THE SYSTEMATICS, PHYLOGENY, AND ZOOGEOGRAPHY OF THE BLISSINAE OF THE WORLD (HEMIPTERA, LYGAEIDAE)

JAMES A. SLATER

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BULLETIN AMERICAN MUSEUM OF NATURAL HISTORY VOL. 165

ABSTRACT

The subfamily Blissinae is defined and the following aspects treated: a historical review, a summary of the biology and feeding habits, a checklist of world species, and an analysis of characters of taxonomic and phylogenetic importance. A proposed phylogeny is presented with cladograms. The distribution of the taxa is discussed and an interpretation is given. Keys are included for adults of all genera and species of the world, and for fifth instar nymphs of all species for which they are known. A diagnosis is given for each genus as well as its original reference, synonymy, type species, and general distribu-

tion. A figure is included for at least one species of each genus as well as 170 figures of anatomical details and 19 diagrams showing phylogenetic relationships. Slaterellinae Drake and Davis is placed as a junior synonym of Blissinae. Blissus navis Slater is transferred to Capodemus, Macropes tinctus Distant to Cavelerius, and Dimorphopterus aleocharoides Jakovlev to Ischnodemus. Neoblissus Bergroth is placed as a junior synonym of Blissus Burmeister. Pseudoblissus is erected as a new genus to contain Blissus trispinosus Slater.

INTRODUCTION

The Blissinae constitute a monophyletic group of lygaeid bugs. The subfamily is characterized not only by several morphological features such as the unique spiracle arrangement (two-six dorsal, seven ventral) but also by its unique trophic position within the Lygaeidae. This is the only subfamily of the Lygaeidae that feeds entirely on the sap of plant tissues rather than on the seeds, and the only one where most of the species live between leaf sheaths (laminaphiles of Slater, 1977).

I have devoted a considerable portion of my research efforts of the past 15 years to understand the systematic relationships and biological and ecological characteristics of members of the subfamily. Most of the descriptive work that can be accomplished from existing material has been completed. It thus seems important and desirable to attempt to bring together in one paper not only keys to all genera and species but also to attempt a preliminary interpretation of the phylogenetic and zoogeographic relationships within the subfamily.

In the present paper I include a cladistic analysis of the blissine genera, a discussion of the zoogeographic patterns, a checklist of world species, keys to all genera and species and, in some cases, a phylogenetic analysis of the species within the genus.

The biological section is brief as I (Slater,

1976) have previously discussed the host plant relationships in considerable detail. On the other hand, a key to species is included even where a recent key has been published. A dorsal view illustration is given for one species of each genus and where possible an illustration of the sperm reservoir of at least one species of each genus. Other illustrations are included where necessary to clarify the text.

ACKNOWLEDGMENTS

I express my appreciation to the following for their help: the late Mrs. Darleen Wilcox (University of Connecticut) who collaborated with me on many aspects of blissine studies and who prepared preliminary drafts of several of the more complex keys presented in this paper; to Dr. Peter D. Ashlock (University of Kansas) for collaborative work on the phylogeny of the Blissinae; to Ms Jane O'Donnell (University of Connecticut) for extensive assistance with the manuscript, dissections, interpretation and illustration of genitalia; to Dr. Abdul Hamid (University of Sokoto, Nigeria) for aid in dissection and illustration; to Dr. Randall T. Schuh (American Museum of Natural History), Dr. Merrill Sweet (Texas A & M University), Mr. Jack Munting (Pretoria, South Africa), Mr. Samuel Slater, Dr. R. M. Baranowski (University of Florida) and Dr. B. J. Harrington (University of Wisconsin) for aid in fieldwork in Africa and the West Indies (Dr. Sweet was also helpful in providing me with an interpretation of the coxal cavities and Dr. Schuh for critical review of the entire manuscript and for discussion of phylogeny and zoogeography); to Ms Mary Jane Spring for preparation of the charts and graphs; to Ms Mary Jane Spring, Dr. Molly Stock (University of Idaho), Mrs. Kathy Schmidt (American Museum of Natural History), Mr. Arthur Smith (formerly British Museum [Natural History]), Mr. Stephen Thurston (University of Connecticut), and Dr. Paul Godfrey (University of Mas-

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Special thanks are extended to the National Science Foundation for not only supporting the work but for assistance in publishing the manuscript, and to the University of Connecticut Research Foundation for financial assistance to Dr. Ashlock to enable us to work together in 1974 on portions of the phylogenetic analysis.

HISTORICAL REVIEW

The subfamily Blissinae was established by Stål (1862) as "Blissida" in a key and has remained a rather noncontroversial concept ever since. There were, of course, species and genera described prior to that time.

The first blissine species to be described was, surprisingly, the Neotropical species Cimex fulvipes by DeGeer in 1773. In 1794 Fabricius described Acanthia gibbus from the Orient and in 1803 he described Lygaeus oblongus from the Neotropics. The first European species to be described was Lygaeus sabuleti by Fallen in 1826. In 1831 Thomas Say described the two abundant and widespread Nearctic species Lygaeus leucopterus Lygaeus falicus. These early species were, of course, all described in expansive early omnibus genera such as Cimex, Lygaeus, and Acanthia. In 1835 Burmeister described the nominate genus Blissus with an African species hirtulus as monotype. Prior to 1861 only six generic names were used in the subfamily and two of these have proved to be junior synonyms. It thus further attests to the genius of Carl Stål who conceived and erected a subfamily based upon four genera and only 30 species that has been capable of adequately encompassing the present complement of over 50 genera and 400 species.

Subsequent to these early descriptions the history of systematic work on the Blissinae for many years followed the path of numerous

other Hemiptera groups. Descriptions of new species and genera were added by such serious students of Hemiptera as Spinola, Signoret, Stål, Reuter, Bergroth, Horvath, Berg, Walker, Distant, Van Duzee, Barber, and many others. By 1960 when Slater (1964) catalogued the literature, 50 authors were listed as describers of new species of the Blissinae (excluding species synonymized by 1960). The bulk of the descriptive work was accomplished by northern European, British, and North American workers with W. L. Distant at the British Museum having described by far the most species (45). Most of the blissine descriptions in the long period from 1850 to 1960 were either done as parts of extensive faunal works, or as parts of reports of expeditions and collecting trips. No careful revisional study had been accomplished on any large genus and no phylogenetic or zoogeographic analysis had been undertaken.

Just prior to 1960 the present author began a series of revisional and faunal studies of the world blissine fauna. The work of my colleagues (Ashlock, Harrington, Miyamoto, Ahmad, and especially the late Darleen Wilcox) and myself has resulted in a series of revisional studies in the course of which 46 percent of the present genera and 60 percent of the species have been described.

It is appropriate to say a word here concerning the importance of having some historical perspective of the maturity of knowledge about a group before one undertakes cladistic and/or zoogeographic analysis. The descriptive phase of taxonomy in ornithology and mammalogy especially is so nearly "complete" that students can often undertake phylogenetic analyses without engaging in detailed descriptive work. This is a very dangerous procedure in most entomological groups and certainly in Lygaeidae. If such an analysis had been attempted in the Blissinae prior to 1960 it would have included little more than one-half of the fauna then existing in collections and interpretations of generic relationships would certainly have been faulty. This is not to say that the present analysis is more than preliminary, but it should be many times closer to the truth than could previously have been possible. There is sometimes an unfortunate tendency to deprecate the value of descriptive systematics, but it has become clear to me during the course of this work that not only is such work necessary but without it most classifications will be as soundly based and as permanent as sand castles on a beach.

The history of biological and ecological knowledge of the Blissinae is more difficult to understand. It is safe to say that for most of the species there is no knowledge. We have at best only a general idea of the distribution of most species, we do not know anything of their host plants, life histories, ecological requirements, parasites, predators, mimetic associations, etc. Nevertheless, a considerable amount of information has accumulated for several species, some of it of a quite sophisticated nature.

The North American Chinch Bug *Blissus* leucopterus (Say) has been a serious pest of corn (Zea mays) in the central United States for

more than a century and as a result its biology and ecology have been studied in detail. As early as 1856 Fitch published a detailed study of the species and subsequent detailed analyses have been written by Walsh and Riley (1869), Riley (1875), Thomas (1877), Howard (1887), Webster (1899, 1907), Forbes (1905), Headlee and McCulloch (1913), and many others. The eastern subspecies *Blissus leucopterus hirtus* Montandon, which is a lawn pest rather than a crop pest, also has developed an extensive literature as has the southern Chinch Bug *Blissus insularis* Barber.

The European *Ischnodemus sabuleti* (Fallen) has been studied carefully especially by Tischler (1960), Putshkov (1969), and the earlier literature is nicely summarized by Southwood and Leston (1959).

For the other species, what biological information exists consists chiefly of host plant associations, some habitat information, descriptions and duration of immature stages and occasionally comments on abundance and dispersal. Slater (1976) has summarized and analyzed the host plant relationships in detail and has discussed their significance both for the plant taxa involved and for adaptations of the insects to their hosts. This discussion will not be repeated here other than to point out the following: All true known breeding hosts of the Blissinae are monocots.

The majority of host plants are members of the Gramineae but host plants are also known in the families Cyperaceae, Restionaceae, Zingiberaceae, Juncaceae, Sparganiaceae, Typhaceae, and Haemodoraceae. Slater's (1976) paper gave host plant information for 21 genera and 111 species of Blissinae.

ANALYSIS OF CHARACTERS

MALE GENITALIA

SPERM RESERVOIR: The plesiomorphic condition in the Blissinae appears to be that of a large ovoid bulb with narrow straplike wings extending basad (fig. 1P, W). This condition is widespread in other subfamilies of Lygaeidae and is found in blissines that have, what I believe, are numerous other plesiomorphic

character states, and this type of sperm reservoir is found in species and genera of Blissinae that are otherwise quite dissimilar. I interpret this to be the retention of a plesiomorphic condition rather than the development of apomorphic ones. For sperm reservoirs of this type to be considered apomorphic would require the condition to have evolved several times within

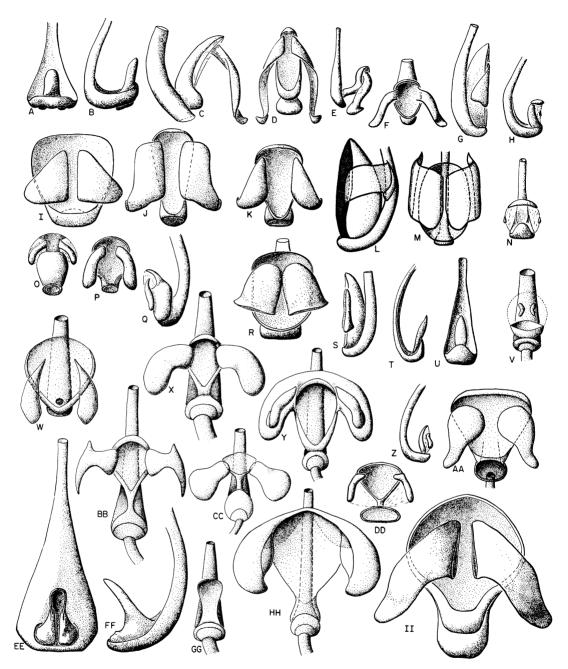


FIG. 1. Sperm reservoir, dorsal and lateral views. A,B. Slaterellus hackeri. A. Dorsal view. B. Lateral view. C,D. Patritius laevus. C. Lateral view. D. Dorsal View. E,F. Extarademus macer. E. Lateral view. F. Dorsal view. G. Ischnodemus sp., lateral view. H. Ischnodemus genei, lateral view. I. Ischnodemus sp. (Manchuria), dorsal view. J. Ischnodemus caspius, dorsal view. K. Ischnodemus sabuleti, dorsal view. L. Ischnodemus basalis, lateral view. M. Ischnodemus ochripes, dorsal view. N. Ischnodemus genei, dorsal view. O. Ischnodemus sordidus, dorsal view. P. Talpoblissus latus, dorsal view. Q. Ischnodemus sordidus, lateral view. R,S. Ischnodemus nr. suturalis (Syria). R. Dorsal view. S. Lateral view. T,U. Heinsius explicatus. T.

the Blissinae. This must not be taken to mean that this type of sperm reservoir is necessarily the plesiomorphic condition for the Lygaeidae. No serious attempt has yet been made to study this structure throughout the family. If the interpretation of the plesiomorphic condition of the sperm reservoir is correct then two distinct trends are evident within the subfamily.

One trend is toward enlargement of the wings into relatively huge platelike sheets (fig. II, J, K, M, R) that often are "curled over" at the outer margins (fig. IL, M). This apomorphic condition is found in all "advanced" species of *Ischnodemus*. In the most apomorphic taxa these wings shift toward the outer end of the bulb, become strongly curled under laterally and are somewhat reduced in total area (fig. IL). Since these wings function to depress the membrane of the bulb and force fluid from the bulb out the ejaculatory tube, the functional value of large robust wings is evident (see Slater and Harrington, 1970).

A second trend is that of reduction. In contrast to the development of platelike wings, reduction has not been a single evolutionary event but has developed a number of times and in a number of different ways. Frequently the basal area of the bulb becomes solid and forms a basal "stalk" (fig. 1E, O, Q) which reduces the functional bulb area and often the size of the wings. A sequence of reduction events can be distinguished. First the bulb and wings become smaller and smaller, then the wings eventually disappear entirely, and finally only a minute hook remains at the base of the ejaculatory tube (fig. 1A, B, T, U) to indicate where the sperm reservoir was attached. It is remarkable to dissect some of the largest and most robust blissines, such as Bochrus and find only such a minute remnant present. At some point in this reduction sequence the sperm reservoir must become nonfunctional. It is very difficult to distinguish between what are independent reductions and what are reduction sequences within single phyletic lines. Reference to the accompanying "cladograms" and illustrations indicate an interpretation of what has occurred in the Blissinae.

