979) specimens are treated as a new species closely related to D. haasi, termed D. sp. aff. haasi.

Deloops haasi (Morzadec, 1983)

Greenops haasi Morzadec, 1983: 125, pl. 10, figs. 1-7, pl. 11, figs. 1-11. (See for complete synonymy.)

TYPES: Holotype defined in Morzadec (1983: pl. 11, fig. 6).

DIAGNOSIS: Same as for genus.

DISCUSSION: This species is known from the Massif Armorican, upper Givetian. It shares some similarities with some species traditionally assigned to Greenops by Morzadec (1983), in particular his G. osismorum, here Stummiana osismorum (Morzadec. 1983). However, the shared similarities appear to be primitive retentions or convergences. Similarities between S. osismorum and D. haasi include the condition of S3 and the overall shape of the genal spines. However, these taxa differ in a variety of features, including: the condition of the terminal pygidial lappet, which is relatively narrower (tr.) in S. osismorum; the cephalic axial furrows, which diverge more strongly anterior to S1 in D. haasi, but not in S. osismorum; the medial edge of the genal spine, which is flattened in D. haasi but not in S. osismorum:

the distance between the frontal lobe and the facial suture, which increases at the midline relative to laterally in *S. osismorum*, whereas this distance is constant in *D. haasi*; and by the possession of a greater number of pygidial axial rings (14) in *D. haasi*.

Originally, Morzadec (1983) treated D. haasi and S. osismorum as sister taxa. Here, the phylogenetic analysis suggests that they are closely related, but D. haasi is sister to a clade including S. osismorum and Kayserops.

STUMMIANA, NEW GENUS

Type Species: Greenops arkonensis Stumm, 1953.

Assigned Taxa: Greenops osismorum Morzadec, 1983.

DIAGNOSIS: S3 straight, anterolateral edge deflected posteriorly; S2 weakly declined posteriorly at lateral edge; maximum of seven lenses in a dorsoventral file on visual surface; anterior margin of frontal glabellar lobe smoothly rounded; distance between frontal lobe and facial suture at midline increases slightly relative to laterally; anteriormost portion of palpebrum opposite PMI; anterior cephalic border rounded in dorsal view; fixigena with distinct bulge medially; tubercle on L0; L3 flattened dorsally; line from posterior to anterior edge of eye forms a 30° angle to sagittal line; axial furrows more strongly divergent anterior to anterior half of L3; interior margin of genal spine evenly deflected laterally posteriorly; terminal pygidial lappet narrow (tr.), subrectangular, does not project as far posteriorly as fifth pair of pleural lappets; fifth pair of pleural lappets project furthest posteriorly; lateral margins of pygidial lappets curved, distal tips developed as blunt triangles; pygidial axis constricts posterior to fifth axial ring; anterior margins of pygidial axial rings posterior to constriction of pygidial axis smoothly convex anteriorly, anterior to constriction margins straight laterally, strongly arched medially anteriorly; pygidial pleural furrows moderately incised, short (exsag.); 11 axial rings.

ETYMOLOGY: This genus is named for the paleontologist Erwin Stumm, who contributed so much to our knowledge of the Middle Devonian faunas of the Michigan Basin.

DISCUSSION: This genus contains two spe-

cies, one from the Middle Devonian of the Michigan Basin of ENA and one from the Middle Devonian of Armorica. It is one of only two cases in the phylogeny of the Asteropyginae presented herein where a species from Armorica is sister to a species from ENA. In all other cases, a species from Armorica or ENA is sister to a larger clade from ENA or Armorica.

Both taxa in this genus had originally been assigned to the genus *Greenops*. However, the results presented here suggest that these two taxa must be excluded from that genus. Characters suggesting exclusion of species assigned to *Stummiana* from the *Greenops* clade include: the condition of the anteriormost portion of the palpebrum, which is opposite the PMI in *Stummiana* but opposite a point between the medial part of L3 and the PMI in *Greenops*; the fixigena, which has a distinct bulge medially in *Stummiana* but is evenly sloping from the axial furrow to the palpebrum in *Greenops*.

The two species in *Stummiana* share a large number of synapomorphies, but they can be discriminated on the basis of the relative position of the eyes, differences in the development of tubercles, and differences in the form of the pygidial pleural segments and lappets.

Stummiana osismorum Morzadec, 1983

Greenops osismorum Morzadec, 1983: 124, pl. 9, figs. 1-12.

?Greenops osismorum Morzadec, 1983: 124, pl. 9, fig. 13.

Types: Holotype defined in Morzadec (1983: pl. 9, fig. 3).

DIAGNOSIS: Lateral to eye facial suture before flexing posteriorly flexes anteriorly to point between distal margins of S1 and S2; anterior cephalic border in dorsal view rounded; S1 crescent shaped, medial portion smooth curve directed anteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; posterior and anterior margins of palpebrum deflected at an equivalent angle to transverse line; medial margins of pygidial pleural lappets straight; terminal pygidial lappet subrectangular, convex posteriorly; anterior and pos-

terior portions of pygidial pleural segments of equal elevation.

DISCUSSION: This species is known from the Givetian of Armorica. Morzadec (1983) suggested that this species was closely related to G. boothi, but the phylogenetic analysis conducted herein does not entirely support his conclusions. Stummiana osismorum in fact appears to be part of a larger clade (including several genera) that is sister to the four ENA taxa assigned to Greenops. Morzadec (1983) also suggested that this species was closely related to Arbizu's (1979: pl. 3, figs. 1, 2) Greenops sp. cf. boothi. However, as discussed above, these specimens appear to be more closely related to D. haasi on the basis of pygidial characteristics, and they are tentatively referred to as D. sp. aff. haasi until more detailed material can be obtained. Morzadec's (1983: pl. 9, fig. 13) is a pygidium that he questionably assigned to G. osismorum, because it appears to have slightly longer pleural lappets than the holotype. It does not appear to differ in the condition of any other characteristics, and until more detailed material is obtained, it is retained within S. osismorum.

Morzadec (1983) treated his G. osismorum as a direct lineal descendant of what he termed Greenops struvei. This direct ancestor-descendant relationship between the two taxa cannot be upheld herein. As discussed above, G. struvei must henceforth be assigned to a new genus, Modellops, and it is not particularly closely related to any of the species assigned to the genera Greenops or Stummiana. It shares a close, though not a sister group, relationship with Philipsmithiana, new genus. Differences between Philipsmithiana and Greenops are discussed under the heading of the former.

Stummiana arkonensis Stumm, 1953 Figure 8.8

Greenops arkonensis Stumm, 1953: 134, pl. 8, figs. 10–14.

Greenops aequituberculatus Stumm, 1953: 131, pl. 7, fig. 5, 6, non 4, pl. 8, figs. 6–8, non 5, 9. ?Greenops aequituberculatus (Stumm). Stumm, 1967: 115, pl. 2, fig. 6.

?Greenops (Greenops?) collitelus (Green). Struve, 1959: O480, fig. 379.5.

TYPES: Holotype defined in Stumm (1953: pl. 8, fig. 13).

DIAGNOSIS: Genal spine extends back to fifth thoracic segment; lateral margin of genal spine at posterior end straight; posterior edge of visual surface anterior to transverse line tangent to L0; fixigena with distinct bulge medially; tubercle on L0 present; cephalic axial furrows diverge at constant angle anterior and posterior to \$1; axial furrows at anterior half of L3 relative to posterior much less strongly divergent; isolated patches of tubercles present on L2 and L3; facial suture lateral to eye before flexing posteriorly flexes anteriorly to point opposite S2; S3 straight, at anterior end recurves posteriorly; posterior margin of palpebrum relative to anterior margin of palpebrum deflected at an angle to transverse line that is significantly larger; medial tubercle present on pygidial and thoracic axial rings; one row of circular fenestrae on anterior part of thoracic pleural segments; medial margins of pygidial pleural lappets curving; distal tips of pygidial lappets blunt triangles; posterior parts of pleural segments elevated above anterior parts; pygidial interpleural furrows arch more strongly posteriorly distally; medial margins of pygidial lappets curving laterally; pygidial pleural lappets long; 11 pygidial axial rings.

MATERIAL: AMNH 45249.

DISCUSSION: This species is known from the following Givetian localities in the Traverse Group, Michigan Basin, ENA: Gravel Point Formation (Cazenovian), Longispina emmetensis zone, Gravel Point west of Charlevoix, Michigan, on shore of Lake Michigan; the Alpena Limestone (Cazenovian), the Huron Point Cement Company, Alpena, Michigan, AMNH locality 3059; Dock Street Clay (Cazenovian), abandoned quarry of the Thunder Bay Limestone Company, eastern edge of Alpena, Michigan, AMNH locality 3062; the Arkona Shale (Cazenovian), Hungry Hollow on the Aux Sable River, north of Arkona, Ontario, AMNH locality 3053; the Widder Formation, Rock Glen, Arkona, Ontario, AMNH locality 3055; and the Norway Point Formation (Tioughniogan), bank of Thunder Bay River on Potter Farm, about 1 mile below Four Mile Dam, Michigan. Thus, this species is known from the Cazenovian and Tioughniogan of the Michigan Basin, based on the stratigraphic correlations presented in figure 2.

Stumm (1953) considered this species to be endemic to the Michigan Basin, and this conclusion appears to be correct. However, his assignment of this species to the genus Greenops can no longer be supported. Stummiana arkonensis can be distinguished from Greenops boothi by several characters, including the following: in G. boothi there are a maximum of six lenses pre dorsoventral file on the visual surface, whereas in S. arkonensis there are a maximum of seven lenses: in G. boothi the facial suture lateral to the eye before flexing posteriorly flexes anteriorly to point between the distal tips of S1 and S2, whereas in S. arkonensis it flexes to a point opposite the distal tip of S2; in G. boothi medially the anterior cephalic border in dorsal view is pointed, whereas in S. arkonensis it is rounded; in G. boothi the posterior margin of the palpebrum relative to the anterior margin is deflected at an angle to a transverse line that is roughly equivalent, whereas in S. arkonensis the angle the posterior margin forms with a transverse line is relatively larger; in G. boothi the axial furrows anterior to S1, relative to those posterior to S1, diverge more strongly, whereas in S. arkonensis they diverge at the same angle; in G. boothi the medial margins of the pygidial lappets are straight, whereas in S. arkonensis they are curved; in G. boothi the anterior and posterior parts of the pygidial pleural segments are of equal elevation, whereas in S. arkonensis the posterior part of the pygidial pleural segment is elevated above the anterior part; in G. boothi the tops of the pygidial pleural segments are flat, whereas in S. arkonensis they are rounded; in G. boothi in the medial region of adjacent pleural segments, the posterior region of the anterior segment is equal in length (exsag.) to the anterior part of the posterior segment, whereas in S. arkonensis the posterior region of the anterior segment is longer (exsag.) than the anterior part of the posterior segment; in G. boothi the terminal pygidial lappet is relatively broad, whreas in S. arkonensis it is relatively narrow; and in G. boothi there are two transverse rows of circular fenestrae on the anterior band of thoracic pleural segments, whereas in S. arkonensis there is only one row.