One conclusion that study of the blissine sperm reservoir has brought forcibly home to me is the impracticality of attempting to establish higher group relationships by sampling only a few taxa. Such a procedure would certainly have led to erroneous or superficial conclusions with this character.

CLASPERS (Parameres): The blissine clasper is for the most part a simple conventional structure consisting of a large, curving, rather sickle-shaped "blade," a short thick "shaft," a rounded, thumblike "outer projection" that marks the juncture of blade and shaft, and a rounded or sharp "subbasal inner angle" often near the base on the inner side (fig. 2A). This type of clasper is widespread in the Blissinae and occurs in many other subfamilies of the Lygaeidae as well.

In *Macropes* and related genera the "shaft" is greatly elongated, the blade reduced and the outer projection often absent so that a relatively slender, undifferentiated clasper results (fig. 2B). In *Spalacocoris* and related taxa the simplification of the clasper has proceeded to the point where only a short blocklike structure remains (see Slater and Ahmad 1968).

FEMALE GENITALIA

SPERMATHECA: A distinct spermatheca is present in all species of the Blissinae that have been examined. It is generally simple in structure with a bulb, pump, and tube (fig. 2C-R) (Dupuis, 1955). The conformation of the bulb and length of the pump show considerable variability, the structure, however, despite its value in other groups, appears to be of limited phylogenetic value in the Blissinae. Not only is the spermatheca frequently quite similar in otherwise dissimilar taxa but sometimes closely re-

Lateral view. U. Dorsal view. V. Riggiella vianai, dorsal view. W. Ischnocoridea elegans, dorsal view. X. Scintillademus gemmatus, dorsal view. Y. Xenoblissus lutzi, dorsal view. Z. Blissiella sp. (Madagascar), lateral view. AA. Heteroblissus anomilis, dorsal view. BB. Iphicrates nigritus, dorsal view. CC. Dentisblissus venosus, dorsal view. DD. Blissiella sp. (Madagascar), dorsal view. EE,FF. Australodemus elongatus. EE. Dorsal view. FF. Lateral view. GG. Micaredemus pusillus, dorsal view. HH. Ramadademus sakalava, dorsal view. II. Cavelerius illustris, dorsal view.

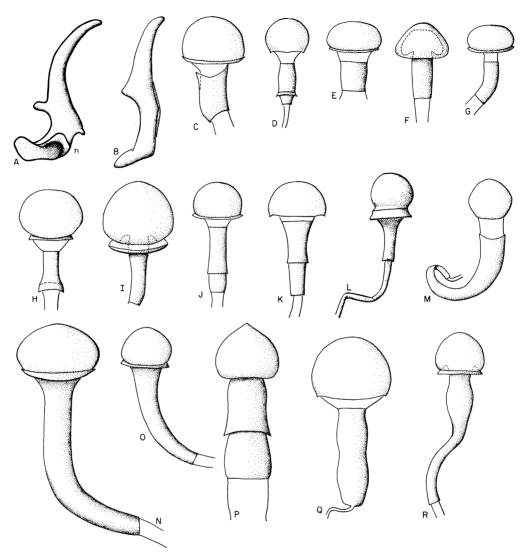


FIG. 2. A,B. Clasper of male. A. Ischnodemus ochripes. B. Macropes varipennis. C-R. Spermatheca of female. C. Aradademus mirificus. D. Macropes femoralis. E. Blissiella castanea. F. Heinsius explicatus. G. Barademus attenuatus. H. Caveloblissus americanus. I. Heteroblissus anomilis. J. Macropes raja. K. Iphicrates nigritus. L. Macchiademus capensis. M. Micaredemus obscurellus. N. Dimorphopterus pallipes. O. Dimorphopterus annulatus. P. Dentisblissus divisus. Q. Ischnocoridea elegans. R. Atrademus capeneri.

lated congeneric species will have striking spermathecal differences. For example, in the Neotropical genus *Reticulatodemus* (see Slater and Wilcox, 1966) the species *umbrosus*, *orbiculatus*, and *nitidus* have differences in the spermatheca that are greater than that found between species of many different blissine genera.

In some Oriental and Australian genera the pump has a transverse expansion midway along its length so that it appears to consist of two discrete parts (fig. 2D, J, K, P). This condition occurs in at least some species of *Iphicrates*, *Macropes*, and *Dentisblissus* and is clearly an apomorphic condition.

It is possible that other features of the sper-

matheca such as the elongation of the pump in species of *Dimorphopterus* will prove to be of phylogenetic value when more species are studied.

FORE COXAL CAVITIES: Slater and Ashlock (1976) have discussed the significance of this character, pointing out that closed coxal cavities are, among the Lygaeidae, found only in certain taxa of the Blissinae. They noted that open coxal cavities (fig. 3A, B, C) are not the result of wing reduction and resultant decrease in the size of the posterior pronotal lobe, nor is there direct correlation between closed coxal cavities (fig. 3D, E, F) and body elongation. It is true that, in general, elongate blissines have closed coxal cavities and short-bodied ones have the coxal cavities open. However, the general correlation of open coxal cavities and short bodies is due to the fact that most plesiomorphic Blissinae are short bodied.

I am indebted to Dr. Merrill Sweet (personal commun.) for suggesting a functional explanation for modifications of the fore coxal area in the Blissinae. Sweet suggests that the median prosternal spine that extends posteriorly between the fore coxae braces the prothorax against the mesosternum and strengthens the segmental connection between pro- and mesothorax. Thus the segmental connection is able to withstand the stress of ventral (downward) flexion without loss of dorsal mobility.

This explanation is reasonable for, in addition to open coxal cavities being plesiomorphic

and thus not useful in forming a group, it is an oversimplification to separate the Blissinae into two groups on the basis of "open" or "closed" fore coxal cavities. Once the coxal cavities have "closed" there has been a series of specializations that serve to increase the efficiency of the area. The prosternal spine becomes increasingly spatulate and the adjacent pleural area extends inward until the pleural "bridges" and the spatulate prosternal spine make contact and a strongly "closed" coxal cavity is formed. Subsequently increasing thickness of the pleural bridge would continue to increase the strength of the connection. Sweet notes that an elongate insect moving about the linear stems and leaves would be dangerously exposed to downward flexion and that the method of feeding requires dorsal flexion to raise the stylets.

Thus it is easy to understand why as the Blissinae become more and more closely associated with the linear stems and leaves of grasses and sedges there would be selection for body elongation and closing of the fore coxal cavities.

SURFACE PRUINOSITY: The pruinose "bloom-like" area that are conspicuous features of the pronotum, head, and scutellum of many Blissinae have been shown by Slater and Harrington (1970) to be composed of minute hairs. The importance of pruinosity for the classification of *Ischnodemus* has been discussed by Slater and Wilcox (1969) and its significance in cladistics by Slater and Ashlock (1976). The

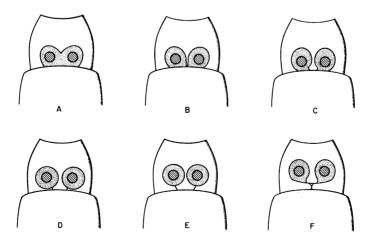


Fig. 3. A-E. Schematic representations of fore coxal cavity modifications.

latter paper points out that pruinosity loss, particularly that of the prothorax, does not occur randomly. The most common sequence appears to be as follows:

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Prothorax: Shining areas posteriorly on the pronotum consisting of three individual units, a transversely elongate central shining stripe separated laterally by pruinosity from an ovoid shining area on each humeral elevation (fig. 4B).

Coalescence of the three shining areas into a single shining strip across the posterior portion of the pronotum (fig. 4C-E). There may or may not be a very narrow pruinose area retained along the extreme posterior margin of the pronotum.

Development of a series of tiny shining dots that collectively define a triangular area on each callus of the anterior pronotal lobe (fig. 4B).

Coalescence of the calli "dots" to form a pair of shining triangular patches on the anterior pronotal lobe (fig. 4D, E).

Expansion of the intrahumeral shining strip to include all or almost all of the posterior pronotal lobe (fig. 4F).

Expansion of the dorsal shining areas until pruinosity is reduced to a narrow band behind the pronotal "collar" and as a pair of mesally narrowing wedges (fig. 4K, L) across the transverse impression that may or may not meet at the midline.

A completely shining dorsal pronotal surface (fig. 40) but with propleuron completely pruinose even above a well-differentiated longitudinal impressed stripe (fig. 5Q).

Restriction of pleural pruinosity to below the lateral impressed stripe (fig. 5N, O).

Reduction of pleural pruinosity to the ventral half to cover an area roughly bounded dorsally by the upper edge of the acetabulum.

Restriction of pruinosity to in front of and behind acetabula, the latter shining (fig. 5T).

Restriction of pruinosity to a narrow ventral area before and between coxae.

Complete loss of all pruinosity.

The great majority of the Blissinae may be placed in one of the above categories insofar as their prothoracic pruinosity pattern is concerned. There are a number of submodifications and variations as might be expected. However, divergence from the above is by no means

random. For example, one never finds a blissine with dorsal pruinosity and shining nonpruinose pleural and sternal areas; one never finds blissines with shining areas across the depressed transverse impression and behind the "collar" and pruinose areas on the calli and between the humeri. In other words there is an observable sequence of pruinosity loss and in general this proceeds as outlined above. Examples of major deviations include the following:

An entirely shining anterior one-half to the pronotum and a completely pruinose posterior one-half. (Praeblissus, Cavelerius, loblissus) (figs. 27, 28, 29, 67).

Development of the series of calli spots to form a triangular outline, or even large shining calli patches without the development of shining intrahumeral bands (fig. 4A) (this may well be a precursor condition to the preceding).

Loss of pruinosity on the posterior one-half of the propleuron but not the anterior one-half (fig. 5N).

Development of a broad shining stripe across the humeri and a similar one across the area of the calli (figs. 31, 61).

The best generalization that can be made is that pruinosity is first lost dorsally and on those dorsal surfaces that are the most elevated, and lost last in those areas most depressed; that pleural and sternal pruinosity loss never occurs until dorsal pruinosity loss is essentially complete and that it progresses from the dorsal margin of the propleuron ventrally.

Scutellum: Pruinosity disappears first distally along the raised median ridge or elevation (fig. 5K) and subsequently proximally on the meson (fig. 5J), then laterally on the anterior one-half (fig. 5I) and is lost last in the depressed areas laterally (fig. 5M).

Head: Pruinosity loss on the head does not appear to follow such a well-defined sequence as it does on the prothorax. This may well be because the eyes are frequently raised above the remainder of the head surface and the functional advantage of a smooth "sliding" surface is not present. In general, the tylus is not pruinose; frequently there is a somewhat "commashaped" shining strip adjacent to the inner margin of each ocellus that in many cases becomes bifid anteriorly. Further shining areas do follow some progression, the vertex mesally becoming

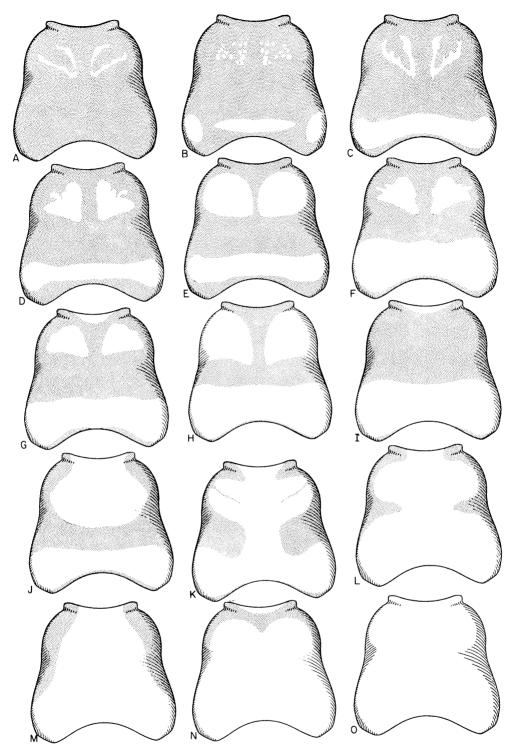


Fig. 4. Pronotum, dorsal view. Stippling indicates pruinosity. A. Ischnodemus oculatus. B. Ischnodemus fulvipes. C. Ischnodemus wittei. D. Ischnodemus ochripes. E. Ischnodemus bosqi. F. Ischnodemus linearis. G. Ischnodemus ocellaris. H. Ischnodemus schoutedeni. I. Ischnodemus lactipennis. J. Ischnodemus genei. K. Capodemus elegiae. L. Ischnodemus umbrosus. M. Capodemus herbosus. N. Ischnodemus torquatus. O. Ischnodemus brevicornis.

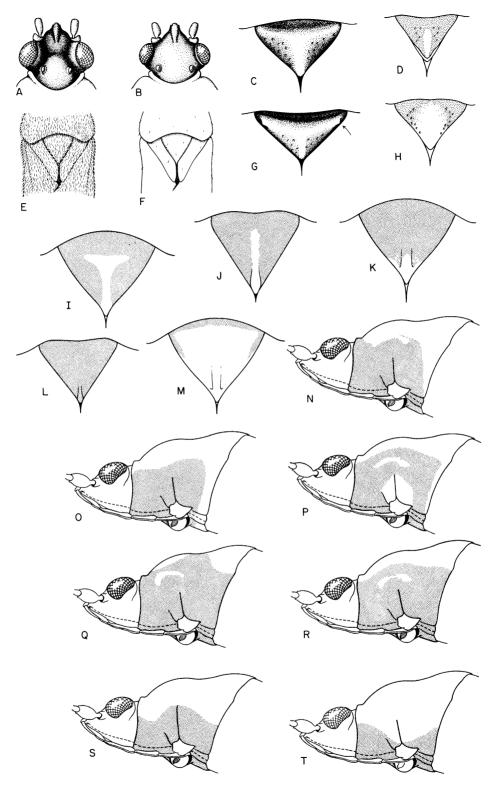


Fig. 5. A,B. Head, dorsal view. A. Micaredemus eleganoides. B. Micaredemus wilcoxae. C,D. Scutellum, dorsal view. C. Geoblissus mekongensis. D. Capodemus wilcoxae. E,F. Middle one-third of body, dorsal

shining before it does laterally and the head losing pruinosity dorsally before it does so laterally and ventrally.