Stumm (1953) suggested that S. arkonensis was closely related to G. aequituberculatus Stumm, 1953. On some level, this is correct; however, Stumm's (1953) conception of G. aequituberculatus appears to have encompassed at least three distinct species: S. arkonensis, G. chilmanae, and Kayserops traversensis. Most of the paratypes of G. aequituberculatus including USNM 117872 should be referred to S. arkonensis by the following logic. The cephala Stumm (1953) assigned to Greenops traversensis and G. alpenensis are identical, and the species they comprise is assigned to the genus Kayserops and is discussed more fully below. The cephalon Stumm (1953: pl. 7, fig. 4) designated as a paratype of G. aequituberculatus shows no differences from the cephala designated the holotypes of G. alpenensis and G. traversensis. This suggests that this cephalon formerly designated G. aequituberculatus has been misassigned. However, the thoracopygidium and pygidia Stumm (1953) assigned to G. aequituberculatus appear to belong to S. arkonensis rather than to K. traversensis. The pattern of fenestrae on the thoracic pleural segments is serially repeated on the pleural segments of the pygidium, and the holotype of "Greenops" traversensis, here assigned to Kayserops, has a single row of circular fenestrae on the posterior band of the thoracic pleural segments, and this is also seen on the paratype of Kayserops traversensis. By contrast, examining the specimens used to establish G. aequituberculatus indicates that there are a few circular fenestrae on the posterior band of the thoracic pleural segments near the thoracic axis, not a row of fenestrae. This is the character state found in S. arkonensis. This character state is also found in some of the pygidia that Stumm (1953: pl. 8, figs. 6, 7) assigned to G. aequituberculatus, and these pygidia clearly differ from the pygidia of Kayserops traversensis and G. chilmanae Stumm, 1965, in the condition of many other characters. In addition, although the patterns of the fenestrae are not visible on the thoracopygidium or the pygidium in Stumm's (1953) plate 7, figures 5 and 6, the lappets, the condition of the interpleural furrows, and the number. form, and condition of the pygidial axial rings, i.e., all those characters of the pygidium used in the phylogenetic analysis, appear to be identical to those shown in Stumm's (1953) plate 8, figures 6 and 7, and the specimens are treated as conspecific. These character states are identical to what is found in S. arkonensis, both in Stumm's (1953) type material and in material housed at the AMNH. Therefore, these specimens formerly assigned to G. aequituberculatus are placed in S. arkonensis, and the species name G. aequituberculatus is no longer considered valid.

Stumm (1967: pl. 2, fig. 6) figured a pygidium in ventral view that he assigned to *Greenops aequituberculatus* but that is questionably referred to herein as *S. arkonensis*. Although not well preserved, it bears a terminal pygidial lappet very similar in shape to those known for *S. arkonensis*. In addition, the form of the pygidial lappets is very similar to what is known for *S. arkonensis*.

Struve (1959) figured a hypostome from Ontario that he assigned to *Greenops* (*Greenops*?) calliteles (Green). It is conceivable that this hypostome belongs to *Greenops widderensis*, one of only two species of asteropyginines described from Ontario. However, the hypostome could also belong to *Stummiana arkonensis* (Stumm, 1953), and thus this hypostome is questionably assigned to either of these taxa. Recovery of articulated specimens with hypostomes for each of these species would make it possible to resolve this issue.

The condition of S3 is treated as straight in S. arkonensis rather than convex anteriorly. Nearly the entire portion of S3 is straight, as it is in G. chilmanae and G. boothi. In these two taxa, at its anterior end, S3 is transverse, whereas in S. arkonensis, it recurves posteriorly. This is considered to be an autapomorphy of S. arkonensis rather than homologous with the structure of S3 in such taxa as D. haasi (Morzadec, 1983), which appear to have a large portion of S3 convex.

ARMORIGREENOPS, NEW GENUS

TYPE SPECIES: Armorigreenops leoi, new species.

Assigned Taxa: Armorigreenops sp. aff. leoi.

DIAGNOSIS: S3 straight, anterolateral edge

transverse; S2 weakly declined posteriorly at lateral edge; S1 crescent shaped, medial portion developed as straight line directed anteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0: distance between frontal lobe and facial suture at midline relative to lateral margins increases slightly; frontal glabellar lobe smoothly rounded in dorsal view; anterior cephalic border in dorsal view pointed, developed as narrow lip; facial suture lateral to eye, before flexing posteriorly, flexes anteriorly to point opposite S1; line from posterior to anterior edge of eye forms a 30° angle to sagittal line; axial furrows anterior to S1 diverge at same angle as furrows posterior to S1, less strongly divergent anterior to anterior half of L3; fixigena evenly sloping from axial furrow to palpebrum; posterior and anterior margins of palpebrum deflected at an angle to transverse line that is roughly equivalent; lateral margin of genal spine parallels sagittal line, at posterior end recurves posteriorly; genal spine long flange extending back to sixth thoracic segment, medial edge significantly thickened relative to lateral edge, plane from medial to lateral ledge evenly inclined, interior margin parallels lateral margin before deflecting posteriorly; tubercle present on medial part of pygidial and thoracic axial rings; tops of pygidial pleural segments flat; lateral margins of pygidial lappets curved, medial margins straight, distal tips developed as blunt triangles; anterior and posterior parts of pygidial pleural segments of equal elevation; laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment; terminal pygidial lappet developed as blunt triangle.

ETYMOLOGY: Name derived by combining Armorica, the region the genus is known from, with Greenops to denote its resemblance to those taxa herein assigned to Greenops that hail from ENA.

DISCUSSION: This genus contains a single described species that shows some similarity to taxa treated here as belonging to *Greenops*, and this species was originally treated as a form closely related to *G. boothi* by Gandl (1972). However, this taxon differs

from *Greenops boothi* in the condition of the following characters: 5, 8, 13, 22, 26, 28, 30, 32, 35, 36, 39, 41, 44, 59, 61, and 63.

On the basis of the phylogenetic analysis conducted herein, Armorigreenops leoi is more closely related to taxa such as Kayserops Delo, 1935. Thus, this species must be excluded from Greenops, or Greenops would be paraphyletic. It would also be possible to place A. leoi in a greatly expanded Greenops that contained a variety of new and old genera including Kayserops. This latter taxonomic decision is not followed in order to retain the structure in the Asteropyginae, and instead the species is placed in a new genus.

Armorigreenops leoi, new species

Greenops (Greenops) sp. Gandl. Carls and Gandl, 1968: 462.

Greenops (Greenops) sp., cf. boothi (Green). Gandl, 1972: 137, pl. 14, figs. 1-3.

TYPES: Holotype Gandl's (1972: pl. 14, fig. 1), Senckenberg Museum, Frankfurt (SMF) 21681.

DIAGNOSIS: Same as for the genus.

ETYMOLOGY: Named for my other grandfather, Leo Lieberman, who recently passed away.

DESCRIPTION: Cephalic length (sag.) 60% of width. Axial furrow nearly straight anterior to S0, diverging forward at about 30°, narrow, shallow. Cranidial anterior border short exsagittally and medially, lying in constant dorsoventral plane, pointed medially in dorsal view. Preglabellar furrow moderately incised and narrow. Cranidial anterior border lengthens slightly medially, developed as narrow lip. Facial suture anterior to eyes flexes laterally then medially, with smoothly convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly then posteriorly laterally. Glabellar length (sag.) equal to width (tr.) across frontal lobe; frontal lobe ellipsoid, about 60% of length of glabella; L1-L3 flat (sag.); frontal lobe at same level as posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI not visible. S3 moderately incised, straight, anterolateral edge flexes posteriorly, branches diverging at about 140°. Sagittal region of L3-L2 nearly flat (tr.), lateral lobes weakly

declined abaxially. L3 wedge shaped, lengthening distally, outer edge posteriorly flexes laterally, anterior end flexing less strongly laterally. S2 weakly convex anteriorly, moderately incised, shallow adjacent to axial furrow, weakly declined posteriorly at lateral edge. S1 deep, crescent shaped, medial part developed as straight line directed anteriorly, contacting axial furrow. L1 short (exsag.), flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. So convex anteriorly medially. deep. L0 55% of width (tr.) of frontal lobe, moderately arched (tr.), longest medially. Large Eye Index 0.40. Anterior edge of eye adjacent to axial furrow opposite antero-distal corner of L3; posterior edge of eye opposite L1, posterior to transverse line tangent to L0; eye raised slightly above frontal lobe. Fixigena evenly sloping from axial furrow to internal margin of eye. Postocular fixigenal field 15% of length (exsag.) of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse, anterior margin nearly straight. Genal spine long flange extending back to sixth thoracic segment, cephalic lateral border furrow bisects spine; medial and lateral edges of genal spine evenly inclined; medial edge thickened; lateral margin of genal spine parallels sagittal line; lateral margin of genal spine at posterior end recurves; interior margin of genal spine parallels lateral margin before deflecting laterally. Lateral border furrow narrow, shallow, lateral border weakly widens posteriorly. Anterior margin of cephalic doublure bluntly pointed. Prosopon of fine tubercles.

Thorax of 11 segments. Axial ring about 30% width (tr.) of thorax, of equal length (exsag., sag.), at distal ends and medially flexed forward. Ring furrow of equal length, moderately incised, flexing forward medially and distally. Anterior margin of ring well defined sagittally, inclined posterodorsally, most elevated near posterior edge; raised prominent tubercle on medial portion of axial ring. Articulating half ring set slightly below axial ring. Proximal part of pleura transverse, approximately horizontal; anterior segments rather evenly convex (tr.), outer part of pleurae more strongly flexed downward, straight in posterior segments. Posterior band long

(exsag.), anterior margin straight, deflected at constant small angle posteriorly relative to transverse line, laterally, length of posterior band increases, becomes planar; posterior surface declined posterodorsally; anterior surface declined anterodorsally, with concave depression. Anterior pleural band short (exsag.), medially transverse, deflecting strongly posteriorly past approximate midpoint, at roughly same level as posterior band, pinching out distally.

Pygidium broadly triangular in outline excluding marginal lappets, length (sag.) about 55% of width (tr.) excluding lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 20° anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Axis 25% of pygidial width (tr.) anteriorly, with 11 rings; rings of nearly equal length (exsag., sag.) distally and medially, longest medially; anterior and posterior edge of axial rings anterior to fifth axial ring from midline to distal edge straight, then strongly arch anteriorly. Ring furrows moderately incised. Lateral margins of pygidial lappets curved, medial margins straight, distal tips developed as blunt triangles. Axial terminus triangular, pointed posteriorly, projecting further posteriorly than fifth pair of pleural lappets, narrow, 1.1 times width of posterior portion of pygidial axis; postaxial region 30% of pygidial length (sag.). Pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; anterior and posterior pleural segments of equal elevation; tops of segments flat; pleural furrows deeper than interpleural furrows, moderately incised, short (exsag.). Laterally pygidial interpleural furrows arching more strongly than medially. In medial region of adjacent pleural segments, posterior region of anterior segment equal in length (exsag.) to anterior part of posterior segment.

DISCUSSION: This species is known from the Givetian of northeastern Spain. It bears a strong resemblance to *Greenops boothi*, and Gandl (1972) originally suggested that it might indeed belong to that species. However, the taxa differ in the condition of several characters, discussed above, and it must be assigned to a new genus.