Hemelytra: The most common pruinosity loss is along the raised radial vein. In some species the lateral third or half of the corium is shining and contrasts strongly with the pruinose inner portion (never the reverse). I do not know of any case where the basal half of the corium is of one composition and the distal half the reverse.

The membrane of the forewing is usually either completely shining or dull but in some large forms, particularly certain species of *Macropes*, there may be alternating transverse bands of shining and dull surfaces. This probably is not actually pruinosity or lack of it but gives a similar effect when viewed with the light microscope.

Although the functional significance of pruinosity loss has not actually been demonstrated, it is evident that the dorsal surfaces of the body that most frequently come into contact with leaf sheaths are the first areas to show loss of pruinosity. For a blissine that moves between two closely appressed surfaces, as most of them do, the dorsal surface is the one most directly in contact with the plant, for the ventral surface is slightly removed because of the presence of the legs.

These varying pruinosity patterns are very useful in blissine taxonomy as they are surprisingly constant for a given species. It is a character, however, that must be used with much caution in phylogenetic interpretation as it is evident that similar progression series have occurred in several quite distinct phyletic lines (see Slater and Ashlock, 1976).

SEXUAL DIMORPHISM: Marked sexual dimorphism is not widespread in the subfamily. Morphological specializations are generally present in males when the sexes are dimorphic. One of the striking features is the relatively high frequency of sexually dimorphic taxa in the Oriental Region as compared to other zoogeographic regions.

In the majority of blissine taxa the sexes are very similar in most external features. The only modifications usually evident is that females tend to average somewhat larger and to have a comparatively broader abdomen.

The following cases are examples of secondary sexual dimorphism known to occur in the subfamily.

ELONGATION OF THE MALE BUCCULAE: In the genus *Iphicrates* males of all species have bucculae that extend considerably anterior to the apex of the tylus and in many species they are broadened distally (fig. 6F). The females of many *Iphicrates* have the bucculae somewhat more produced anteriorly than in other Blissinae, but in almost all species those of the males are much larger. *Aradacrates*, a monotypic Madagascar genus, also has produced bucculae (fig. 18), but apparently these have evolved independently. (Females of this genus are unknown.)

ELONGATION OF THE JUGA: This condition is confined to certain species of *Iphicrates* where the male juga become very long and tapered in contrast to the conventionally subtriangular blissine condition found in females of the same species.

ELONGATION OF THE GENAE: Males of the genus *Dentisblissus* have greatly elongated and anteriorly produced genae that appear as heavy "tusklike" extensions and are usually bifid apically (fig. 31). In the females of *Dentisblissus* and males and females of species of *Scintillademus* and *Patritius* a very short protrusion is present in the genal area.

EYE SHAPE: Marked sexual dimorphism of the eyes occurs in many species of *Pirkimerus*. The eyes of females are very large and protrude markedly from the lateral margin of the head. The eyes of males, by contrast, are relatively narrow, protrude only slightly, and with the rest of the lateral margin of the head, form an almost even, gently convex arc.

ENLARGEMENT OF THE HIND FEMORA AND TIBIAE: In males of the genus *Bochrus* and some species of *Patritius* and *Extarademus* the

view. E. Blissiella castanea. F. Blissiella micans. G-M. Scutellum, dorsal view. G. Geoblissus siccus. H. Capodemus variabilis. I. Macropes pseudofemoralis. J. Ischnodemus mendax. K. Macropes nigrolineatus. L. Ischnodemus inornatus. M. Macropes australis. N-T. Thoracic pruinosity, ventrolateral view. N. Capodemus pentameri. O. Capodemus tenuatus. P. Capodemus wilcoxae. Q. Capodemus variabilis. R. Capodemus stuckenbergi. S. Ischnodemus brunnipennis. T. Ischnodemus slossoni.

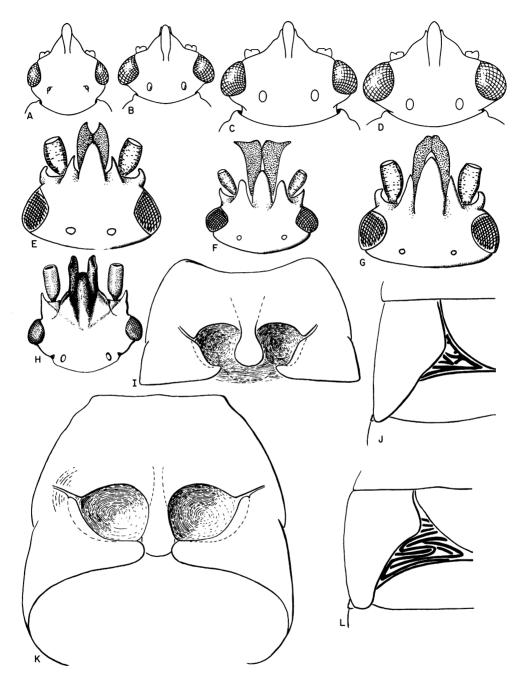


Fig. 6. A-G. Head, dorsal view. A. Macchiademus acuminatus. B. Macchiademus capensis. C. Ischnodemus diplachne. D. Ischnodemus oculatus. E. Iphicrates pseudolineatus. F. Iphicrates lativentris. G. Iphicrates montaguei. H. Iphicrates rex. I. Open fore coxal cavities, Capodemus sabulosus. J. Mesonotal striations, Capodemus rusticoides. K. Closed fore coxal cavities, Atrademus capeneri. L. Mesonotal striations, Capodemus rusticus.

hind femora are much thicker and more incrassate than are those of the females. In at least one species of *Bochrus* there is also a large spine developed on the male tibiae. Some *Pirkimerus* species also have more strongly enlarged and spinose tibiae in the males than in the females.

ENLARGEMENT OF THE FORE FEMORA: Many of the Blissinae have strongly incrassate and spinose fore femora. In *Macropes spinimanus* this is much more pronounced in the males than in the females, and in a number of other *Macropes* the male fore femora are somewhat more swollen.

ABDOMINAL SPINES: Males of *Extarademus* have a series of spines on the posterior abdominal sterna (fig. 8A) and sometimes elongation of the connexiva (the very elongate connexiva of *Toonglasa* (fig. 79) may be sexually dimorphic but only a single specimen is known).

FEMORAL SPINES: Many species of the Blissinae have all femora mutic. However, the occurrence of a single spine ventrally on the distal third of the fore femora occurs in many taxa, two spines in this area are not uncommon, and there is a whole group of genera that have multispinose fore femora. Spines on the middle and hind femora are much less common and are rarely present as other than a series of spines and then almost always only in species that also have multispinose fore femora. Variation in the number of fore femoral spines intraspecifically has been discussed by Slater and Wilcox (1973) for the South African fauna and is not repeated here other than to say that generally species with only a single spine present only rarely have more, but in species with more than one spine, variation in the number is greater. Species that are generally mutic appear never to develop spines, and the reverse is so rare as to almost be an aberration.

If used with care, the presence, absence, or number of femoral spines is very helpful in blissine taxonomy, however, its use in phylogeny is very difficult. This is so in part because, like pruinosity patterns, both different and similar femoral spine conditions are found in different phyletic lines even within the same genus. Of more importance is the difficulty of deter-

mining whether the plesiomorphic condition for the Blissinae is mutic or spinose. Study of related outgroups is not especially valuable, for while many of the Lygaeidae have multispinose fore femora, many others do not. Further, spined and mutic fore femora both occur within tribes and subfamilies almost throughout the family, and of course we do not yet have a cladistic analysis of the higher taxa of the Lygaeidae. I suggest that mutic fore femora may be ancestral, but if so, a single fore femoral spine developed early. I suggest this because of the possibility that the Orsillinae is the sister group of the Blissinae (Ashlock, 1967) (if not, the sister group of the Blissinae probably is the Ischnorhynchinae or Lygaeinae) and all have many taxa with mutic femora. Also blissines with multispinose fore femora usually have a number of apomorphic characters and in closely allied rather generalized blissine taxa such as Blissus-Dimorphopterus and Capodemus-Macchiademus, those species with other apomorphic characters are those most likely to have fore femoral spines present. There appears to be a functional advantage in having more femoral spines for insects that move through closely appressed leaf sheaths, in that spines can serve as a "brace" to hold the folded tibiae against the ventral surface of the femur. Thus, one can hypothesize selection for the acquisition and elaboration of the spines. However, species without spines do not seem to be concentrated in those taxa whose species live less closely associated with closely appressed surfaces. This is not to say that secondary loss of spines has not taken place. It appears to have occurred in Ischnodemus and Pirkimerus and may well have done so in other

Admittedly the evidence for the plesiomorphic condition is not strong and greatly limits the cladistic value of the character.

The mutic condition is certainly plesiomorphic for the middle and hind femora. Spines appear on these femora only in taxa with many other derived features.

APICAL CORIAL MARGIN: The value, variability, and cladistic limitations of this character are discussed in detail by Slater and

Ashlock (1976). Outgroup comparison seems to show conclusively that a straight corial margin is plesiomorphic. Concavity of the apical corial margin is a general statement that subsumes a variety of modifications of the margin, some of which at least have certainly occurred independently, and thus a concave apical corial margin in itself does not necessarily constitute a synapomorphy.

ANTENNAL SHAPE: Terete antennae appear to be the plesiomorphic condition. The evidence is similar to that for the apical corial margin, i.e., it is the predominate condition in other Lygaeidae and other families of Hemiptera. Strongly clavate antennae usually occur in taxa that have many other derived characters. (I understand the danger of circular reasoning here.) Like the concave apical corial margin, "clavate antennae" is by no means a "yes or no" condition. Many species that have what are called terete antennae show a tendency toward a slight enlargement of the second and third segments distally. In relatively plesiomorphic taxa clavate antennae seem generally to be associated with shortening of the body.

ABDOMINAL SCLERITES OF NYMPHS: Although the nymphs of the Blissinae appear to have a number of features important to an understanding of the phylogeny of the subfamily, it is difficult at present to use these characters in phylogenetic work, as nymphs of many species (including all species of a number of genera) remain unknown. Nymphs of most the Blissinae have a series of plates on the dorsal and ventral surfaces of the abdomen. Usually these are larger and more heavily sclerotized on the posterior segments. Slater and Wilcox (1973) present a coding system for recognition of the various sclerites (fig. 7). These plates (Slater, 1976) are most extensively developed in species that live between closely appressed leaf surfaces. They appear to be useful as "skid plates" to facilitate the movement of the body through tightly appressed surfaces. Many of the elongate Blissinae live in such habitats and are capable of moving backward almost as readily as they move forward. Thus, the value of strong plates at the posterior end of the abdomen is evident. The plates along the midline

are usually the largest on the posterior segments and appear to develop progressively from the posterior end of the body anteriorly, so that the plesiomorphic condition is one in which median dorsal plates (TM) are present on terga six and caudad, and the most apomorphic condition is where there is a TM plate on all terga from segment two posteriorly. The lateral plates form a similar although less complete series. The most apomorphic condition of these plates of any blissine whose nymphs are yet known is found in the Ethiopian and Madagascar genus Barademus (fig. 7P). The most plesiomorphic conditions are found in such genera as Dimorphopterus and Blissus.

ABDOMINAL TRICHOBOTHRIA: I have not personally studied this set of characters. Schaefer (1975) has investigated either nymphs and/or adults of 21 species of Blissinae, representing 13 genera (chiefly South African and North American). He believes that two "trends" may be present in heteropteran trichobothrial evolution. In the "main trend" the condition in which an increased number of trichobothria become present in earlier and earlier instars is apomorphic. In the "counter trend" the apomorphic condition is where fewer trichobothria are present in the later instars and the adult number is reduced.

Schaefer (1975) believed the "counter trend" best explains the situation in the blissines he studied, i.e., reduction of the full adult complement is apomorphic. This seems reasonable and is supported by the fact that although Schaefer did not indicate the adult trichobothrial complement of species of *Capodemus*, Slater and Sweet (1972) have previously shown that in members of this genus only two trichobothria each were present on sterna three and four. Clearly, there has been a reduction or "counter trend" in this genus at least.

The trichobothrial pattern in the Blissinae may be of some value within genera or closely allied groups of genera. However, frequently we are involved with a reduction phenomenon and in such cases synapomorphy can be established only with the greatest caution. Schaefer also notes that there is considerable individual variation (sometimes within the same individual

on a given segment) and that in this subfamily it is often very difficult, if not impossible, to differentiate trichobothria from other setae. It is my belief that a really extensive survey is necessary with long series studied to allow for understanding of individual variation (Davidova and Stys, 1976) before trichobothrial patterns can be used in the phylogeny of the Blissinae.

FORE TIBIAE: In most of the Blissinae the fore tibiae are cylindrical for much of their length, although they are frequently somewhat enlarged near the distal ends and bear a series of spines. The most strikingly apomorphic conditions occur in the Blissus-Dimorphopterus phyletic line. The most apomorphic states occur in such genera as Geoblissus and Talpoblissus (fig. 7G, H) where the fore tibiae are modified into enlarged and flattened "scoops" to enable the insect to burrow readily into sandy soil. In species of these two genera the "digging" modification appears to have evolved independently. In Geoblissus the entire tibia is flattened and bears a closely set series of teeth along both margins (fig. 37). These teeth are sometimes worn down to blunt stubs in some "old" specimens (Slater, Ashlock, and Wilcox, 1969). In Talpoblissus the terminal portion of the tibiae is strongly splayed out, somewhat "scalloped" with a heavy "tooth" present at the point of each "scallop" (fig. 7G, H). Many species of Dimorphopterus and Blissus have somewhat enlarged, flattened, and toothed fore tibiae.

In such genera as *Spalacocoris* (fig. 77) and *Chelochirus* (fig. 30) the fore tibiae are very short and thick, usually curved with a few large teeth or hooks on the distal ends. Although the habits of these bizarre blissines are unknown, it is possible that the fore tibial modifications are also fossorial adaptations.

Labial Length: This is a very useful character for specific discrimination, especially in large and complex genera such as *Ischnodemus*, but it is of limited value at higher group levels. The plesiomorphic condition appears to be one in which the labium extends posteriorly onto the extreme anterior portion of the mesosternum. A labium of this length is found in many species of a large number of genera of the

Blissinae. Apomorphic conditions are represented by both shortened and elongated labia and there is obviously a considerable amount of parallelism. The labial length probably functions in some manner in resource partitioning as often sister species within a genus have rather different labial lengths. How mouthparts function in insects moving about between appressed leaf surfaces is somewhat of a puzzle. In some other lygaeids the utility of a very elongate "beak" is quite apparent. For example, Oxycarenus maculatus and Dinomachus marshalli have greatly elongated labia which in the adults reach to or beyond the middle of the abdomen, and in early instar nymphs actually "trail" posteriorly beyond the end of the abdomen. Oxycarenus maculatus feeds on the seeds of African proteas, which lie deep within the flower heads. Dinomachus marshalli feeds on the seeds of Ficus sycomorus (and probably other Ficus spp.) and presumably feeds by inserting the stylets through the syconium to reach the seeds within. In the Blissinae, however, it is not evident how an elongate (or a shortened) "beak" is adaptive. This offers an interesting area for investigation, for how most of the Blissinae actually feed is completely unknown.

MESOSTERNAL FURROW: In most of the Blissinae there is a fine groove running longitudinally along the midline of the mesosternum. In some species, a definite deep troughed furrow is present. I previously considered this to be of phylogenetic importance. However, it now appears that there has been substantial parallel development and that it is at least in part correlated with elongation of the labium. The labium lies in this furrow and it can readily be seen that such a development should be advantageous to an insect moving through closely appressed leaf surfaces.

MEMBRANE TEXTURE: The plesiomorphic condition is one in which the membrane is noticeably thinner than the clavus and corium but semiopaque, thicker than the hind wing and of uniform texture. Two opposing apomorphic "trends" are evident; in one the membrane becomes increasingly leathery and opaque and can scarcely be differentiated in texture from

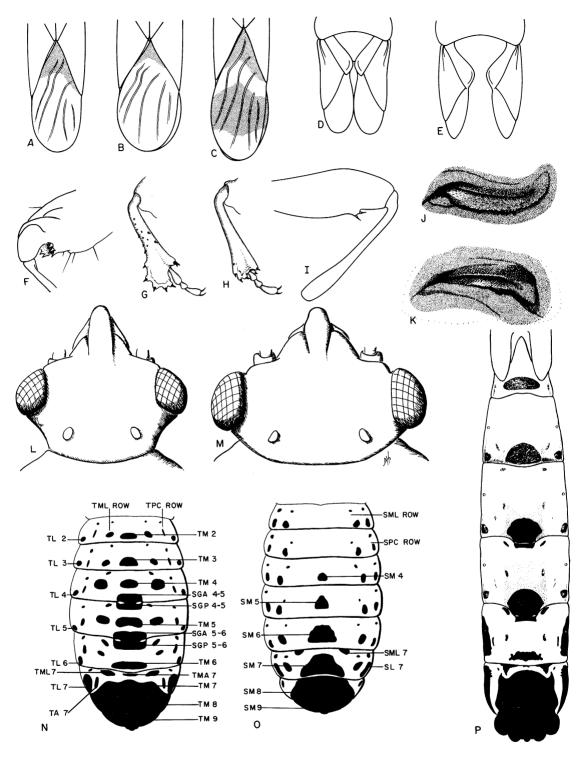


Fig. 7. A-C. Membrane of forewing. Stippling indicates pruinosity. A. Macrops lobatus. B. Macropes privus. C. Macropes albosignatus. D,E. Mesothoracic wings. D. Ischnodemus slossoni. E. Ischnodemus

the rest of the hemelytron. Such a condition is well exemplified by some of the large flattened species of Riggiella, Scansidemus, Chelochirus and members of the Macropes raja-complex. The converse apomorphic condition is one in which the membrane becomes very thin and hyaline and is either partially or completely transparent. In some species of Dimorphopterus the proximal part of the membrane may be opaque and the distal portion transparenthyaline. It is apparent that intermediate conditions exist and that the hyaline membrane has developed more than once. It is also apparent that there is, in some cases, a strong correlation of wing texture with other synapomorphic characters and that used with caution this feature is of some value in establishing cladistic relationships.

The membrane of some of the Blissinae such as *Reticulatodemus*, *Xenoblissus*, and *Gelastoblissus* is composed of a network of tiny cells (figs. 72, 80, 36). I have interpreted this for the most part as synapomorphy as is discussed under the individual taxa.

In some complexes of *Macropes* and related taxa the surface of the membrane has alternate bands of dull (pruinose) and bands of shining surface. This is an important feature but difficult to use in practice, as any accumulation of grease or dirt will obscure or obliterate the pattern, and even with material in good condition it requires some practice and care in orientation of the light source to make accurate comparative observations.

OVIPOSITOR LENGTH: This is a very useful character for specific discrimination and does have value at higher group levels. The major problem is in understanding what the plesiomorphic condition is because of the great variability of ovipositor length in other lygaeids and lack of knowledge of what is the sister group of the Blissinae. Certainly when the ovipositor has become platelike as it has in

Spalacocoris and Pirkimerus the condition is highly apomorphic, in fact approaching at least superficially the condition found in the Pentatomoidea. Extreme elongation of the ovipositor is also certainly relatively apomorphic. Beyond this it is really not possible to be certain. However, in studying a genus such as Ischnodemus one becomes impressed by the appearance of a progression from an ovipositor that does not completely divide the sixth sternum (fig. 8D) through types that do divide sternum six (fig. 8C, E, G), those that partly divide sternum five (fig. 8G), those that completely divide sternum five (fig. 8C), to those that partially divide sternum four. The shorter ovipositor also usually occurs in species that have a number of other plesiomorphic characters. Therefore, I believe the most reasonable hypothesis is to consider that the plesiomorphic condition of the ovipositor in the Blissinae is one in which the ovipositor is distinctly geniculate and laciniate but short, and when "at rest" does not completely divide sternum six. Thus, increasing modification of the anterior sterna by ovipositor elongation is relatively apomorphic.

In the Australian genera *Heinsius* and *Australodemus* the ovipositor is prolonged posteriorly beyond the remainder of the abdomen (fig. 8H, I), a strongly synapomorphic feature (see Slater and Sweet, 1963).

OCELLI: Most of the Blissinae have a pair of conventional small ocelli on the vertex. Occasionally in morphs with a high degree of wing reduction the ocelli will be reduced or lost. This is true in number of lygaeid groups and, although an apomorphic condition, is certainly a convergence or parallelism phenomenon.

In the Spalacocoris-Pirkimerus-Chelochirus clade the ocelli are much enlarged and I believe they represent an important synapomorphic character uniting these taxa.

ANTENNIFEROUS TUBERCLE: The plesiomor-

hesperius. F. Fore femur, Micaredemus quadratus. G,H. Fore tibia. G. Talpoblissus cydnoides. H. Talpoblissus latus. I. Fore femur, Ischnodemus sinuatus. J,K. Metathoracic scent gland auricle. J. Patritius fusconervosus. K. Patritius colombianus. L,M. Head, Dorsal view. L. Capodemus distinctus. M. Capodemus herbosus. N,O. Abdomen of nymph showing sclerite code. N. Dorsal view. O. Ventral view. P. Abdomen of nymph, dorsal view, Barademus attenuatus.

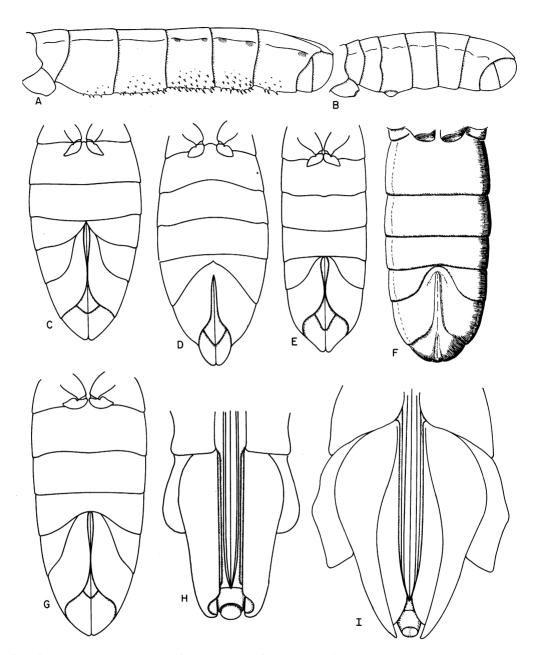


FIG. 8. A,B. Abdomen, lateral view. A. Extarademus macer. B. Macropes obnubilis. C-I. Abdomen of female, ventral view. C. Ischnodemus montanus. D. Ischnodemus fulvipes. E. Ischnodemus stali. F. Ischnodemus basalis. G. Ischnodemus sp. (Tanzania). H. Heinsius explicatus. I. Australodemus elongatus.

phic condition in the Blissinae (and indeed in the Lygaeidae) is that of a short truncate protrusion from the body wall, anterior to and slightly below the eye (fig. 6A, B). Species of Iphicrates, Reticulatodemus, and Extarademus have the outer margin of the tubercle produced forward and curved, so as to appear hooked or like a cow's horn on to the tubercle (fig. 6E-

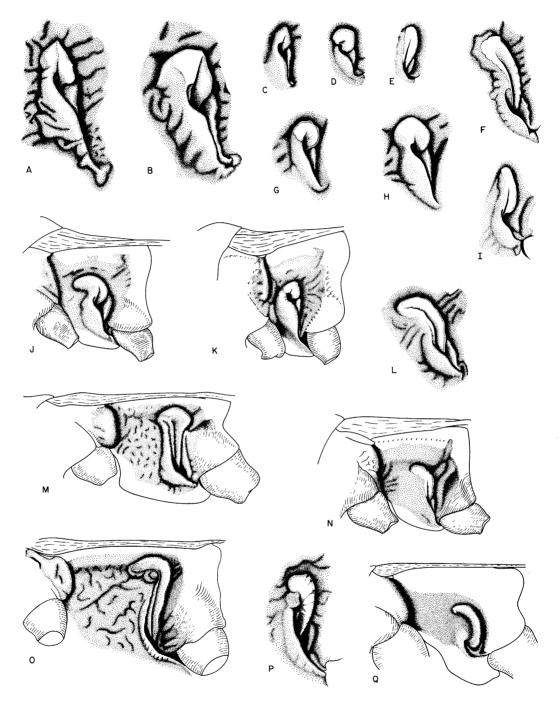


Fig. 9. Metathoracic scent gland auricle. A. Ischnodemus basilewskyi. B. Ischnodemus grossinigrus. C. Capodemus sabulosus. D. Macropes lobatus. E. Macropes uniformis. F. Macropes raja. G. Heteroblissus anomilis. H. Ischnocoridea elegans. I. Macropes spinimanus. J. Ischnodemus brincki. K. Blissus leucopterus. L. Extarademus humerus. M. Aradademus mirificus. N. Extarademus macer. O. Aradademus oculatus. P. Macropes albosignatus. Q. Barademus attenuatus.

G). Outgroup comparison suggests that this probably is a synapomorphic condition and is important because of the Neotropical-Oriental relationship thus established.

COLORATION: Color generally is affected so strongly by the immediate environment as to be of little value in cladistic analysis. It is relatively constant in some species, and leg and antennal color, as well as hemelytral patterns, have proved to be of considerable ancillary value in species segregation. The dark spot in the middle of the membrane that occurs in so many blissine lines probably has some disruptive pattern value and is sometimes useful in recognizing clades within large However, at the generic level, color does not seem to be useful in systematic work on this subfamily.

Savanna species, as might be anticipated, tend to be pale yellow, whereas there is a tendency for species living in more mesic habitats to be variegated, but we really know too little of the habits of most species to generalize further.

METATHORACIC SCENT GLAND AURICLE: This structure is of considerable phylogenetic importance in the subfamily. Not only is there a great variety of forms within the Blissinae but frequently a given genus will have a given type of auricle, and it is often possible to observe what appear to be synapomorphies in the shape of the structures that aid in the recognition of clades. In some genera the auricle is an important indicator of which of the species is the most plesiomorphic.

The plesiomorphic metathoracic scent gland auricle is a short ovoid button-like structure, probably somewhat narrowed at the distal end (fig. 9D, G, H). Evidence for this is the wide-spread occurrence of this type of auricle in other groups of the Lygaeidae and its presence in a number of blissine taxa. Also this type of auricle occurs in species of some genera, other members of which have different auricle shapes. Finally, this is the type of auricle that is most often present in taxa that have a number of other plesiomorphic characters. The above is not true of any other auricle type.