Haas (1970) figured a pygidium from Mid-

dle Asturia in northern Spain, at Vaca de Luanco in the lower Candas Kalke, that he assigned to *Greenops boothi*. This species is very similar to *A. leoi*, differing chiefly in the number of pygidial axial rings, 12 in Haas's species versus 11 in *A. leoi*. In addition, in Haas's species the anterior margin of the first five pygidial axial rings follows the same general course found in *A. leoi* and in all other species of the genus *Greenops*; however, the anterior flexion of the axial ring medially is not so strong as in *A. leoi* or in the other species of *Greenops*. Haas's (1970) figured pygidium is tentatively treated as a new species, *Armorigreenops* sp. aff. *leoi*.

Arbizu (1979) also figured a specimen that he referred to as Greenops (G.) cf. boothi. However, this specimen is not conspecific with A. leoi, counter to Morzadec's (1983) conclusions. In particular, they differ in the condition of the distal tips of the pygidial pleural lappets; in the shape of the terminal pygidial lappet; in the condition of S3, which are convex anteriorly in Arbizu's specimen; and in the structure of the genal spine, which is flattened medially and smoothly arched laterally in Arbizu's figured specimen. The species appears to be closely related to G. haasi Morzadec, 1983, here Deloops haasi, and is discussed above under the heading of that genus.

Gandl (1972) treated A. leoi (his G. (G.) sp. cf. boothi) as closely related to the form he called G. (Neometacanthus?) sp. M. aff. aequituberculatus. This species was discussed above under the heading of the genus Neometacanthus, and it appears that Gandl's pygidium cannot be assigned to that genus. The pygidium also differs from Armorigreenops in the condition of several characters. In particular, the anterior margin of the five anteriormost pygidial axial rings, when proceeding from the axial furrows to the midline, is smoothly convex anteriorly instead of being straight and then flexing strongly anteriorly; the anterior margins of the pygidial axial rings posterior to the fifth pygidial axial rings are straight instead of convex anteriorly; the pygidial interpleural furrows do not flex more strongly posteriorly laterally; and the tops of the pygidial pleural segments are rounded instead of flat. The evolutionary affinities of Gandl's (1972) isolated pygidium are difficult to ascertain; however, it seems clear that it is not closely related to Armorigreenops leoi, new species. Gandl (1972) also asserted that Neometacanthus stellifer was a close relative of A. leoi. As discussed above, Neometacanthus is sister to a clade containing Rhenops and Asteropyge, and thus N. stellifer is not a particularly close relative of A. leoi.

KENNACRYPHAEUS, NEW GENUS

TYPE SPECIES: Kennacryphaeus harrisae, new species.

DIAGNOSIS: S3 straight medially, anterolateral edge transverse; S1 developed medially as smooth curve convex posteriorly; posterior edge of visual surface posterior to transverse line tangent to anterior edge of L0; maximum of eight lenses per dorsoventral file on visual surface; facial suture lateral to eye, before flexing posteriorly, flexes anteriorly to point between distal tips of S1 and S2; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; distance between facial suture and frontal lobe at midline relative to distance between facial suture and frontal lobe at lateral margins of frontal lobe equivalent; medially anterior cephalic border in dorsal view rounded; spine on L0; anterior cephalic border medially not deflected anteriorly beyond lateral margins of cephalon; PMI developed as rounded depression; posterior margin of palpebrum relative to anterior margin deflected at an angle to transverse line that is roughly equivalent; line from posterior to anterior edge of eyes forms an angle to sagittal line that is approximately 30°; change in cephalic topography where librigena meets lateral border furrow insignificant; genal spine medially developed as flattened edge, laterally evenly inclined; medial edge of genal spine significantly thickened relative to lateral edge; raised ridge on dorsal surface of genal spine, at juncture of posterior border furrow and lateral border furrow, is smoothly curved, convex distally; axial furrows anterior to S1 relative to posterior to S1 diverge at same angle; visual surfaces present within sagittal line drawn tangent to lateral margins of frontal lobe; interior margin of genal spine parallels lateral margin before deflecting laterally; interior

margin of genal spine more strongly deflected laterally posteriorly; tubercles present on medial portion of pygidial and thoracic axial rings; terminal pygidial lappet developed as blunt triangle, does not project as far posteriorly as fifth pair of pleural lappets; lateral margins of pygidial lappets curved, medial margins straight; distal tips of lappets pointed; anterior margins of first five pygidial axial rings straight; pygidial axis constricts posterior to fifth axial ring; pygidial pleural field flanking posterior portion of pygidial axis faintly excavated; anterior and posterior parts of pygidial pleural segments of equal elevation; tops of pygidial pleural segments rounded; pleural furrows moderately incised, short (exsag.); laterally pygidial interpleural furrows flex more strongly posteriorly; in medial region of adjacent pleural segments posterior region of anterior segment equal in length (exsag.) to anterior portion of posterior segment; 14 pygidial axial rings; anterolateral margins of first five pygidial axial rings going from axial furrows to midline straight, then strongly arched anteriorly; longest pygidial lappet short (exsag.), length from proximal to distal tip equal to length (sag.) of first two pygidial axial rings.

ASSIGNED TAXA: None (monotypic).

ETYMOLOGY: Named for Malcolm McKenna, Department of Vertebrate Paleontology, AMNH, whose advice and assistance over the years are greatly appreciated.

DISCUSSION: This genus was created to accommodate *K. harrisae*, new species. The genus *Kennacryphaeus* is sister to the genus *Kayserops* Delo, 1935. *Kennacryphaeus harrisae* could have been lumped with *Kayserops*, but this would have required substantial modification of the diagnosis of *Kayserops*. Instead, a new monotypic genus is erected.

Kennacryphaeus harrisae, new species Figures 8.5, 8.7, 27

Greenops (Greenops) boothi (Green). Eldredge, 1971a: 54, text fig. 1.

TYPES: Holotype AMNH 45298, a cephalon from the Mottville Member (Cazeonovian), Skaneateles Formation, Pratt's Falls, 2.4 miles northwest of Pompey Center, New York. Paratypes AMNH 45302, a pygidium from the Mottville Member, road cut on U.S.

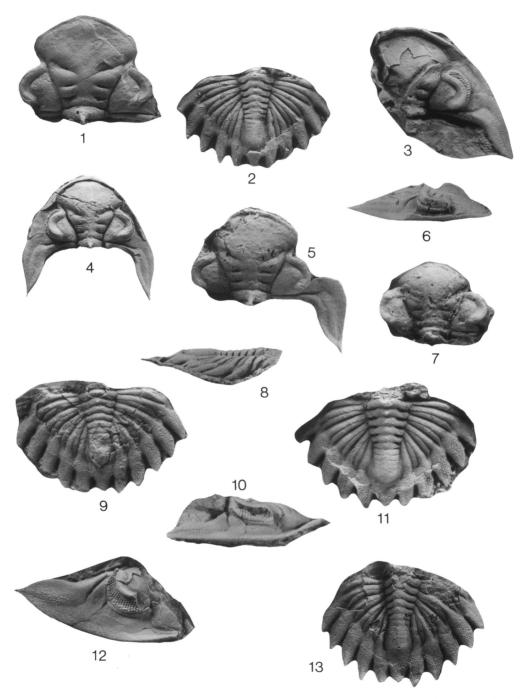


Fig. 27. Kennacryphaeus harrisae, new species, Mottville Member, Skaneateles Formation, New York. 1. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45304, dorsal view of cranidium, ×2.1. 2. U.S. Route 20, road cut just west of Pompey Center, AMNH 45302, dorsal view of paratype pygidium, ×2.3 3, 10. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45300, dorsal and lateral views of partial cephalon, ×2.5. 4, 6. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45298, dorsal and lateral views of holotype cephalon, ×2. 5. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45301, dorsal view of partial cephalon, ×2.2. 7. Pratt's Falls, 2.4 miles northwest of

Route 20, just west of Pompey Center, New York; AMNH 45297, a pygidium from the same locality as the holotype; and AMNH 45299, a pygidium from the same locality as the holotype.

DIAGNOSIS: Same as for genus.

ETYMOLOGY: Named for Ms. Meg Harris, who contributed some of the specimens used to diagnose this species.

MATERIAL: AMNH 29006, 45248, 45297–45306.

DESCRIPTION: Cephalic length (sag.) 55% of width (tr.). Axial furrow nearly straight anterior to S1, diverging forward at about 30°, narrow, shallow. Cranidial anterior border short exsagittally and medially, lying in constant dorsoventral plane, rounded medially in dorsal view. Preglabellar furrow moderately incised and narrow. Cranidial anterior border lengthens slightly medially, developed as narrow lip. Facial suture anterior to eyes flexes laterally then medially with smooth convex margin laterally. Posterior branch of facial suture flexes weakly anteriorly, then posteriorly laterally. Glabellar length equal to width across frontal lobe; frontal lobe ellipsoidal, about 50% of length of glabella; L1-L3 flat; frontal lobe with same elevation as posterior glabellar region; anterior part of frontal lobe moderately declined forward, smoothly rounded in dorsal view. PMI developed as rounded depression. S3 moderately incised, straight, anterolateral edge flexes posteriorly, branches diverging at about 140°. Sagittal region of L2 and L3 nearly flat (tr.), lateral lobes weakly declined abaxially. L3 wedge shaped, lengthening distally, outer edge posteriorly flexes laterally, anterior end flexing less strongly laterally. S2 weakly declined posteriorly at lateral edge, moderately incised, shallow adjacent to axial furrow. S1 deep, crescent shaped, medial portion developed as smooth curve convex posteriorly, contacting axial furrow. L1 short, flexed forward strongly medially and weakly abaxially, depressed only slightly below L2 laterally. S0 of nearly uniform length, convex anteriorly at medial portion in dorsal view, moderately incised. L0 45% of width (tr.) of frontal lobe, moderately arched (tr.), longest sagittally medially, bearing spine. Large Eye Index 0.45. Anterior edge of eye adjacent to axial furrow opposite anterodistal corner of L3; posterior edge of eye opposite L1, posterior to transverse line tangent to anterior edge of L0; eye raised above frontal lobe. Fixigena evenly sloping from axial furrow to palpebrum. Postocular fixigenal field 15% of length (sag.) of eye. Librigenal field moderately declined to lateral border furrow. Posterior border furrow nearly transverse medially, laterally flexing anteriorly, anterior margin nearly straight, shallow distally. Cephalic lateral border furrow bisects genal spine; medially genal spine flattened ledge, laterally evenly inclined; medial edge thickened; lateral margin of genal spine at posterior end straight; interior margin of genal spine parallels lateral margin before deflecting laterally. Lateral border furrow narrow, shallow. Anterior margin of cephalic doublure rounded. Prosopon of fine tubercles.

Thorax not known.