The simplest modification of this lobed or

button-like auricle is for the auricle to elongate (fig. 10A). This has occurred in several phyletic lines, and in advanced states the auricle sometimes expands irregularly at the distal end as in *Dentisblissus venosus* (fig. 11I), *Aradademus mirificus* (fig. 9M), and *Iphicrates rex* (fig. 11F). Another apomorphic condition is for the auricle to elongate and curve forward in a somewhat scimitar-shaped arc (fig. 9Q). In *Barademus*, *Micaredemus*, and *Extarademus* this appears to show cladistic relationship, but a somewhat similar condition apparently has been attained independently in *Macropes*.

The scent gland auricle is then one of the really important external features available for use in the classification and phylogenetic reconstruction of the taxa of the Blissinae.

WING POLYMORPHISM: Reduction of wings is widespread in the Blissinae. I have previously discussed the general phenomenon and proposed a classification of wing types (Slater, 1975, 1977). In the latter paper the Blissinae were placed in a special category which I called the "laminaphiles" (living between leaf sheaths) to distinguish them from the ground-living species (geophiles) and those that live on trees, shrubs, and forbs above the ground (arboreals). This special category is necessary as the development of flightlessness in the Blissinae much more closely parallels the condition found in the geophiles than it does that of the arboreals.

Whether deterioration of wing muscles occurs after a dispersal or mating flight period has not been investigated in this subfamily. When wing reduction occurs it takes the form of progressive shortening of the wing and never results in the formation of a coleopteroid or staphylinoid "shell" of the type that is so common in the Rhyparochrominae (see Slater, 1975 for definition of terms). Submacroptery (fig. 18), where the membrane is slightly shortened, and brachyptery, where the membrane, corium. and clavus are all shortened, are both common. Microptery (fig. 68) occurs but appears to be limited to taxa living under relatively stable ecological conditions. In some cases such as Praetorblissus and Capodemus the microptery is extreme, the forewing being reduced to a

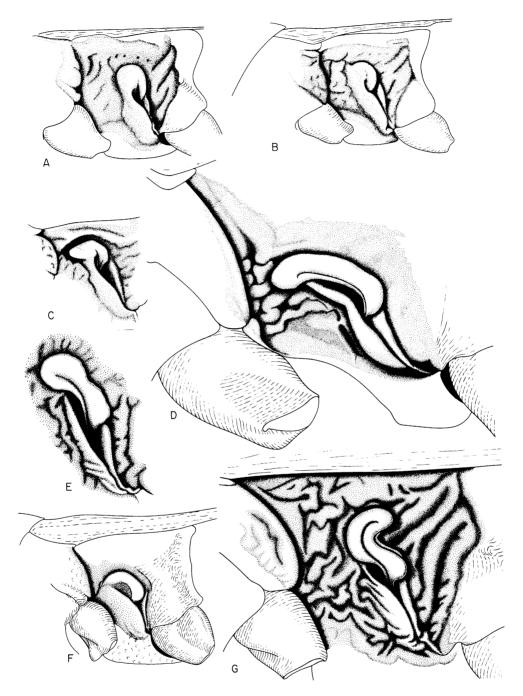


Fig. 10. Metathoracic scent gland auricle. A. Ischnodemus badius. B. Ischnodemus falicus. C. Ischnodemus asciaformis. D. Spalacocoris nigritus. E. Chelochirus confertus. F. Micaredemus pilosulus. G. Chelochirus talpus.

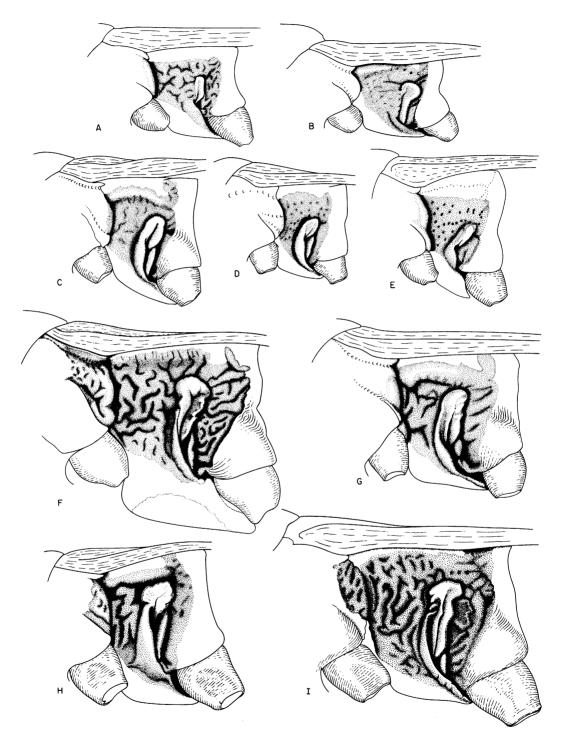


Fig. 11. Metathoracic scent gland auricle. A. Reticulatodemus nitidus. B. Iphicrates malayensis. C. Iphicrates angulatus. D. Iphicrates lativentris. E. Xenoblissus lutzi. F. Iphicrates rex. G. Iphicrates nigritus. H. Scintillademus gemmatus. I. Dentisblissus venosus.

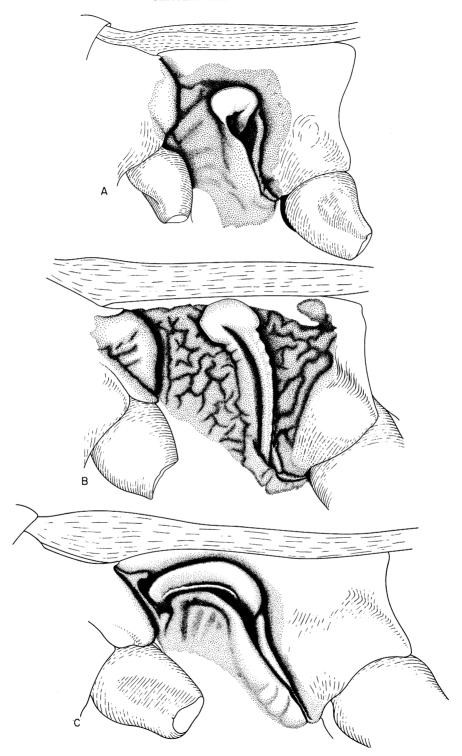


Fig. 12. Metathoracic scent gland auricle. A. Procellademus venenatus. B. Riggiella vianai. C. Ramadademus sakalava.

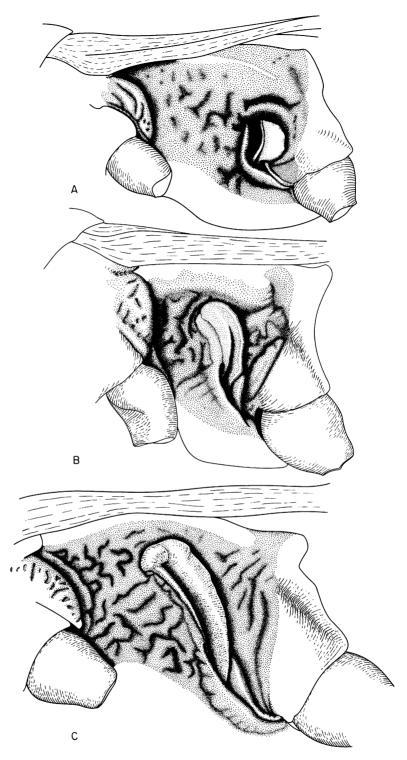


Fig. 13. Metathoracic scent gland auricle. A. Pirkimerus japonicus. B. Macropes major. C. Bochrus poecilopterus.

minute scalelike pad that does not extend posteriorly to cover even the metanotum.

Wing polymorphism is very important ecologically, but of almost no phylogenetic value, as it occurs sporadically in various species of diverse genera and is certainly more an expression of habitat stability than of cladistic relationship.

TARSAL SEGMENTS: Most blissine bugs have conventional tarsi composed of three segments, the first segment being the longest. Only in some of the most apomorphic taxa is there any significant modification. In some species of *Pirkimerus*, for example, the first segment is greatly elongated; in such genera as *Macropes*, *Chelochirus*, and their relatives the second segment is much reduced and the first segment swollen. Only two tarsal segments are rarely present. Since all nymphs have only two tarsal segments, adults with two segments may be neotenic.

PROPLEURON: In *Macropes* species and in some of the other large Oriental blissines the anterior portion of the propleuron is deeply excavated. Such species have the fore femora strongly incrassate and the propleural modifica-

tion appears to be an adaptation to allow the femur to "fit" into the body form when the insect is moving through narrow spaces. Nevertheless, not all blissines with enlarged fore femora show this modification, and I believe it to be of value cladistically. It is a difficult character to use in practice as actually the condition is an not either-or one but a series of increasingly deep impressions in the side of the prothorax.

Host Plants: Host plants and morphological adaptations of blissine bugs to their hosts have been extensively discussed previously (Slater, 1976) and are not repeated here. I note only the restriction of these insects to monocots and the concentration of species on the Gramineae. There is marked host specificity but hosts are unknown for so many taxa that this information can be used phylogenetically only in a very limited fashion. This does not mean, however, that eventually the insect-host relationship will not be extremely valuable (it almost certainly will be), but only that information is still insufficient for detailed comparative studies.

PHYLOGENETIC RELATIONSHIPS 1

The cladogram (fig. 15) is constructed on the basis of shared apomorphies rather than plesiomorphies. Trichotomies and polytomies occur in the generic cladogram and in some of the specific cladograms. Paraphyletic groups are recognized (*sensu* Hennig, 1966 and Ashlock, 1971). The question of the desirability of recognizing paraphyletic groups is discussed by Mayr (1974), Ashlock (1974), Nelson (1978), Michener (1978) and Brothers (1978), among others.

In attempting to reconstruct the phylogeny, reliance has been placed upon uniquely derived characters such as the extensive posterior sclerotization of the abdomen of the nymphs, the

'This section was done in collaboration with Dr. P. D. Ashlock (U. of Kansas) and is considered to be co-authored, although J. A. Slater is responsible for the final wording.

presence of a genital tubercle, and the development of broad platelike wings on the sperm reservoir. Such characters as clavate or terete antennae, degrees of thoracic, head and scutellar pruinosity, presence or absence of fore femoral spines, etc., while useful, are of secondary value since they appear to have developed a number of times in the evolution of the group. The broad flattened body, multispinose fore femora, deeply concave apical corial margin, and structural characteristics of this kind are somewhat intermediate between the unique characters and those that reappear upon a number of occasions. Thus, there is a galaxy of characters that must be used with considerable care if evidence for a reasonable approximation of the phylogenetic history of the group is to be marshalled in support of the hypothesis presented here.

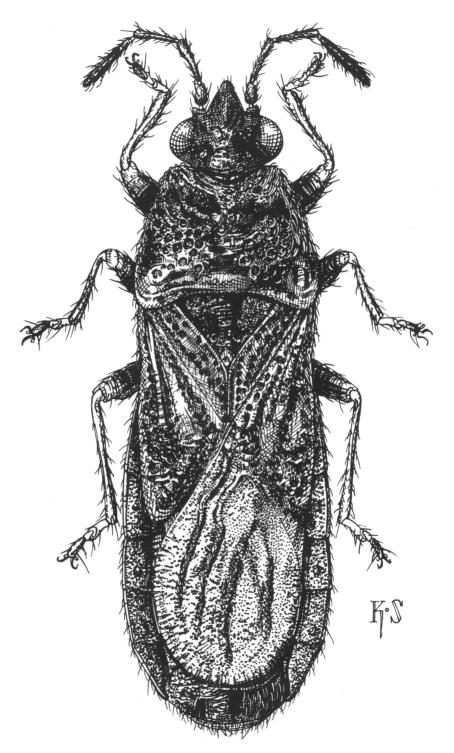


Fig. 14. Capodemus navis (Slater). Example of a generalized blissine adult.

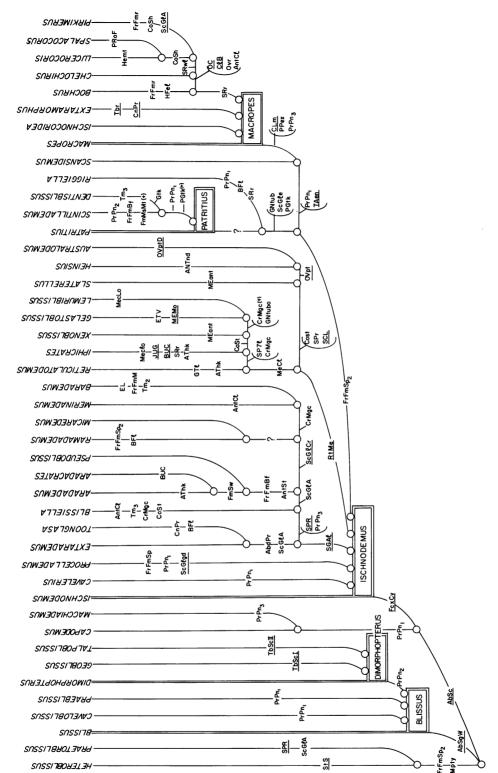


Fig. 15. Generic cladogram of the Blissinae. Genera in boxes are considered to represent paraphyletic genera and such generic names are carried to the top of the cladogram by a double line. Synapomorphic characters that are underlined are considered to be particularly "strong" characters in the construction of the cladogram. The distribution of the genera may be readily obtained from the accompanyiing check list of species.

In the cladogram (fig. 15) there is no appreciable deviation from the partial cladogram presented by Slater and Ashlock (1976) and that portrayed on the left side of the present cladogram.

In the cladistic analysis of species of Ischnodemus primary reliance has been placed on the condition of the phallus and especially the nature of the sperm reservoir. This is a complex structure and thus the various apomorphic conditions present are less likely to develop twice, or more than twice, than are most external features such as pruinosity patterns, fore femoral spines, "stalked" eyes, hyaline membranes, color patterns, grooved sterna, labial and ovipositor lengths, and so forth. It seems probable that certain characters found in the nymphs as well as the host plant relationships will ultimately prove to be of great phylogenetic importance, and they have been introduced into the Ischnodemus analysis as well as the generic analysis in several places. Unfortunately, this type of information is lacking for many species and genera.

At the base of the generic cladogram there is a trichotomy. As mentioned by Slater and Ashlock (1976) this is probably due to the unavailability of nymphs of Heteroblissus and Praetorblissus. It seems likely that *Heteroblissus*, although having several highly apomorphic characters, will prove to belong in a clade with Blissus and Dimorphopterus, whereas Praetorblissus may be one of the most plesiomorphic elements of a "line" now characterized by AbSc leading to the rest of the Blissinae. In any event, the sclerotization of the posterior segments of the abdomen of the nymphs (AbSc) is a very strong character which unites all the other genera of the Blissinae except those associated with the Blissus and Dimorphopterus clades which have been discussed by Slater and Ashlock (1976). This synapomorphy applies at a high level of generality within the group as it indicates the relationship of two genera (Capodemus and Macchiademus) that are essentially endemic in the southwest Cape Region of Africa with the *Ischnodemus* complex from ancestral components of which most of the genera of Blissinae appear to have been derived. Capodemus and Macchiademus, while agreeing with Ischnodemus and its descendant groups in having these uniquely derived sclerotized plates at the posterior end of the nymphal abdomen, retain the plesiomorphic condition of narrowly open fore coxal cavities. In such otherwise highly derived genera as Lucerocoris and Heinsius this posterior nymphal sclerotization is also absent. However, these genera are so closely related to others in which it is well developed that there seems little doubt that this is a case of secondary loss.

One of the most important features on the cladogram is the acceptance of the closed fore coxal cavities (FcxCv) as being a uniquely derived condition. If this is true, it follows that ancestors similar to modern species of Ischnodemus represent the sister groups of most of the more apomorphous genera. It is possible that a few species that are placed in Ischnodemus because of the closed coxal cavities (particularly such species as *Ischnodemus par*athoracicus, and thoracicus) may have a sister group relationship with some species of Dimorphopterus. The sperm reservoirs of parathoracicus and such a species of Dimorphopterus as zuluensis are very similar. This appears to indicate symplesiomorphy and to not necessarily be indicative of cladistic relationship. The similarity, however, is very striking. To accept the closing of the fore coxal cavities independently two or more times would necessitate the acceptance of the independent development of the posterior abdominal sclerotization two or more times.

The species included in *Ischnodemus* appear to constitute a paraphyletic group. There seem to have been two major "trends" in operation within the complex. One trend is the development of large, flattened platelike wings on the sperm reservoir and a concomitant increase in the size of the bulb of the reservoir itself. A second trend (which probably involves several different evolutionary lines) is one in which reduction takes place. This reduction frequently results in the retention of only a minute bulb, with or without tiny wings present, but in other cases results in the development of a thickened, sclerotized thumblike projection above the ejaculatory tube without wings developed at all.

From various *Ischnodemus* ancestors the cladogram shows five separate lines diverging (a possible sixth line leading to *Patritiodemus* is not shown). Two of these lines "lead," however, only to a single genus each (*Cavelerius* and *Procellademus*). The cladogram of *Ischnodemus* (fig. 42) indicates probable areas within the genus where the sister groups of these five various lines lie. There is a great deal of difference in the strength of the synapomorphies holding these together.

The genus Cavelerius could be synonymized with Ischnodemus. The species differ from species of Ischnodemus primarily in the unusual nature of the pruinosity of the pronotum where the anterior half is completely shining and the posterior half completely pruinose (PrPn1). This has occurred independently in two other cases in the Blissinae both derived from (Caveloblissus Blissus-like ancestors and Praeblissus). In view of the fact that within Ischnodemus we have retained species that are completely pruinose and completely shining one certainly can question the desirability of retaining this as a separate genus. The most apomorphic species have the posterior angles of the pronotum extremely strongly produced backward in a great arc, whereas the more plesiomorphic species do not, and it is this high degree of apomorphy which makes it questionable to synonymize Cavelerius. Almost exactly the same phenomenon is found in the South American genus Patritiodemus. This latter genus has a completely pruinose head and pronotum (plesiomorphic) and rather stalked eyes (apomorphic). Its most apomorphic members also have very enlarged fore femora with great bifid spines present ventrally. The most generalized members are very similar to some species of Ischnodemus. Thus, both Cavelerius and Patritiodemus could conceivably be included in Ischnodemus.

As transformation series occur within "groups" of *Ischnodemus* the above cases are really of a different order of magnitude. Since anagenesis is a consideration in the classification used in this paper *Cavelerius* and *Patritiodemus* are treated as distinct genera.

A very different situation exists for the three other phyletic lines that we believe have developed from "Ischnodemus-like" ancestors. The first of these includes a group of genera held together by a synapomorphic character in the nymphal abdomen. This is the possession of a very large elliptical, ovoid sclerotized plate (SGAI) which extends forward from the anterior margin of each dorsal abdominal scent gland. Other species of the Blissinae have small sclerotized plates around the scent glands (as do almost all the Lygaeidae) but in this case the sclerite is very large and is similar in all taxa that possess it. This is a very strong synapomorphy and is most interesting in that it includes two South American genera and a series of genera from Madagascar, three of which also occur on the African continent. As discussed in the zoogeographic section the South American-Madagascar-African relationship probably indicates relatively early divergence of this stock (West Gondwanaland). Within this clade the Western Hemisphere taxa are the most plesiomorphic. They have a less reduced sperm reservoir and retain thoracic pruinosity which is lost (PrPn3) in Madagascar-African genera. Nevertheless, it is important to point out that this Western Hemisphere clade, which consists of Extarademus and Toonglasa, is in several ways a very specialized one. Extarademus itself shows uniquely derived conditions of spines and secondary sexual projections upon the abdomen which are unusual in the Blissinae. Toonglasa, which is known only from the holotype, is essentially a large flattened Extarademus with long projections of the abdominal connexivum. The flattened body of Toonglasa gives it a superficial resemblance to such genera as Bochrus, Chelochirus, Scansidemus, etc., but the cladistically significant relationships are with Extarademus. Ramadademus is a large flattened blissine. Fortunately, the nymph is known and the abdominal sclerotization is of the type which characterizes the Extarademus clade. Ramadademus is very similar in habitus to such genera as Scansidemus (Oriental) and the two genera have similar (and highly derived) scent gland auricles. Until nymphs of Scansidemus become available there is no way to determine whether Scansidemus belongs in a clade with Ramadademus or with Bochrus and its relatives.

The remaining members of the above clade, which is "held together" by the anteriorly enlarged nymphal scent glands, includes chiefly a Madagascan complex of eight genera, one of which is undescribed. All of these genera have an extremely reduced sperm reservoir, with only a small median, usually curved, hooklike central sclerite present. It is doubtful that the sperm reservoir is functional. Of these seven Madagascan genera (in addition to Ramadademus) three are known to occur in Africa and the other four are restricted to Madagascar. The most plesiomorphic of these appears to be Blissiella, which is also the most widespread on the African continent. This is the only genus of the clade which has a plesiomorphic rounded earlike metathoracic scent gland auricle, all the others having it modified either by elongation or by forming a crescent shape (fig. 9Q). Blissiella also has a short, thickened corium with a concave apical margin. In many ways species of Blissiella are reminiscent of species of *Dimorphopterus* and they certainly appear to remain a number of plesiomorphic conditions.

Of the remaining genera Aradademus, Aradacrates, and Pseudoblissus have a synapomorphous stalked fourth antennal segment. This feature is quite distinctive for these three autochthonous Madagascan taxa. Each of the three is a very highly specialized genus whose species have a number of autapomorphous features. Aradacrates is particularly striking because of the extremely enlarged bucculae which project forward as plates anterior to the end of the tylus (figs. 6E-H, 40), similar to the condition found in some species of *Iphicrates*. However, these two genera, despite the presence of these large bucculae, are not closely related and in Aradacrates enlargement of the bucculae is certainly an independent evolutionary event. The remaining genera of the clade Micaredemus, Merinademus, and Barademus form a very closely related group which is held together by an uniquely shaped metathoracic scent gland auricle which in all the species curves forward in a lunate or crescent-shaped arc (fig. 9Q, 10F).

The other two large complexes of genera that are shown as arising from an *Ischnodemus* ancestry are held together by synapomorphies that are less compelling than those discussed above. The first of these constitutes a clade of genera placed together on the basis of having a reticulate membrane (RtMe). There are difficulties with this interpretation in that two of the genera included actually do not have a reticulate membrane and in those genera that do, the type of membrane surface is not uniform. For example, Reticulatodemus, Xenoblissus, and Gelastoblissus have a series of minute hexagonal cells present over the entire surface of the membrane. In Australian genera such as Heinsius and Slaterellus there is a "dimpling" or corrugating of the surface which may or may not be homologous to the true reticulations. The similarity of many species of *Iphicrates* to species of Reticulatodemus on the one hand and to Xenoblissus on the other seems to justify the inclusion of a genus such as Iphicrates (which does not have a reticulate membrane) in this group and to support the belief that the reticulate membrane is secondarily lost. Iphicrates is an Australian and Oriental genus and Reticulatodemus and Xenoblissus are both Neotropical. It seems highly unlikely that such characters as the hooked antenniferous tubercles and the unusual scent gland auricle shape (which are apomorphic conditions and common to the three genera) can all be the result of convergence.

The final clade derived from Ischnodemus is a group of very large, usually flattened genera with multispinose femora which is held together primarily by the presence of these numerous spines (FrFmSp2). Since multispinose fore femora appear to have arisen independently in the case of Ramadademus there is, of course, no reason why it could not have occurred independently again. Most of the included genera are, however, related to most other genera on the basis of other characters (see fig. 15) even though I have not been able to discover a strong apomorphic character common to all genera. Within this supposed clade the South American genus Patritius is certainly the most plesiomorphic. Not only do these species show a more *Ischnodemus*-like, elongate, relatively slender body form but the pruinosity on the dorsal surface is much more generalized than in the other genera included here. Patritius

is a particularly important genus and one shown on the cladogram as possibly being paraphyletic.

The rationale for treating *Patritius* as possibly paraphyletic is due to lack of information. Several species of *Patritius* with relatively derived pronotal pruinosity patterns are known only from females. These species have short genal projections rather similar to those found in females of such New Guinea genera as *Dentisblissus* and *Scintillademus* where the male gena, particularly in *Dentisblissus*, project forward as hugh "tusks." The pruinosity patterns and scent gland auricles are also similar in these species of *Patritius* to those of the New Guinea genera.

The relationships of the various species of Macropes have been treated by Slater and Wilcox (1973) and that paper should be consulted for details. In the cladogram Macropes is treated as a paraphyletic group. Ischnocoridea and Extaramorphus appear to have sister group relationships with different sections of Macropes. Ischnocoridea is very closely related morphologically to some species of Macropes and it is questionable whether it should have generic status. Extaramorphus, while obviously derived from a Macropes-like ancestor, has a number of striking autapomorphies and certainly on phenetic grounds deserves generic status. The remaining genera that are derived from an ancestral group of Macropes-like species consist of five genera of large generally flattened or cylindrical Oriental insects: Bochrus, Chelochirus, Lucerocoris, Spalacocoris, and Pirkimerus. All of these genera have a greatly reduced sperm reservoir (SRr) and all but Bochrus, which is somewhat isolated, form a very distinctive clade by themselves with synapomorphies evident in the hugh ocelli (OC), strongly clavate antennae (AntCl), a greatly modified, reduced and blocklike clasper (ClB), and a greatly reduced ovipositor (Ovr).

The position of the Oriental genus Scansidemus and the Neotropical genus Riggiella, which are held together within the clade by the multispinose fore femora, is more difficult to understand. It is possible that Scansidemus is the sister group of Ramadademus, but if so this can only be verified by a study of the nymphs. These two genera do have very similar metathoracic scent gland auricles. The phylogenetic position of Riggiella is perhaps the greatest enigma in the Blissinae. There is nothing in the Neotropics that even remotely resembles this genus with its extremely flattened body and huge multispinose legs. The habitus is reminiscent of some of the large Oriental genera. However, the scent gland of Riggiella is quite distinctive and it does not have the enlarged first tarsal segment which is characteristic of the Oriental clade. Again, study of nymphs would be extremely desirable here. It is possible that Riggiella represents an extremely early divergence of a West Gondwanaland stock which is now absent from Africa. It may be the sister group of the Oriental clade.

One of the most interesting features that emerges from the generic cladogram—and which was arrived at completely independently of the zoogeographic analysis—is that in each of the three major phyletic lines leading from *Ischnodemus* the most plesiomorphic genera in each clade are Neotropical. Note that *Praetorblissus* and *Heteroblissus* (fig. 15) that are treated as possibly the most plesiomorphic of all the Blissinae are also Neotropical.