Pygidium broadly triangular in outline excluding marginal lappet. Length (sag.) about 60% of width (tr.) excluding lappets, with five pairs of pleural lappets. Axial furrows shallow, narrow, converging at 20° anterior to fifth pygidial axial ring, roughly parallel posterior to fifth pygidial axial ring. Anterior margins of pygidial axial rings posterior to constriction of pygidial axis straight, anterior to constriction laterally straight, medially arched anteriorly. Axis 35% of pygidial width (tr.) anteriorly, with 14 rings. Terminal pygidial lappet blunt triangle, narrow, does not project as far posteriorly as other lappets; postaxial region 20% of pygidial length

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Pompey Center, AMNH 45305, dorsal view of small cranidium, ×5. 8, 9. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45297, lateral and dorsal view of paratype pygidium, ×2.5. 11. U.S. Route 20, road cut just west of Pompey Center, AMNH 45306, dorsal view of pygidium, ×2.6. 12. Pratt's Falls, 2.4 miles northwest of Pompey Center, AMNH 45303, lateral view of partial cephalon, ×2.6. 13. Pratt's Falls, 2.4 miles northwest of Pompey, AMNH 45299, dorsal view of paratype pygidium, ×2.5.

(sag.). Lateral margins of pygidial lappets curved, medial margins straight, distal tips pointed. Tops of pygidial pleural segments rounded. Pleural furrows moderately incised, short (exsag.), laterally flex more strongly posteriorly.

DISCUSSION: This species comes from the following localities in the Cazenovian (Givetian) of the Appalachian Basin, ENA: Mottville Member (Cazenovian), Skaneateles Formation, road cut, Route 20, west of Pompey Center, New York; Mottville Member, Pratt's Falls, New York; ?Mottville Member, Fitch Road, New York; Stafford Limestone, Skaneateles Formation, road cut, Route 488, north of the bridge that crosses Flint Creek, Phelps, New York.

Eldredge (1971a) originally treated this species as identical to Greenops boothi. However, the two taxa can be distinguished in the condition of several characters, including the following: in G. boothi the interior margin of the genal spine is evenly deflected laterally all the way back, whereas in K. harrisae the interior margin parallels the lateral margin before deflecting laterally; in G. boothi there is a maximum of six lenses per dorsoventral file on the visual surface, whereas in K. harrisae there is a maximum of eight lenses; in G. boothi the distance between the frontal lobe and the facial suture at the midline relative to laterally increases slightly, whereas in K. harrisae it is constant; in G. boothi there is a single tubercle on L0, whereas in K. harrisae there is a spine; in G. boothi a plane from the medial to the lateral edges of the genal spine is evenly inclined, whereas in K. harrisae medially the genal spine is developed as a flattened ledge; in G. boothi the medial edge of the genal spine is not significantly thickened relative to the lateral edge, whereas in K. harrisae the medial edge is significantly thickened relative to the lateral edge; in G. boothi the axial furrows anterior to S1 deflect more strongly laterally relative to the furrows posterior to S1, whereas in K. harrisae they are equally divergent anterior and posterior to S1; in G. boothi the anterior margins of the pygidial axial rings posterior to the constriction of the pygidial axis are smoothly convex anteriorly, whereas in K. harrisae they are straight; in G. boothi the tops of the pygidial pleural segments are flat, whereas in *K. harrisae* they are rounded; in *G. boothi* there are 11 pygidial axial rings, whereas in *K. harrisae* there are 14 pygidial axial rings; in *G. boothi* the terminal pygidial lappet is subrectangular anteriorly and convex posteriorly, whereas in *K. harrisae* it is a bluntly pointed triangle; and in *G. boothi* the longest pygidial lappet is relatively long (exsag.), whereas in *K. harrisae* it is relatively short, equal to the length (sag.) of the first two pygidial axial rings.

Kennacryphaeus harrisae is the only species of asteropyginine described from the Skaneateles Formation (Cazenovian) of the Hamilton Group of the Appalachian Basin, and it appears to be restricted to the lower part of this formation.

Kennacryphaeus harrisae does show similarity with Bellacartwrightia phyllocaudata. In particular, they both have: the medial portion of the genal spine significantly thickened relative to the lateral edge; the interior margin of the genal spine more strongly deflected laterally posteriorly; the medial portion of S1 developed as a smooth curve convex posteriorly; the frontal lobe smoothly rounded in dorsal view; the distance between the frontal lobe and facial suture at the midline relative to laterally constant; a spine on the medial portion of L0; the shape of the terminal pygidial lappet triangular, and not projecting as far as the other lappets; the medial margins of the pygidial lappet straight; the tops of the pygidial pleural segments rounded; and laterally the pygidial interpleural furrows flexing more strongly posteriorly. However, these two taxa differ in the condition of several characters. In particular, in K. harrisae S3 is straight with the anterolateral edge transverse, whereas in B. phyllocaudata S3 is sinusoidal; in K. harrisae S2 is weakly declined posteriorly at its lateral edge, whereas in B. phyllocaudata S2 is roughly transverse; in K. harrisae the posterior edge of the visual surfaces is posterior to a transverse line tangent to the anterior edge of L0, whereas in B. phyllocaudata it is anterior to such a line; in K. harrisae there are a maximum of eight lenses per dorsoventral file on the visual surfaces, whereas in B. phyllocaudata there are a maximum of nine lenses; in K. harrisae the anterior cephalic border is rounded in dorsal view, whereas in B. phyllocaudata it is point-

ed; in K. harrisae the anterior cephalic border medially is not deflected far laterally beyond the lateral margins of the glabella, whereas in B. phyllocaudata it is deflected far anteriorly; in K. harrisae a line from the posterior to the anterior edge of the eve forms an angle to a sagittal line that is approximately 30°, whereas in B. phyllocaudata the angle is approximately 5°; in K. harrisae medially the genal spine is a flattened ledge, whereas in B. phyllocaudata a plane from the medial to the lateral edges of the genal spine is evenly inclined; in K. harrisae the interior margin of the genal spine parallels the lateral margin before deflecting laterally, whereas in B. phyllocaudata it just deflects laterally; in K. harrisae the axial furrows anterior to S1 relative to posterior to S1 diverge at the same angle, whereas in B. phyllocaudata they diverge more strongly; in K. harrisae the axial furrows at the anterior half of L3 relative to the posterior half of L3 are equally divergent, whereas in B. phyllocaudata the axial furrows are less strongly divergent at the anterior half of L3; in K. harrisae there are tubercles on the medial portions of the pygidial and thoracic axial rings, whereas in B. phyllocaudata there are spines; in K. harrisae the anterior margins of the pygidial axial rings posterior to the constriction of the pygidial axis are straight, whereas in B. phyllocaudata they are smoothly convex anteriorly; in K. harrisae the pygidial axis constricts posterior to the fifth axial ring, whereas in B. phyllocaudata it constricts posterior to the sixth ring; in K. harrisae the pygidial pleural field flanking the posterior portion of the pygidial axis is faintly excavated, whereas in B. phyllocaudata it is prominently excavated; in K. harrisae the pygidial pleural furrows are moderately incised and medially short (exsag.), whereas in B. phyllocaudata they are relatively long (exsag.); in K. harrisae in the medial region of adjacent pleural segments, the posterior region of the anterior segment is equal in length (exsag.) to the anterior part of the posterior segment, whereas in B. phyllocaudata the posterior region is longer (exsag.); in K. harrisae there are 14 pygidial axial rings, in B. phyllocaudata there are 15 or 16; in K. harrisae the terminal pygidial lappet is a blunt triangle, whereas in B. phyl-

locaudata it is a sharp triangle; in K. harrisae the terminal pygidial lappet is relatively narrow, whereas in B. phyllocaudata it is relatively broad; and in K. harrisae the longest pygidial lappet is short, with its length (exsag.) equal to the length (sag.) of the first two pygidial axial rings, whereas in B. phyllocaudata it is long (exsag.), with the length equal to the length (sag.) of the first three pygidial axial rings.

Thus, although there are a fair number of character states that *K. harrisae* and *B. phyllocaudata* share, there are also many in which they differ. Phylogenetic analysis indicates that it is more parsimonious to view the characters they differ in as being shared with other taxa than it is to view the characters they share as synapomorphies, and according to the cladogram in figure 4 they are distantly related. Thus, their shared features must be viewed as either plesiomorphy or homoplasy. However, it is important to point out that these taxa may converge on each other in certain aspects of their morphology.

KAYSEROPS DELO, 1935

Type Species: Cryphaeus kochi Kayser, 1883.

ASSIGNED TAXA: Kayserops inflatus Morzadec, 1983, Kayserops, new species, aff. inflatus Morzadec, 1983, Greenops traversensis Stumm, 1953, and ?Cryphaeus kerfornei Pichard, 1930.

DIAGNOSIS: S3 convex anteriorly; L2 and L3 prominently inflated; isolated patches of tubercles present on L3; eyes substantially elevated above glabella; line from posterior to anterior edge of eye forms 30° angle with sagittal line; anterior cephalic border does not deflect far beyond lateral margins of frontal glabellar lobe; S1 crescent shaped, medially developed as smooth curve convex posteriorly; anterior margin of frontal glabellar lobe smoothly rounded in dorsal view; distance between frontal lobe and facial suture increases slightly from lateral margins to midline; anterior cephalic border pointed medially; PMI absent; fixigena evenly sloping from axial furrow to palpebrum; posterior margin of palpebrum relative to anterior margin deflected at angle to transverse line that is roughly equivalent; demarcation between librigena and lateral border furrow weak; tubercle on L0; medial edge of genal spine significantly thickened relative to lateral edge; plane from medial to lateral edges of genal spine evenly inclined; lateral margin of genal spine parallels sagittal line; axial furrows less strongly divergent at anterior half of L3 than at posterior half; interior margin of genal spine parallels lateral margin before deflecting laterally; medial tubercles on pygidial and thoracic axial rings present; posterior part of pygidial pleural segment elevated above anterior part; tops of pygidial pleural segments rounded; terminal pygidial lappet narrow, triangular; pygidial lappets flanking terminal lappet project further posteriorly than terminal lappet; distal tips of lappets pointed; anterior margins of pygidial axial rings posterior to fifth ring smoothly convex anteriorly; pygidial axis constricts posterior to fifth axial ring; pygidial pleural furrows moderately incised, short (exsag.); anterolateral margins of first five pygidial axial rings do not arch strongly anteriorly; pygidial lappets long (exsag.); five pairs of pygidial pleural lappets present.

DISCUSSION: The species in this genus are known from the upper Emsian of Germany and the Armorican Massif, and from the Givetian of the Michigan Basin of ENA. When first erected by Delo (1935), this genus was thought to consist of a single species. However, it now appears that additional taxa can be assigned to Kayserops. These are figured in Morzadec (1983) and Stumm (1953). There exist several additional taxa that Gandl (1972), Arbizu (1979), and Smeenk (1983) assigned to Kayserops but that can no longer be retained within that genus. These taxa are separated by several nodes on the cladogram in figure 4 from those taxa treated as Kayserops herein on the basis of phylogenetic and morphological evidence, and they are discussed above under the headings of the genera Bradocryphaeus Haas and Mensink. 1969, Hallandclarkeops, new genus, and Braunops, new genus.