CODE TO CLADOGRAM ABBREVIATIONS

A

AbdPr males with spines and projections on abdominal sternum

AbScabdominal sclerites on nymphs
AbSgWbasal abdominal segments white

AntSt....."stalked" fourth antennal segment
AThk.....antenniferous tubercle hooked

В
BF1broad flattened body BUCmale bucculae greatly produced
C
C1B
E
ELextreme body elongation ETVtransverse eyes
F
FcxCv closed fore coxal cavities FmSw all femora short, stout and swollen FrFmBf fore femoral spine bifid FrFmM fore femora mutic FrFmr fore femora slender and reduced FrFmSp1 fore femora with one (or rarely two) spines FrFmSp2 fore femora multispinose FmMsMt (-) middle and hind femora mutic
G
GNtub genital tubercle prominent GNtubo secondary loss of genital tubercle GT genital tubercle Gtk males with genal tusks enlarged GTI genital tubercle secondary loss
Н
Hemttwo-textured hemelytral membrane HFelhind femora of males greatly enlarged
J
JUGmale juga strongly projecting
М
MEant
o
OCocelli enlarged OVptovipositor paratergites elongated

OVptDovipositor paratergites secondarily divided OVrovipositor reduced and platelike
P
pGtkproclivity for genal tusks PPexpropleuron excavated PRoFmedian longitudinal pronotal furrow PrPn1reduced pronotal pruinosity (dorsally) PrPn2no pronotal pruinosity (dorsally) PrPn3no pleural pruinosity
R
RtMe reticulated membrane on fore wing
s
SCL body with flattened scalelike hairs ScGIA metathoracic scent gland auricle modified ScGIb metathoracic scent gland auricle curved backward ScGICr crescent shaped metathoracic scent gland auricle ScGle metathoracic scent gland auricle elongated ScGlgd metathoracic scent gland auricle widened and enlarged SGAI enlarged SGA sclerite in nymphs SPR, SPr modified sperm reservoir SP71 seventh abdominal spiracle lateral SRr sperm reservoir reduced SRwl sperm reservoir wings absent StS stridulatory structure present
T
TAen

ZOOGEOGRAPHY

There is no meaningful fossil history of the Blissinae and therefore zoogeographic analysis beyond the descriptive must be based upon inference and interpretation of modern distributions.

I have been considerably influenced by the operational logic of the vicariance "model" insofar as it considers that the burden of proof in disjunct distribution situations is on those who believe that such distributions are the results of dispersal. (Platnick and Nelson, 1978). On the other hand, I believe that insects including the Blissinae do disperse. Nevertheless, the com-

plete lack of the Blissinae on oceanic islands¹ of the Pacific and Indian oceans and the paucity of the West Indian fauna cannot help but persuade one that the major features of the distribution are not the result of recent dispersal.

The major features of the distribution of the Blissinae are:

¹Dimorphopterus pilosus (Barber) was described from Yap Island but this probably is a parthenogenetic species and thus stands somewhat apart from the "normal" dispersal potential of the subfamily. *Macropes obnubilus* (Distant) occurs on the Bonins and on Guam but is almost certainly introduced by man.

- (1) Extremely close relationship between the *Ischnodemus* faunas of Africa and South America and paucity of this taxon in the Oriental and Australian regions.
- (2) Presence of a considerable number of highly apomorphic genera (not closely related between regions) endemic to either the Oriental or Neotropical region.
- (3) Closer relationship of the (scanty) Palearctic and Nearctic faunas to the Old and New World Tropics respectively rather than to one another, i.e., no Holarctic element.
- (4) Apparent tropical relationships (not *Ischnodemus*) between the Oriental and Neotropical faunas.
- (5) Impoverishment of the Australian fauna, with a few isolated endemic genera, but with the greater part of the fauna congeneric with Oriental genera and restricted to the north and east.
- (6) Occurrence of several "sister" groups between South America and Africa.
- (7) Radiation of several "stocks" on Madagascar, almost all of which appear to be most closely related to forms from Africa.
- (8) Absence of the subfamily from New Zealand.

Before attempting a general interpretation of the present distribution of the Blissinae, I outline the situation in each major faunal region so that general conclusions can be placed in some perspective relative to more specific ones.

PALEARCTIC FAUNA

The blissine fauna is depauperate, with 18 species present in *Ischnodemus*, *Dimorphopterus*, and *Geoblissus*. All three of these genera occur in both the Ethiopian and Oriental regions and are more extensively developed there. In the genus *Ischnodemus* the majority of species belong to the *sabuleti*-group, which is essentially a Palearctic complex and is somewhat isolated from the extensive Ethiopian fauna. There is no evidence of a direct faunal relationship between the Palearctic and Nearctic regions. Certainly blissine stocks did not disperse through Beringia in the late Tertiary.

NEARCTIC FAUNA

Like that of the Palearctic, the blissine fauna is depauperate (three genera, 29 species). There

has been a rather extensive radiation of chinch bugs (Blissus), especially in western North America and a more limited radiation of the Ischnodemus falicus-group in the southeastern states.

There seems little doubt but that the Nearctic fauna is derived from the Neotropical. Blissus has a number of Neotropical species and there are closely related derivative genera (Praeblissus, Caveloblissus) in the Neotropics.

Two "groups" of *Ischnodemus* are present in North America; one, consisting of *robustus*, *praecultus*, *rufipes*, ¹ and *fulvipes*, appears to represent two or three independent introductions (dispersals) of Neotropical lines into the southeastern states in relatively recent time. *Ischnodemus praecultus* and *fulvipes* are widespread Neotropical species and the others are very closely related to Neotropical taxa.

The second, the *I. falicus* group, is more difficult to understand. It includes all the other species of Ischnodemus in North America. The species are most numerous in the southeast (especially Florida) but there has been speciation in the Great Plains and in the midwestern prairies. The falicus group species are strongly apomorphic, not at all similar to any Palearctic species (and absent from the far western states), but are not obviously closely related to any existing Neotropical species. Most of these falicus group species have a micropterous morph in contrast to the first group noted above. I (Slater, 1977) have suggested that when the living habits of two groups of species are similar, wing reduction in species of one group can be interpreted as indicating that, in the area under consideration, such a group is the older.

Thus, it seems probable that the *falicus* group has been in North America for a relatively long time, and I suggest that both *Blissus* and *I. falicus*-group ancestral stocks entered the United States no later than the Miocene, and probably coincident with the rise and spread of the Madro-Tertiary flora. The only other blissine known to occur north of the Mexican border is *Extarademus macer* (Van Duzee).

¹This species was synonymized with *praecultus* by Slater and Wilcox, 1969. Subsequent collecting in Florida has raised the question whether or not they may be distinct species and until this is resolved I treat them as distinct.

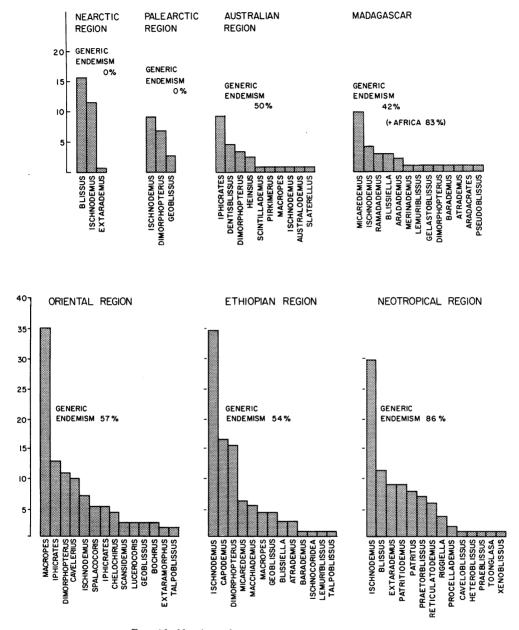


Fig. 16. Number of genera per zoogeographic region.

This is a widespread Neotropical species and a member of an otherwise Neotropical genus. *E. macer* is known to occur north to extreme western Iowa. There it is associated with xeric habitats that contain remnant disjunct colonies of dry adapted western plants. The Madro-Ter-

tiary hypothesis is consistent with a secondary speciation center in Florida. The extension of a "dry adapted" fauna and flora into Florida is well documented and evidenced today by many organisms such as burrowing owls and gopher tortoises.

AUSTRALIAN FAUNA

The Australian blissine fauna is also relatively depauperate (only 24 species) but contains 10 genera half of which are endemic. Despite the similarity in number of species present, the situation in Australia is quite unlike that in the Northern Hemisphere where there is no generic endemicity. This is to be expected in an isolated continent like Australia, but it is puzzling why there are so few species present. It may in part be due to lack of collecting, but is probably primarily due to loss of fauna under conditions of increasing Tertiary aridity.

Zoogeographic analysis is further complicated by the difficulty of determining for the Blissinae just what the limits of the Australian Region should be. If one restricts it to Australia proper one obscures the close relationship between the fauna of northern and eastern Queensland with that of New Guinea. On the other hand, the inclusion of New Guinea and its associated islands (as is done conventionally for mammals) does introduce what is undoubtedly an Oriental element. On the whole, I believe the more inclusive concept to be preferable.

Three genera are restricted to Australia proper (Australodemus, Heinsius, and Slaterellus). All three of these are morphologically isolated, and show a number of autapomorphic features but together probably do form a clade with synapomorphic features such as scalelike hairs, short thick antennae, and structure of the sperm reservoir. I believe that these taxa are remnants of a very old element in the Australian blissine fauna, one that has probably been decimated by increased aridity during the Tertiary. That aridity may have decimated a once more extensive fauna is indicated by the presence of Ischnodemus sordidus

'Slaterellus Drake and Davis was described as representing a distinct subfamily, the Slaterellinae, and has never formally been placed in the Blissinae although several authors have commented on the close relationships. It is unquestionably a blissine and related by several features to *Heinsius* New Synonymy.

Slater in extreme southwestern Australia. Although Ischnodemus is a relatively plesiomorphic genus and is represented in all major zoogeographic regions, this is the only Australian representative. Ischnodemus sordidus is also quite plesiomorphic within Ischnodemus and is the sister species of I. noctulus Distant which occurs in India, Ceylon, and southern Africa. I do not believe such a disjunct distribution can realistically be attributed to dispersal but rather that it is a vicariance pattern reprean old African-Oriental-Australian senting "track" in Croization terminology. If this is true. L. sordidus has been able to survive in the winter rainfall area of the southwest coast of Australia, but has been eliminated by aridity over most of a formerly more extensive range. This certainly seems to parallel the situation for many plants and insects that are now restricted to the southwest corner of Australia.

The remainder of the Australian blissine fauna has had a completely different history. All of these genera either occur in the Oriental Region as well or are closely related to Oriental taxa. They all occur on New Guinea and, where they also occur in Australia, they are restricted for the most part to the tropical and subtropical north and east. Macropes has radiated in the Oriental Region and one widespread species reaches eastern Oueensland. At least half of the species of *Iphicrates* occur on New Guinea and associated islands. In Australia one species extends down the east coast and reaches Tasmania. There is also a species on New Caledonia. The absence of blissines from New Zealand suggests that the presence of a species on New Caledonia may be a dispersal rather than a vicariant event. Pirkimerus is an Oriental genus with one bizarre species found on New Guinea. Dimorphopterus is widespread in the Orient. Dentisblissus and Scintillademus are closely related, strongly apomorphic taxa restricted to New Guinea and northern Queensland.

ETHIOPIAN FAUNA

The fauna is large and diverse (13 genera, 92 species). There is a striking endemic ele-

ment (Capodemus, Macchiademus, and several species in other genera) in the Southwest Cape. The South African fauna has been dealt with extensively by Slater and Wilcox (1973) and the host plant relationships by Slater (1976) and are not elaborated here other than to point out that these Southwest Cape endemics are very ancient plesiomorphic taxa.

Much of the African fauna is a savanna element made up largely of species of *Ischnodemus* and *Dimorphopterus*. As with many other African savanna insects the range of any given species tends to be extensive, often extending from the dry inland areas of the West African savanna in a great arc through east Africa into the low veld of the Transvaal or coastal Natal. There is evidence of speciation in the isolated East African mountains but it is less striking than in many other animal groups (including many of the Lygaeidae).

The distribution of such genera as Lemuriblissus, Barademus, Micaredemus, and Atrademus may reflect a more mesic period in Africa and these taxa may well be at a competitive disadvantage relative to species of Ischnodemus and Dimorphopterus. All four of the above genera also occur on Madagascar where Micaredemus has speciated considerably. Unfortunately, little is known about the biology of these insects.

There is, as in many other taxa, an Oriental relationship in African blissines which is sometimes at the specific level conspecific (Ischnodemus noctulus, I. thoracicus, Geoblissus hirtulus), and sometimes consists of sister taxa (Talpoblissus. Macropes. Dimorphopterus). This would suggest Late Tertiary faunal interchange, but is more likely an older vicariant event. It should be noted that the dominant Ischnodemus groups of Africa are totally absent from the Oriental Region and the numerous highly apomorphic Oriental blissine taxa are absent from Africa. Indeed, one of the striking features of the Ethiopian Blissinae lies in the close resemblance of the Ethiopian and Neotropical Ischnodemus faunas and the absence of Oriental components.

Despite the large number of species and genera in the Ethiopian Region, the species

tend to be without the bizarre sizes, shapes, and ornamentations that occur so frequently in Oriental taxa.

MADAGASCAN FAUNA

The fauna of this great island probably remains poorly known. Most of the 12 genera and 29 species I have described in the past few years. No Hemipterist, much less a blissine or lygaeid specialist, has collected intensively on the island. Nevertheless, a general pattern is evident. I (Slater 1967) previously thought that the greater part of the Madagascan fauna could have resulted from the introduction of only one or two African stocks. However, subsequent African collecting has shown that *Barademus* and *Lemuriblissus*, previously thought to be Madagascan endemics, are present in Africa.

One is tempted to see a vicariance pattern for the Madagascan-African relationships, but unless limited dispersal is admitted this would apparently require the Blissinae to be present in the Triassic and thus such an idea may be untenable.

Most, if not all, of the Madagascan blissine fauna is related to that of Africa, not the Orient. There are Madagascar species that are very closely related to a group of African species but with only a single species on Madagascar. There are other taxa that are congeneric with African species but in which considerable speciation has occurred on Madagascar, and finally there are genera with such strongly autapomorphic characteristics that their affinities are at present difficult or impossible to clearly understand.

Ramadademus, which is an Madagascar genus of large, strongly flattened bugs with multispinose femora, is similar in shape to several Oriental genera and to the Neotropical Riggiella. However, nymphs of this genus have elliptically enlarged sclerites anterior to the scent glands which I consider a synapomorphic character that relates Ramadademus to other Madagascar genera. Thus, as noted above, the similarity of body configuration is convergent.