A slightly emended diagnosis is presented to that of Delo (1935) and Struve (1959), as phylogenetic analysis indicated that a few species could be assigned to *Kayserops*. These taxa could have been assigned to separate genera, but instead they were treated as

belonging to a single genus. Some of the species that have been assigned to Kayserops herein were not subjected to phylogenetic analysis. One of these species, K. kerfornei (Pichard), appears to be very similar morphologically to K. inflatus. However, they do differ by the former's possession of a very long spine on the medial portion of the anterior cephalic border. Kayserops kerfornei also may lack the tubercle on L0, a diagnostic feature of the genus (the occipital ring of this taxon was too poorly preserved to ascertain this for sure). This species also lacks another character treated as diagnostic for Kayserops, the presence of a tubercle on the medial region of the thoracic and pygidial axial rings. Thus, the taxon appears to differ in characters diagnostic for the genus as presented here. However, it is conceivable that phylogenetic analysis would show this species as sister to the rest of Kayserops, and therefore it would be reasonable to place the species within an emended Kayserops. This could only be done if this taxon were incorporated into the phylogenetic analysis. This was not done herein, and thus it is only possible to questionably place this species within Kayserops and point out the characters in which it may depart from the rest of the genus.

Unfortunately, the type of the genus, *K. kochi*, also was not available for study. However, on the basis of figures and diagnoses, it appears to be very similar to *K. inflatus*, and it seems reasonable to use the phylogenetic position of the latter to anchor the phylogenetic topology of the genus *Kayserops*. It would be unlikely, but not impossible, that analysis of this species instead of, or in addition to, *K. inflatus* might change the topological position of the genus *Kayserops*. Such analysis was not possible at this time but will be pursued at a later date.

On the basis of the phylogenetic analysis conducted herein, *Kayserops* appears to be part of a large clade that includes exponents originally assigned to *Greenops*, as well as some new taxa, and this clade is in turn sister to *Greenops*. Several authors have commented on the phylogenetic relationships of this genus. Some of their conclusions differ from those promulgated herein, as their conceptions of the genus *Kayserops* do not co-

incide with that developed here using cladistic evidence. Haas (1970) claimed that Kavserops was a subgenus of Comura and was either derived from a paraphyletic Comura (Comura) or part of a basal polytomy along with all the subgenera in Comura (see fig. 5A). Comura was not treated as paraphyletic herein, nor for that matter were any of the other genera of the Asteropyginae. In addition, species of Comura were not subjected to phylogenetic analysis herein, making the issue of paraphyly of Comura impossible to test. The cladogram in figure 4 shows that Kayserops is distantly related to Tolkienia, a close relative of Comura, and Haas's (1970) phylogenetic topology does not appear to be upheld in the case of Kayserops. Gandl (1972) suggested that Kayserops was closely related to the genera Neometacanthus and Paracryphaeus (see fig. 5B). This contention is not supported by the analysis conducted herein, though it must be stated that Gandl's (1972) concept of Kayserops corresponds to the genus Braunops herein. However, some of the members of the large clade that includes Kayserops, such as Stummiana osismorum (Morzadec) and S. arkonensis (Stumm), as well as Deloops haasi (Morzadec), had traditionally been assigned to the genus Greenops, which had been treated, incorrectly, as closely related to Neometacanthus by some authors. Arbizu (1979) suggested that Kayserops was derived from a paraphyletic Pseudocryphaeus and was relatively closely related to Greenops and Neometacanthus (see fig. 5C), and Morzadec (1983) essentially advocated the same phylogenetic placement of Kayserops (see fig. 5E). The relatively close relationship between Kayserops and Greenops predicted by these authors appears to be supported. Smeenk (1983) suggested that Kayserops was derived from a paraphyletic Paracryphaeus (see fig. 5D). This does not appear to be supported. However, Smeenk's (1983) conception of Kayserops corresponds to what is treated herein as *Braunops*.

This genus is known from the upper Emsian of Armorica and the Givetian of ENA.

Kayserops inflatus Morzadec, 1983 Kayserops inflatus Morzadec, 1983: 127, pl. 14, figs. 1-8, pl. 15, figs. 1-4, 6. TYPES: Holotype defined in Morzadec (1983: pl. 15, fig. 1).

DIAGNOSIS: S3 convex anteriorly: PMI not visible; line from posterior to anterior edge of eye forms 30° angle to sagittal line; prosopon of fine granules; isolated coarse tubercles present on L3 and palpebrum; librigena not visible in dorsal view between lateral margin of eye and lateral border furrow; broad ledge of anterior cephalic border not visible anterior to anterolateral portions of frontal lobe; maximum of eight lenses per dorsoventral file on visual surface: lateral to eye, facial suture before flexing posteriorly flexes anteriorly to point opposite S1; tubercle on L0 present; L3 distinctly swollen; genal spine evenly inclined from medial to lateral edge; lateral margin of genal spine at posterior end straight; medial margins of pygidial pleural lappets curving laterally; distal tips of pygidial lappets pointed; tops of pygidial pleural segments rounded; posterior part of pygidial pleural segments elevated above anterior part; terminal pygidial lappet triangular and narrow; at pygidial margins surface of pygidium dorsally inflates at contact of lappets and margins; 13 pygidial axial rings.

DISCUSSION: This species is known from the upper Emsian of the Armorican Massif. One of the pygidia that Morzadec (1983: pl. 15, fig. 5) figured appears to differ from the other pygidia of K. inflatus in a few features. These taxa possess two fewer pygidial axial rings, have broader pygidial lappets, and have the pygidial interpleural furrows deflected less strongly posteriorly laterally. As all the other pygidia Morzadec (1983) figured are identical, and these are the more abundant, these were used to code K. inflatus for character analysis. The pygidium in Morzadec (1983: pl. 15, fig. 5) appears similar to pygidia typical of K. inflatus in several other respects, and it is treated as Kayserops new species aff. inflatus Morzadec. Kayserops inflatus is also probably closely related to K. kerfornei (Pichard, 1930). Morzadec (1983) suggested that K. inflatus is also closely related to Gandl's (1972) "Kayserops" new species N. This species appears to lack several of the characters diagnostic of Kayserops and cannot be considered to be closely related to that genus. It may belong to Philipsmithiana, new genus. The relationships of Gandl's (1972) species were not considered further at this time.

The illustrations in Morzadec (1983), particularly plate 14, figure 4a, suggest that circular fenestrae on the thoracic pleural segments are present in this taxon, and thus this species was coded as having these structures. However, until the actual specimens can be obtained, this cannot be determined with certainty.

Kayserops traversensis (Stumm, 1953) Figure 12.5

Greenops traversensis Stumm, 1953: 132, pl. 8, figs. 1-4.

Greenops alpenensis Stumm, 1953: 133, pl. 7, fig. 7.

Greenops aequituberculatus Stumm, 1953: 131, pl. 7, fig. 4, pl. 8, fig. 5.

TYPES: Holotype an external mold of cephalothorax, USNM 117871, from the Traverse Group, Gravel Point Formation, upper blue shale, abandoned "Bell" quarry and ledges on shore about 2 miles east of Bay Shore, Emmet County, Michigan, designated by Stumm (1953: pl. 8, figs. 1, 3, 4). Paratype an external mold of pygidium, UMMP 28668, upper Gravel Point Formation, Emmet County, Michigan.

DIAGNOSIS: S3 convex anteriorly; S2 weakly declined posteriorly at lateral edge; tubercle present on L0; PMI absent; line from posterior to anterior edge of eye forms an angle to sagittal line that is approximately 30°; prosopon of coarse tubercles; genal spine evenly inclined laterally; cephalic axial furrows diverge at same angle anterior and posterior to S1; maximum of seven lenses per dorsoventral file on visual surface: broad ledge of anterior cephalic border absent anterior to anterolateral edges of glabella; one transverse row of circular fenestrae on anterior and posterior bands of thoracic pleural segments; circular fenestrae present at lateral margins of thoracic axial rings, and scattered about medial part of thoracic axial rings; lateral margins of pygidial lappets curved; distal tips of lappets pointed; pygidial pleural field flanking posterior part of axis moderately excavated; posterior part of pygidial pleural segment elevated above anterior part; tops of pygidial pleural segments rounded; in medial region of adjacent pleural segments posterior region of anterior segment longer (exsag.) than anterior part of posterior segment; terminal pygidial lappet blunt triangle, narrow anteriorly; 14 pygidial axial rings.

MATERIAL: AMNH 45250.

DISCUSSION: This species is known from the following stratigraphic horizons and localities in ENA from the Middle Devonian Traverse Group of the Michigan Basin: the Gravel Point Formation (uppermost Cazenovian), upper blue shale, zone 6, abandoned "Bell" quarry and ledges on shore about 2 miles east of Bay Shore, Emmet County, Michigan; the Dock Street Clay (lowermost Tioughniogan), abandoned quarry of the Thunder Bay Limestone Company, eastern edge of Alpena, Alpena County, Michigan; the Norway Point Formation (Tioughniogan), shale bank on south side of Thunder Bay River on Potter Farm about 1 mile below Four Mile Dam, Alpena County, Michigan; and the Thunder Bay Limestone (Taghanic), bluffs on northeastern shore of Partridge Point, 4 miles south of Alpena, Alpena County, Michigan. Thus, in terms of New Yorkian stages, it ranges from the latest Cazenovian into the Taghanic.

Stumm (1953) appears to have parsed a single species into three separate species: his Greenops aequituberculatus, G. traversensis, and G. alpenensis. The holotypes of G. aequituberculatus, G. alpenensis, and G. traversensis, all cephala, are identical and they are treated as conspecific. However, the single glabella and two pygidia that Stumm (1953) illustrated as paratypes of G. aequituberculatus are referable to G. arkonensis Stumm, 1953, and are discussed in greater detail above under the heading of that species. Stumm (1953) illustrated only two specimens of G. alpenensis, a cephalon and pygidium. The pygidium (Stumm, 1953: pl. 7, fig. 8), a paratype, differs significantly from all other asteropyginines except for ?Greenops comis (Hall and Clarke, 1888), which is known from a single pygidium found in the Onondaga Limestone of Ontario, and it is treated as closely related if not conspecific with that species. Both species share the small, triangular, postaxial process, as well as the relatively short (exsag.) pygidial pleural lappets, the narrow, convex overall form of the pygidium, and the pygidial axis with approximately 11 axial rings. However, at this time no generic appellation can be given to this species because of the many autapomorphies it bears, as well as the absence of a thorax and cephalon. It is clear that it should not be assigned to Greenops because it lacks most of the diagnostic pygidial characters of that genus. It is possible that Stumm's (1953: pl. 7, fig. 8) pygidium belongs to the same species as the cephalon of G. alpenensis, but this does not appear likely on the basis of the following logic. Although Stumm (1953) claimed that his G. alpenensis and G. traversensis differed in the condition of the shape of the tubercles present on the head shield, the tubercles appear identical, and any dissimilarities probably can be accounted for by differences in preservation, as these different cephala are known from rocks with different lithologies. Moreover, they match each other in all features of preserved cephalic morphology. In addition, the holotype of what was referred to as G. traversensis also preserves a thorax that bears a prominent medial tubercle on each of the axial rings. In all asteropyginine taxa with medial tubercles on the thoracic axial rings, these are repeated in a serially homologous fashion on the pygidial axial rings. In Stumm's (1953: pl. 8, fig. 3) paratype pygidium of *G. traversensis*, such medial tubercles are present, whereas they are not developed on the paratype pygidium of *G. alpenensis* that closely resembles ?*Greenops comis*. Thus, it seems likely that the specimens that Stumm (1953) illustrated and designated holotypes and paratypes of *G. traversensis* all belong to the same species.

As mentioned above, the holotype cephalon of G. alpenensis is identical with the holotype cephalon of G. traversensis. Because the types of G. traversensis represent a homogeneous series whereas those of G. alpenensis do not, the latter species is considered no longer valid, and the pygidium is assigned to ?G. sp. aff. comis, while the cephalon is treated as identical with traversensis, which is treated herein as a species of the genus Kayserops. The species Greenops aequituberculatus Stumm, 1953, was discussed in detail above under the heading of Greenops arkonensis Stumm, 1953, and it is no longer considered a valid species. Most of the specimens of G. aequituberculatus appear referable to G. arkonensis; however, one, the glabella, which was a paratype, UMMP 25438, is identical with the glabella of the holotype specimen of Kayserops traversensis, and the two should be treated as conspecific.

MORPHOLOGICAL THEMES IN THE EVOLUTION OF THE ASTEROPYGINAE

On the basis of evidence presented by Edgecombe (1993), the Acastidae Delo, 1935, which includes the Acastinae Delo, 1935, the "Acastavinae" Struve, 1958a, and the Asteropyginae Delo, 1935, is sister to the Calmoniidae Delo, 1935. Several authors have discussed the diversification of the calmoniids as an important example of an adaptive radiation (e.g., Eldredge and Cracraft, 1980; Fortey and Owens, 1990), although the adaptive character of at least part of the calmoniid radiation has been called into question by some (e.g., Lieberman, 1993). Still, it is clear that a substantial taxic proliferation occurred in the diversification of the calmoniids. In one of the clades nested within the Calmoniidae, the "Metacryphaeus group," most of

the morphological differences between taxa are associated with variation in characters of the glabella, such as its furrows and overall shape. As argued cogently by Eldredge (1971a), these morphological features seem to govern food gathering and processing. Thus, any changes in these features through phylogenetic time may indicate changes in mode of feeding. Therefore, the taxic proliferation that occurred in the "Metacryphaeus group" appears to involve a proliferation of feeding or digestive modes.

Although not typically characterized as an adaptive radiation, the asteropyginines are clearly a diverse group of Devonian trilobites, and in their evolution there is a clear predilection toward differentiation in certain

characters or character complexes. In particular, in the asteropyginines the greatest variation is found in characters of the pygidium, such as the shape of the lappets and the axial terminus, in the shape and size of the genal spine, and in the array of fenestrae and furrows on the thoracopygidium. These consistently important character complexes, in terms of their utility for providing resolution in phylogenetic analysis, may indicate the possibility that changes in modes of sediment stabilization and burrowing frequently oc-

curred in the Asteropyginae. Whether or not these character differences involve adaptive diversification is another question that would require functional analysis as well as statistical tests, phylogenetic tests, and assumptions about character optimization, and the answers to such questions are not considered herein. However, the consistent manifestation of differences in these particular character complexes in the Asteropyginae is worthy of mention.

BIOGEOGRAPHIC PATTERNS AND THE APPEARANCE OF THE ASTEROPYGINAE IN EASTERN NORTH AMERICA

As mentioned in the introduction, one of the reasons for studying the phylogeny of the Asteropyginae was to assess how many times the subfamily had invaded ENA. We also wished to pinpoint where these invasions came from, and to deduce whether any invasion of lineages occurred from ENA into other biogeographic regions. Biogeographic patterns in the Asteropyginae can be deduced in a preliminary way by using parsimony algorithms to optimize biogeographic states to ancestral nodes on the phylogeny of the Asteropyginae. With this information, we can witness transformations between biogeographic areas and deduce the number of times that ENA was invaded from a primitively Armorican lineage. Armorica is treated as comprising Spain, the Massif Armorican, Germany, and northern Africa, as well as Afghanistan and Turkey. Eastern North America and northern Gondwanaland (Venezuela) were also treated as separate biogeographic areas. Taking the phylogeny of the Asteropyginae given in figure 4, the biogeographic state of each terminal taxon is mapped onto the tree, with a "0" for Armorica, a "1" for ENA, and a "2" for northern Gondwanaland. Fitch optimization (Fitch, 1971) was used to ascertain the biogeographic states of ancestral nodes on the phylogeny in figure 4. The optimizations, as well as the states of the terminal taxa, are shown in figure 28. These results indicate that there may have been four invasions by the Asteropyginae from Armorica into ENA. In some ways this is not surprising, because the asteropyginines in ENA do not comprise a single monophyletic clade, but it points out how there were major shifts in biogeographic areas occupied by the subfamily, as well as an asymmetrical transformation vector leading from Armorica to ENA.

It is instructive to consider which lineages mark the biogeographically invasive lineages and, in addition, using phylogenetic information in conjunction with information about the first appearance of species in the fossil record, it is possible to put some temporal bounds on when these different invasions occurred. This method follows the ghost-lineage method utilized by Edgecombe (1992). One of the invasions marks the establishment of the genus Bellacartwrightia in ENA; using the ghost-lineage method, this invasion must have occurred by the Eifelian. Species in this genus first appear in the Eifelian Onondaga Limestone, so there is good congruence between the predicted and actual stratigraphic patterns. Another of these invasions marks the establishment of the genus Greenops in ENA (in this analysis, unlike others, we conclude that Greenops is restricted to ENA). According to the ghost-lineage method, based on phylogenetic patterns, this invasion occurred some time between the upper Emsian and the Givetian. Members of this genus actually first appear in the Givetian portion of the Hamilton Group. Thus, there is a gap in the record between the predicted and the actual time of invasion. An-

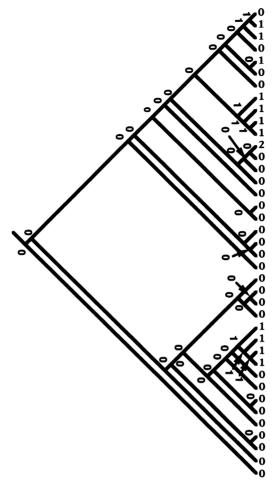


Fig. 28. The mapping of biogeographic states of taxa onto the single most parsimonious tree shown in figure 4. Nodes were optimized using Fitch's (1971) algorithm. "0" indicates presence in Armorica, "1" in Eastern North America, and "2" in northern Gondwanaland.

other of the invasions marks the appearance of Kennacryphaeus harrisae and Kayserops traversensis in ENA, and the ghost-lineage method indicates that this invasion must have occurred by the Upper Emsian. However, the genus Kennacryphaeus does not appear in the fossil record until the latest Eifelian or earliest Givetian. Thus, again there appears to be a significant gap in the history of this genus between the time it invaded from Armorica, based on stratigraphic evidence, and the time it actually appeared. The final invasion marks the appearance of Stummiana arkonensis in ENA, and it occurred during the Givetian, based on phylogenetic topology. This species first appears in the Givetian Traverse Group, and thus it shows close congruence between predicted and actual first occurrence.

The results from this analysis indicate that the timing of invasive events is spread out over the interval from the upper Emsian to the Givetian. Thus, the invasions cannot be consigned to one unique biogeographic event, such as the initiation of the Acadian Orogeny. Instead, the invasions span an interval including both the initial development of the Acadian Orogeny (in the upper Emsian) and the reintensification of that orogeny (in the late Eifelian) (timing of orogenic events from Ettensohn [1985]). This time interval also corresponds to an interval when

there were multiple episodes of relative sealevel rise, probably related to the orogenic events (Johnson et al., 1985; Hallam, 1992). This pattern of multiple episodes of invasion within a clade spanning the upper Emsian to the Givetian corresponds to the pattern recognized to have occurred in the proetid trilobites by Lieberman (1994). In the proetids there were several invasions from different regions into ENA, and these invasions occurred at different times throughout the Lower and Middle Devonian and could not be consigned to a single eustatic or tectonic event. For further discussion of the driving forces behind these biogeographic changes, see Lieberman and Eldredge (1996).

In addition, it is likely that other invasions of asteropyginines may have occurred from Armorica into Laurentia. For instance, some species from the Givetian of the Illinois Basin may be referable to the genus Neometacanthus. Unfortunately, they were under study by another author and could not be incorporated into the phylogenetic analysis conducted herein. If they are actually species of Neometacanthus, it would imply at least one more instance of invasion from Armorica into ENA. Finally, Stumm's (1953) Greenops sp. A may constitute another invasion from Armorica into ENA. This species is probably referable to Tolkienia, new genus, and all other species in that genus are restricted to Armorica. Thus, there are at least four, and perhaps as many as six, invasions by independent asteropyginine lineages into ENA.

The results of this analysis also indicate that at least once an ENA lineage reinvaded Armorica. The latter conclusion matches that of Oliver (1977), who suggested that rugose coral lineages common to ENA had invaded Armorica during the Middle Devonian. The invasions by asteropyginines from ENA into Armorica must have occurred by the Upper Emsian, based on phylogenetic topology. Thus, there appears to be some asymmetry in the pattern of invasion between Armorica and ENA. Perhaps this is not surprising, as Armorican asteropyginine diversity is much greater than that of ENA, and the subfamily is primitively present within Armorica and primitively absent from ENA.

These conclusions about the timing of the

invasion of Armorican lineages into ENA and the timing of reinvasion of ENA lineages back into Armorica are a bit troubling, for they suggest that a portion of the fossil record of the asteropyginines is obscured, as most members of the subfamily do not appear in ENA until the latest Eifelian or Givetian. It is possible that lineages persisted in other biogeographic regions for long periods of time without being preserved in the fossil record.

It is clear from phylogenetic analysis that many invasions of Armorican asteropyginine lineages into ENA occurred, but the exact timing of these invasions cannot always be exactly ascertained, although they were almost certainly between the upper Emsian and the Givetian. In addition, almost all of the other trilobites in the order Phacopida that occur in the Hamilton Group are forms that invaded from Armorica. This pattern of invasion by Hamilton Group trilobites from Armorica into ENA suggests that some of the taxa that populate faunas usually come from different biogeographic areas. In addition, the timing of these invasions was piecemeal. with different elements arriving at different times and certain lineages reinvading ancestral biogeographic regions. This suggests that faunas are often put together from immigrant lineages but, more importantly, it indicates faunas tend to be put together piecemeal contra the conclusions of Brett and Baird (1995).

For some other trilobite lineages that occur in the Hamilton Group, phylogenetic analyses have also been conducted, and these results further reiterate how transition times for invasions from Armorica into ENA show broad scatter. In particular, Phacops Delo, 1935, analyzed by Eldredge (1972) and Burton and Eldredge (1974), invaded ENA from Armorica around the Eifelian-Givetian transition, and the genus Basidechenella Kayser, 1880, analyzed by Lieberman (1994), may have invaded from Armorica into ENA in the Emsian. Thus, it is obvious that those trilobite taxa that appear in the Hamilton Group represent a series of invasions, some of which significantly predate the appearance of the first Hamilton Group sediments.

In summary, recruitment of the trilobite fauna of the Hamilton Group was piecemeal, with different taxa arriving from different regions at different times, counter the predictions of Brett and Baird (1995). This points out that faunal origination, at least in the case

of the Hamilton Group, cannot be consigned to a single evolutionary event (Lieberman, 1994; Lieberman and Eldredge, 1996).

REFERENCES

Arbizu, M.

1979. Asteropyginae (Trilobita) du Dévonien des Montagnes cantabriques (Espagne).
Bull. Soc. Géol. minéral. Bretagne (C)
9: 59-102.

Babcock, L., and S. Speyer

1987. Enrolled trilobites from the Alden Pyrite Bed, Ledyard Shale (Middle Devonian), western New York. J. Paleontol. 61: 539-548.

Babin, C., D. Goujet, H. Lardeux, A. Lejal-Nicol, F. Lethiers, P. Morzadec, Y. Plusquellec, and M. Weyant

1977. La formation des schistes de Porsguen (Dévonien superieur de la rade de Brest, Massif Armoricain). Lithologie, flore, faune. Ann. Soc. Géol. Nord 96: 333-346.

Beaumont, C.

1988. Orogeny and stratigraphy: numerical models of the Paleozoic in the eastern interior of North America. Tectonics 7: 389-416.

Brett, C. E.

1986. The Middle Devonian Hamilton Group of New York: an overview. In C. E. Brett (ed.), Dynamic stratigraphy and depositional environments of the Hamilton Group (Middle Devonian) in New York State, Part 1. New York State Mus. Bull. 457: 1-4.

Brett, C. E., and G. C. Baird

1986. Symmetrical and upward shallowing cycles in the Middle Devonian of New York State and their implications for the punctuated aggradational cycle hypothesis. Paleoceanography 1: 431–445.

1995. Coordinated stasis and evolutionary ecology of Silurian to Middle Devonian faunas in the Appalachian Basin. In D. H. Erwin and R. L. Anstey (eds.), New approaches to speciation in the fossil record, 285–315. New York: Columbia Univ. Press.

Brett, C. E., K. B. Miller, and G. C. Baird

1990. A temporal hierarchy of paleoecologic processes within a Middle Devonian epeiric sea. *In* C. Brett (ed.), Dynamic stratigraphy and depositional environ-

ments of the Hamilton Group (Middle Devonian) in New York State, Part 1. New York State Mus. Bull. 457: 129–156.

Burmeister, H.

1843. Die Organization der Trilobiten. London: Ray Society.

Burns, J.

1991. Fossil collecting in the Mid-Atlantic states. Baltimore: Johns Hopkins Univ. Press.

Burton, C. J., and N. Eldredge

1974. Two new subspecies of *Phacops rana* (Trilobita) from the Middle Devonian of north-west Africa. Palaeontology 17: 349-363.

Carls, P.

1979. Emsian-Eifelian stratigraphy of the eastern Iberian chains. Guide book of the field trip. Intl. Symp. Dev. Syst., Spain, pp. 35-39.

Carls, P., and J. Gandl

1968. The Lower Devonian of the eastern Iberian chains (NE Spain) and the distribution of its Spiriferacea, Acastavinae, and Asteropyginae. Intl. Symp. Dev. Syst. Calgary 1967, 2: 453–464.

Conrad, T. A.

1840. Third Annual Report of the Paleontological Department of the Survey. Rep. New York Geol. Surv., 199–207.

Cooper, G. A., C. Butts, K. E. Caster, G. H. Chadwick, W. Goldring, E. M. Kindle, E. Kirk, C. W. Merriam, F. M. Swartz, P. S. Warren, A. S. Warthin, and B. Willard

1942. Correlation of the Devonian sedimentary formations of North America. Bull. Geol. Soc. Amer. 53: 1729-1794.

Cooper, M. R.

1982. A revision of the Devonian (Emsian-Eifelian) Trilobita from the Bokkeveld Group of South Africa. Ann. S. Afr. Mus. 89: 1-174.

Delo, D. M.

1935. A revision of the phacopid trilobites. J. Paleontol. 9: 402-420.

1940. Phacopid trilobites of North America. Geol. Soc. Am. Spec. Pap. No. 29.

Destombes, J., and J.-L. Henry

1987. Trilobites Calmoniidae de l'Ordovicien

supérieur du Maroc et les origines de la Province Malvino-Cafre. Lethaia 20: 129-139.

Edgecombe, G. D.

- 1991. Systematic studies on phacopide trilobites. Diss., Columbia Univ., New York.
- 1992. Trilobite phylogeny and the Cambrian-Ordovician "Event": cladistic reappraisal. *In* M. J. Novacek and Q. D. Wheeler (eds.), Extinction and phylogeny, 144–177. New York: Columbia Univ. Press.
- 1993. Silurian acastacean trilobites of the Americas. J. Paleontol. 67: 535-548.

Eldredge, N.

- 1971a. Patterns of cephalic musculature in the Phacopina (Trilobita) and their phylogenetic significance. J. Paleontol. 45: 52-67.
- 1971b. The allopatric model and phylogeny in Paleozoic invertebrates. Evolution 25: 156–167.
- 1972. Systematics and evolution of *Phacops rana* (Green, 1832) and *Phacops iowensis* Delo, 1935 (Trilobita) from the Middle Devonian of North America. Bull. Am. Mus. Nat. Hist. 147: 45-114.
- 1979. Cladism and common sense. *In J. Cracraft and N. Eldredge (eds.)*, Phylogenetic analysis and paleontology, 165–198. New York: Columbia Univ. Press.
- 1985. Unfinished synthesis. New York: Oxford Univ. Press.
- 1989. Macroevolutionary dynamics. New York: McGraw-Hill.
- 1993. Fossils. New York: Harry N. Abrams.
- 1995. Reinventing Darwin. New York: Wiley.

Eldredge, N., and L. Branisa

1980. Calmoniid trilobites of the Lower Devonian *Scaphiocoelia* Zone of Bolivia, with remarks on related species. Bull. Am. Mus. Nat. Hist. 165: 181-289.

Eldredge. N., and J. Cracraft

1980. Phylogenetic patterns and the evolutionary process. New York: Columbia Univ. Press.

Eldredge, N., and A. R. Ormiston

1979. Biogeography of Silurian and Devonian trilobites of the Malvinokaffric Realm. *In J. Gray and A. J. Boucot (eds.)*, Historical Biogeography, Plate Tectonics, and the Changing Environment, 147–167. Corvallis: Oregon State Univ. Press.

Ellison, R. L.

1965. Stratigraphy and paleontology of the Mahantango Formation in south-central

Pennsylvania. Pennsylvania Geol. Surv., 4 ser., 298 pp.

Emmrich, H. F.

1839. De trilobitis. Dissertation petrefactologica quam concensu et auctoritate amplissimi philosophorum ordinis. Berlinis, 56 pp.

Ettensohn, F. R.

1985. The Catskill Delta complex and the Acadian Orogeny: a model. In D. L. Woodrow and W. D. Sevon (eds.), The Catskill Delta. Geol. Soc. Am. Spec. Pap. 201: 39–50. Boulder, Colorado: Geological Society of America.

Feist, R.

1991. The Late Devonian trilobite crisis. Hist. Biol. 5: 197–214.

Fitch, W. M.

1971. Toward defining the course of evolution: minimum change for a specific tree topology. Syst. Zool. 20: 406–416.

Fortey, R. A., and R. M. Owens

1990. Evolutionary radiations in the Trilobita.

In P. D. Taylor and G. P. Larwood (eds.), Major evolutionary radiations, 139–164. Syst. Assoc. Spec. Vol. 42. Oxford, UK: Clarendon Press.

Gandl, J.

 1972. Die Acastavinae und Asteropyginae (Trilobita) Keltiberiens (NE-Spanien).
 Abh. Senckenberg. Naturforsch. Gesell.
 530: 1-184.

Gauthier, J. A., R. Estes, and K. de Queiroz

1988. A phylogenetic analysis of Lepidosauromorpha. In R. Estes and G. Pregill (eds.), Phylogenetic relationships of the lizard families. Essays commemorating Charles L. Camp, 15–98. Palo Alto, California: Stanford Univ. Press.

Grabau, A. W., and W. H. Shimer

1910. North American index fossils: invertebrates, vols. 1 and 2.

Green, J.

1832. A monograph of the trilobites of North America. Philadelphia.

1837. Description of several new trilobites. Am. J. Sci. (1) 32: 343–349.

Haas, W.

1968. Trilobiten aus dem Silur und Devon von Bithynien (NW-Turkei). Palaeontographica 130: 60–207.

1970. Zur Phylogenie und Systematik der Asteropyginae und beschreibung einiger neuer arten (Phacopacea, Trilobita). Senckenberg. Leth. 51: 97-131.

Haas, W., and H. Mensink.

1969. Asteropyginae aus Afghanistan (Trilo-

bita). Akad. Wissen. Lit. Mainz, Abh. Math.-Natur. kl. 6: 155-211.

Hall, J.

1861. Descriptions of new species of fossils from the upper Helderberg, Hamilton, and Chemung Groups. New York State Cabinet Nat. Hist., Ann. Rep. No. 14.

1862. Descriptions of new species of fossils from the upper Helderberg, Hamilton, and Chemung Groups. New York State Cabinet Nat. Hist., Ann. Rep. No. 15.

1876. Illustrations of Devonian fossils. Gastropoda, Pteropoda, Cephalopoda, Crustacea, and corals of the upper Helderberg, Hamilton, and Chemung Groups. Paleontology, New York. Albany.

Hall, J., and J. M. Clarke

1888. Descriptions of the trilobites and other Crustacea of the Oriskany, upper Helderberg, Hamilton, Portage, Chemung, and Catskill Groups. Natural history of New York. N.Y. Geol. Surv., Paleontol., vol. 7.

Hallam, A.

1992. Phanerozoic sea-level changes. New York: Columbia Univ. Press.

Hawle, I., and A. J. C. Corda

1847. Prodrowm einer monographie der Bohemischen Trilobiten. Abh. Bohem. Gesell. Wiss., Prague, vol. 5.

Hickerson, W. J.

1992. Trilobites from the late Givetian Solon Member, Little Cedar Formation of eastern Iowa and northwestern Illinois. In J. Day and B. J. Bunker (eds.) The stratigraphy, paleontology, depositional and diagenetic history of the Middle–Upper Devonian Cedar Valley Group of central and eastern Iowa, 123–139. Iowa Dep. Nat. Res., Guidebook No. 16.

Holloway, D. J., and J. V. Neil

1982. Trilobites from the Mount Ida Formation (Late Silurian-Early Devonian), Victoria. Proc. R. Soc. Victoria 94: 133-154.

Huffner, E.

 Beitrage zur Kenntnis des Devons von Bithynien. Jahr. Preuss. Geol. Landes-Anst. 33: 305-395.

Johnson, J. G., G. Klapper, and C. A. Sandberg
 1985. Devonian eustatic fluctuations in Euramerica. Bull. Geol. Soc. Am. 96: 567-587.

Kayser, E.

1878. Die Fauna der altesten devon-Ablagerungen des Harzes. Abh. Geol. spez.

Preuss., Band 2, H. 4, 295 pp. 1880. *Dechenella*, eine Devonische Gruppe der gattung *Phillipsia*. Z. Deutsch. Geol. Gesell. 32: 703–707.

1883. Die orthocerasschiefer zwischen Balduinstein und Laurenberg an der Lahn. Jahrb. Preuss. Geol. Landes. 1–56.

1889. Die Fauna des Hauptquarzits und der zorger schiefer des Unterharzes. Abh. Preuss. Geol. Landes. 1: 1-139.

Kesling, R. V., and R. B. Chilman

1975. Strata and megafossils of the Middle Devonian Silica Formation. Mich. Mus. Paleontol. Pap. Paleontol. No. 8.

Kesling, R. V., R. T. Segall, and H. O. Sorensen 1974. Devonian strata of Emmet and Charlevoix Counties, Michigan. Mich. Mus. Paleontol. Pap. Paleontol. No. 7.

Kielan, Z.

 Les trilobites mesodevoniens des monts de Sainte-Croix. Paleontol. Polon. No. 6.

Kirchgasser, W. T., W. A. Oliver, Jr., and L. V. Rickard

1985. Devonian series boundaries in the eastern United States. Cour. Forschungs. Senckenb. 75: 233–260.

Lesperérance, P., and S. Desbiens

1995. Selected Ordovician trilobites from the Lake St. John district of Quebec and their bearing on systematics. J. Paleontol. 69, Mem. 42: 1-19.

Lieberman, B. S.

1993. Systematics and biogeography of the "Metacryphaeus group" Calmoniidae (Trilobita, Devonian), with comments on adaptive radiations and the geological history of the Malvinokaffric Realm. J. Paleontol. 67: 549–570.

1994. Evolution of the trilobite subfamily Proetinae Salter, 1864 and the origin, diversification, evolutionary affinity and extinction of the Middle Devonian proetid fauna of Eastern North America. Bull. Am. Mus. Nat. Hist. 223: 176 pp.

Lieberman, B. S., G. D. Edgecombe, and N. Eldredge

1991. Systematics and biogeography of the "Malvinella group," Calmoniidae (Trilobita, Devonian). J. Paleontol. 65: 824–843.

Lieberman, B. S., and N. Eldredge

1996. Trilobite biogeography in the Middle Devonian: geological processes and analytical methods. Paleobiology 22: 66–79.

Ludvigsen, R.

1979. Fossils of Ontario part 1: the Trilobites. Toronto: Royal Ontario Museum.

Morzadec, P.

- 1969. Le Dévonien de la rive nord de la rivière du Faou (Finistère). Étude stratigraphique, étude des trilobites. Bull. Soc. Géol. Minér. Bretagne 1968: 1-58.
- 1971. Quelques Asteropyginae (Trilobites, Phacopacea) des schistes et calcaires a Athyris undata (Dévonien inferieur du Massif Armoricain). Palaeontographica 138: 166-184.
- 1976. Trilobita. In Incursion hercynienne dans les faunes rhenanes du Dévonien Inferieur de la Rade de Brest (Massif Armoricain). Palaeontographica (A) 153: 35-45.
- 1981. Les trilobites. In P. Morzadec, F. Paris, and P. Racheboeuf (eds.), La tranchee de la Lezais, Emsien superieur du Massif Armoricain, 279–287. Mem. Soc. Géol. Minér. Bretagne 27: 294 pp., Rennes.
- 1983. Trilobites du Dévonien (Emsien-Famennien) de la rade de Brest (Massif Armoricain). Palaeontographica (A) 181: 103-184.
- 1992. Évolution des Asteropyginae (Trilobita) et variations eustatiques au Dévonien. Lethaia 25: 85–96.

Morzadec, P, and M. Arbizu

1978. Kayserops ? cantarmoricus nov. sp., Trilobite Asteropyginae du Dévonien moyen armoricain et cantabrique. Geobios 11: 925-931.

Oehlert, D.

1877. Sur les fossiles dévoniens du department de la Mayenne. Bull. Soc. Géol. France (3) 5: 578-603.

Oliver, W. A., Jr.

- 1976. Biogeography of Devonian rugose corals. J. Paleontol. 50: 365-373.
- 1977. Biogeography of late Silurian and Devonian rugose corals. Palaeogeogr., Palaeoclimatol., Palaeoecol. 22: 85-135.

Pichard, P.

1930. Contribution a l'étude du sous-genre Cryphaeus dans le Massif Armoricain. Bull. Soc. Geol. Minér. Bretagne 9: 70– 134.

Pillet, J.

- 1954. La classification des Phacopacea (Trilobites). Bull. Soc. Géol. France (6) 3: 817–839.
- 1958. Contribution a l'étude de quelques Asteropyginae. Bull. Soc. Géol. France (6) 8: 3-20.

- Contribution a l'étude des faunes de trilobites du Zemmour. Ann. Fac. Sci. Univ. Dakar 6: 93-119.
- 1969. Un nouveau genre de Trilobite dévonien en Afghanistan (*Neocalmonia* nov. gen., Calmoniinae, Trilobite). C. R. Somm. Soc. Géol. France 5: 144-145.

Ramsköld, L.

1991. The perforated trilobite *Laethoprusia* gen. nov., and the phylogeny of *Koneprusia* and *Isoprusia* (Odontopleuridae, Koneprusiinae). Trans. R. Soc. Edinburgh Earth Sci. 82: 125–141.

Ramsköld, L., and G. D. Edgecombe

1993. Silurian (Ludlow) acastid trilobites from Gotland and Scania, Sweden. Paläontol. Z. 67: 261–285.

Reed, F. R. C.

1925. Some new Silurian trilobites. Geol. Mag. 62: 73.

Richter, R.

- 1909. Beitrage zur Kenntnis devonischer Trilobiten aus dem Rheinischen Schiefergebirge. Vorbericht zu einer Monographie der Trilobiten der Eifel. Diss., Marburg.
- 1912. Beitrage zur Kenntnis devonischer Trilobiten. I. Die Gattung Dechenella und einige verwandte Forment. Abh. Senckenberg. Naturforsch. Gesell. 31: 239-340.
- 1916. Die Entstehung der Abgerollten "Dalaider versteinerungen" und das alter ihrer Mutterschichten. Jahrb. Preuss. Geol. Landes-Anst. 37: 247–259.

Richter, R., and E. Richter

- 1926. Die Trilobiten des Oberdevons. Beitrage zur Kenntnis devonischer Trilobiten. IV. Abh. Preuss. Geol. Landes- Anst. Neue Fol. 99: 1-314.
- 1939. Neue Beitrage zur Kenntnis der Geologie, Palaeontologie und Petrographie der Umgegend von Konstantinopel, 4, Trilobiten aus dem Bosphorus-Gebeit. Abh. Preuss. Geol. Landes-Anst. Neue Fol. 190: 1-47.
- 1943. Trilobiten aus dem Devon von Marokko. Senckenbergiana 26: 116–199.
- 1948. Neometacanthus n. nom. (pro Metacanthus Corda 1847). Senckenbergiana 29: 107.
- 1950. Arten der Dechenellinae (Trilobita). Senckenbergiana 31: 151-184.
- 1954. Die trilobiten des Ebbe-Sattels und zu vergleichende Arten (Ordovizium, Gothlandium, Devon). Abh. Senckenberg. Naturforsch. Gesell. 488: 1–76.

Rickard, L. V.

1989. Stratigraphy of the subsurface Lower and Middle Devonian of New York, Pennsylvania, Ohio, and Ontario. New York State Mus. Map Chart 39.

Rouault, M.

1851. Mémoire sur le terrain paléozoique des environs de Rennes, fossiles du terrain silurien et dévonien. Bull. Soc. Géol. France (2) 8: 1-399.

Schmidt, W. E.

1907. Cryphaeus in der Siegener schichten.Z. Deutsch. Geol. Gessel. 59: 9-12.

Shergold, J. H.

1966. A revision of Acaste downingiae (Murchison) and related trilobites. Palaeontology 9: 183-207.

Shimer, H. W., and R. R. Shrock

1944. Index fossils of North America. Cambridge, Massachusetts: MIT Press.

Smeenk, Z.

1983. Devonian trilobites of the southern Cantabrian Mountains (northern Spain) with a systematic description of the Asteropyginae. Leidse Geol. Mededel. 52: 383-511.

Speyer, S. E.

1987. Comparative taphonomy and palaeoecology of trilobite lagerstatten. Alcheringa 11: 205–232.

1988. Biostratinomy and functional morphology of enrollment in two Middle Devonian trilobites. Lethaia 21: 121-138.

Steininger, J.

1831. Observations sur les fossiles du Calcaire intermediare de l'Eifel. Mems. Soc. Géol. France 1: 331-371.

Struve, W.

1958a. Acastavinae n. subfam. Senckenb. Leth. 39: 221-226.

1958b. Beitrage zur Kenntnis der Phacopacea (Trilobita), 3: Treveropyge n. gen. und Comura (Delocare) n. subgen. (Asteropyginae). Senckenb. Leth. 39: 227– 234.

1959. Asteropyginae. In R.C. Moore (ed.), Treatise on invertebrate paleontology, Part O, Arthropoda 1, 477–483. Lawrence, Kansas: Univ. Kansas Press. Stumm, E. C.

1953. Trilobites of the Devonian Traverse Group of Michigan. Contrib. Mus. Paleontol. Univ. Mich. 10(6): 101–157.

1965. Two new species of trilobites from the Middle Devonian silica shale of northwestern Ohio. Contrib. Mus. Paleontol. Univ. Mich. 19(13): 163–166.

1967. Devonian trilobites from northwestern Ohio, northern Michigan, and western New York. Contrib. Mus. Paleontol. Univ. Mich. 21(6): 109-122.

Swofford, D. L.

1990. PAUP (phylogenetic analysis using parsimony) version 3.0q.

Termier, G., and H. Termier

1950. Paleontologie marocaine, t. II, Invertebres de l'Ere primaire: Annelides, Arthropodes, Echinodermes, Conularides et Graptolites, IV. Paris.

Timm, J.

1981. Zur trilobiten Stratigraphie des silur/devon Grenzbereiches im Ebbe-Antiklinorium (Rheinisches Schiefergebirge). Mitt. Geol.-Pälaontol. Inst. Univ. Hamburg 50: 91–108.

Tomczykowa, E.

1991. Upper Silurian and Lower Devonian trilobites of Poland. Prace Panstwow. Inst. Geol. 134: 1-62.

Tomczykowa, E., and E. Witwicka

1974. Stratigraphic correlation of Podolasian deposits on the basis of ostracodes and trilobites in the Peri-Baltic area of Poland (Upper Silurian). Biul. Inst. Geol. 276: 55–84.

Vrba, E. S., J. R. Vaisnys, J. E. Gatesy, R. De-Salle, and K.-Y. Wei

1994. Analysis of paedomorphosis using allometric characters: the example of Reduncini antelopes (Bovidae, Mammalia). Syst. Biol. 43: 92–116.

Walter, O. T.

1923. Trilobites of Iowa and some related Paleozoic forms. Iowa Geol. Surv. 31: 173–400.

Wiley, E. O.

1979. An annotated Linnaean hierarchy, with comments on natural taxa and competing systems. Syst. Zool. 28: 308–337.