The Madagascan blissine fauna shows no

more Neotropical relationships than does the African.

ORIENTAL FAUNA

The Blissinae of this huge and varied region are remarkable for two reasons: (1) The presence of a series of genera of large size, many species of which have striking and bizarre structural modifications, including various expressions of sexual dimorphism (otherwise minimal in the subfamily), modification of head spines, buccular enlargements, reduction of genitalia, incrassate and spinose hind legs, flattened bodies. (2) The scarcity of species of *Ischnodemus* and the complete absence of the dominant *Ischnodemus* groups of Africa and South America.

The genus *Macropes* has about the same number of described species in the Oriental Region as *Ischnodemus* does in each of the Ethiopian and Neotropical regions, but too little is known of the biology of species of *Macropes* to indicate to what extent ecological replacement may exist.

It is evident that while relatively dry-adapted plesiomorphic taxa are present (Dimorphopterus), the chief feature of the Blissinae of the Oriental Region is the large number of varied and highly apomorphic taxa. There probably are more, as collecting in recent decades has been sporadic, and for a number of genera that have been known for a long time, only a handful of specimens exist in the museums of the world. This suggest unusually specialized habitats and probably is indicative of species adapted to relatively mesic conditions on at least the "fringes" of the varies montane and rain forest flora.

What little is known of the blissine fauna of the Indonesian islands on the Sunda shelf indicates a close relationship to the Asiatic mainland. The Philippines do have an endemic genus (*Lucerocoris*) and many endemic species but the known fauna is for the most part harmonic.

NEOTROPICAL FAUNA

As is true of many other animal groups the Neotropical blissine fauna is characterized by richness and by a very high degree of generic endemism (86% of the Neotropical genera are endemic as compared with 57% for the Oriental Region and 23% for the Ethiopian—54% if Madagascar is included).

Of the Neotropical subregions, the South American fauna is by far the richest. Indeed, Central America and the West Indies appear to have blissine species that are largely of South American relationship. This is strikingly true of the West Indies from which only seven species have been reported. Of these, four are species of Blissus all so closely related that their status needs careful investigation (all are in disturbed habitats). Of 32 Neotropical species of Ischnodemus only one is known from the West Indies. This is I. fulvipes, which is widespread in South and Central America and also occurs in Florida. There is an endemic species of Extarademus (discalis) and one of Patritius (cubensis) on Cuba. The sister groups are South and Central American. These last two are, in my opinion, the only true endemic blissine elements in the West Indies. It is probably significant that these two species occur on Cuba, as this was probably the only island that remained above sea level during the Miocene. The presence of these two species may represent vicariant events; the other species I believe to be the result of recent dispersal.

The Central American fauna is somewhat more extensive than the West Indian (seven genera, 13 species). However, with the possible exception of *Extarademus* species, the stocks are derived from South America. For example, all three of the Central American *Ischnodemus* species also occur in South America. Central America has only a single species of *Reticulatodemus* and of *Praetorblissus*, the other species all being South American. *Extarademus* has a number of Central American and Mexican species (but is also South American) and its sister group is African.

Although there is evidence of dispersal in *Blissus* and *Ischnodemus*, in general the West Indian and Central American faunas appear to present more of a vicariance pattern.

In South America the most striking aspect of the fauna is the very close relationship of the *Ischnodemus* species to those of Africa. Both regions contain a series of species that are so similar that it is difficult to believe that they come from two continents physically so far removed from one another. Presumably some of this resemblance is due to convergence toward elongate bodies and tan coloration in the East and South African savannas on the one hand and the Argentine and adjacent pampas on the other. It is tempting to consider this a direct effect of continental drift. I suggest, however, that while drift is involved, that the relationship is more remote and subtle than that of a series of sister groups.

When one examines the South American blissine fauna as a whole one is struck by the presence of several New World-Old World "sister" groups at the generic level, with the Neotropical component retaining in each pair the more plesiomorphic features. Examples are Blissus-Dimorphopterus, Extarademus-Micaredemus, Riggiella-Scansidemus, and possibly Patritius-Dentishlissus and Reticulatodemus-Iphicrates. However, this is not the situation in Ischnodemus. In Ischnodemus the species with important plesiomorphic characters are African. The Neotropical fauna is relatively apomorphic and to judge by the appearance of the sperm reservoir probably is a monophyletic unit in itself. Despite the presence of many African species with important plesiomorphic characters the most widespread and abundant savanna species of African Ischnodemus are in contrast even more apomorphic than the Neotropical. I conclude that Ischnodemus-like stocks of a relatively apomorphic nature were present when Africa and South America were still close enough together for considerable faunal exchange to occur, and that the diverse species in South America may be members of a single phyletic line the evolution of which has taken place independent of the subsequent evolution of the genus in Africa.

There is also an apparent Oriental-Neotropical relationship which is more difficult to understand but which also is found in a number of plant groups.

We know too little about the distribution of the Blissinae within South America itself to say more than that there appear to be a number of old and endemic taxa in Peru, and that the Chilean fauna does not appear to be as distinctive and isolated as one might have anticipated.

In general then the hypothesis that seems to

best fit the distributional and cladistic relationships of the Blissinae is the following:

- 1) That the Blissinae originated as a tropical group.
- 2) That the taxon originated in Gondwana-
- 3) That considerable diversification took place before the fragmentation of Gondwanaland, certainly before the breakup of West Gondwanaland.
- 4) That the present fauna of the Northern Hemisphere developed from nearby tropical regions.

Evidence to support this hypothesis has been suggested in a few places above but must be stated more fully here. That the Blissinae originated as a tropical group seems evident. Not only are they much more diverse in the tropics but all of the major phyletic lines are represented. Although host plant information remains scattered, there is a concentration upon tropical groups shown especially in the grasses where bambusoid, panicoid, and eragrastoid grasses are most frequently utilized. The north temperate fauna in both hemispheres is very depauperate, the Nearctic and Palearctic faunas not closely related and each quite easily derivable from adjacent tropical areas. In the Southern Hemisphere the Blissinae are absent from New Zealand, and almost absent from temperate Australia. In the rich veld and pampas regions of southern and eastern Africa and in South America the fauna is reasonably diverse and in the former contains some old elements. but it should be remembered that southern Africa is certainly warm temperate and extends to less than 35°S. Recognition of the Blissinae as a tropical group must be considered when one attempts to establish age of the group. The several sister group relationships noted above between Africa and South America strongly suggest that the Blissinae were in existence by or soon after 96 m.y. (B.P.) the date generally accepted for the last (and tropical) connection between South America and Africa. There is no present evidence for an ancient (or Recent) northern dispersal between hemispheres for this subfamily.

The first problem to be faced in attempting to interpret the distributional patterns discussed above is to try to understand how old the Blissinae are. In the absence of any meaningful fossil record, this question must necessarily be approached through the use of indirect evidence and by comparing the distributions with those of other groups.

As I have previously discussed (Slater, 1976), modern Blissinae are known to breed only on monocotyledonous plants. Since many families of monocots serve as host plants there must be an absolute physiological dependence on the part of the insects upon some feature of monocot plants. Therefore, it is highly unlikely that the subfamily would have existed as a recognizable entity before the existence (and before some degree of differentiation) of the monocots. Further, the other subfamilies of lygaeid bugs for the most part feed on the seeds of angiosperms. The origin of the angiosperms is still a matter of much debate. Raven and Axelrod (1972) noted that pollen that can definitely be referred to any living genera is first known from the middle of the Upper Cretaceous (about 75 m.y. B.P.). There is no a priori reason why blissine genera should be older or younger than plant genera, but it is my impression that generally a plant "genus" is a more inclusive category than are most animal genera. Thus, for a working hypothesis it seems reasonable to think that a living animal genus might not be older than all of the plant genera with which the group is associated. This would lead to the conclusion that the Blissinae did not originate earlier than some time in the Upper Cretaceous, and probably arose coincident with the radiation of the monocot angiosperms. Further evidence that the Blissinae may not be appreciably older is their complete absence from New Zealand. This suggests that the group was not present in temperate West Antarctica 80 m.y.B.P. when New Zealand broke away. This, of course, does not mean that blissines did not exist at that time. They appear to have originated as a tropical group, whereas the Late Cretaceous climates of New Zealand and West Antarctica were temperate and possibly cool temperate during most of this period (Cracraft, 1973). The presence of at least three and probably several more sister groups between Africa and South America

would argue for some diversification of the Blissinae before Africa and South America separated in the Upper Cretaceous (some 80-90 m.y.B.P. depending upon which authority one follows). As Raven and Axelrod (1972) pointed out, however, for organisms with some ability to cross water gaps, time of actual marine transgression between two continental masses is certainly not coincident with the last period of extensive faunal exchange. To the contrary, exchange will take place for a very long time subsequently, although at an ever diminishing rate as the two land areas move farther and farther away from each other. Thus, the Blissinae need not have been especially diversified at the time of the final separation of Africa and South America, but they must have been diversified within a reasonable time thereafter.

The result of this is that all the (admittedly scanty) evidence points to the occurrence of the Blissinae with some degree of differentiation in the Upper Cretaceous. This will be used as a working hypothesis in attempting to understand present distributions.

POSSIBLE GONDWANALAND RELATIONSHIPS

AUSTRALIA: The limited "autochthonous" Australian fauna appears to have its closest cladistic relationships with Madagascar (the evidence is not compelling). There is also a relationship to southern Africa and to India. Ischnodemus sordidus is known to occur only in the winter rainfall area of extreme south-Australia. Ιt is relatively plesiomorphous member of the genus and shares several synapomorphic features with Ischnodemus noctulus Distant. The latter has a disjunct distribution in India, Ceylon, Southeast Asia, Indonesia and southern Africa. The two species are moderately closely related morphologically and quite isolated from other species of *Ischnodemus*. It is interesting that neither species occurs on Gramineae but on monocot taxa generally considered to be older (Cyperaceae, Zingiberaceae). It seems unlikely that the disjunct distribution of noctulus between Asia and Africa is a drift phenomenon

but rather is due to Tertiary climatic deterioration between the two areas, as is discussed later. The relationship between noctulus and sordidus may, however, be related to drift through a common ancestor. Cracraft (1973) discussed conflicting viewpoints as to whether or not India was in contact with Australia. Despite the ambiguity of the geological evidence the presence of a thaumastocorid bug in India (Drake and Slater, 1957), a family otherwise known in the Eastern Hemisphere only from Australia, persuades me that former close faunal interchange did take place. Veevers, Jones, and Talent's (1971) reconstruction of Gondwanaland shows southeastern India in close contact with southwestern Australia. My suggestion is that noctulus and sordidus do illustrate a drift relationship although not necessarily indicating or implying actual continental contact.

Heinsius, Australodemus, and Slaterellus, the other autochthonous Australian genera, are highly derived taxa and have no close relatives Africa. India. South America. or Cladistically, they appear to be most closely related to Madagascan taxa (see Cladogram) but the evidence is scarcely compelling. MacKerras (1964, 1970) referred to an "old northern" or "Lemurian" element in the Australian fauna that consists of groups that "have centers of origins in Africa or Madagascar, extended around or across the Indian Ocean." If this fauna is real in the Blissinae, it suggests either astonishing dispersal across the Indian Ocean or extinction (or non-collecting) in the Oriental Region. The Australian taxa are dry-adapted and possibly the remnants of a more extensive fauna. It seems certain that they are not derived directly from the existing Oriental fauna or vice versa. These genera may represent disjunct Gondwanaland elements from a period when Australia, Antarctica, and Madagascar were closer together. However, if MacKerras's "Lemurian" distributions are real, in the absence of Indian representatives, I cannot explain them other than by extinction in India as it drifted through many degrees of latitude. Cracraft (1973) believed "it is not yet possible to extrapolate the continental position (for the

Indian Ocean) of 75 m.y. back to any particular reconstruction of Gondwanaland."

The "tropical" Australian genera do have Oriental affinities. Iphicrates, for example, though richest in species in New Guinea and its associated islands, is also represented in Southeast Asia, India, and Ceylon. The intriguing aspect of this fauna is its apparent relationships to tropical and subtropical genera in South and Central America. It seems most unlikely that such relationships are the result of dispersals through the Antarctic. Nor, despite Raven and Axelrod's (1972) discussion of the massive extinction of plants in Africa, does it seem likely to me that these Australasian-Neotropical relationships are old Gondwanaland elements which have been eliminated in much of the Old World Tropics. I am intrigued by the idea of Nelson and Platnick (personal commun.) who suggest the "hybrid" nature of continental faunas. For South America this would be the result of an early separation in the Pacific that carried part of the "Oriental" fauna eastward until the plate collided with a westward moving plate carrying faunal elements of African relationship. This would explain the composition of the South American blissine fauna very well and make understandable the Neotropical rather than Old World tropical affinities of several of the tropical Australian-New Guinea blissine genera.

SOUTH AMERICA: The cladistic affinities of such South American genera as *Xenoblissus*, *Reticulatodemus*, and *Patritius* to Australian or Asian genera such as *Iphicrates*, *Dentisblissus*, and *Scintillademus* are those discussed above.

There is a closer relationship between several Neotropical and African taxa where definite sister group relationships are present. These are *Dimorphopterus-Blissus*, *Micaredemus-Extarademus*, and the "advanced" sections of *Ischnodemus*. Interestingly, in each of these cases the Neotropical representative is the most plesiomorphic. This seems to be an illustration of the isolation of South America as a great island for most of the Tertiary, thus allowing the survival there of relatively primitive stocks. While the *Dimorphopterus-Blissus* and *Micaredemus-Extarademus* relationships appear to be

drift phenomena and therefore one may postulate that their ancestor existed in West Gondwanaland ca. 75-90 m.y.B.P., the situation with Ischnodemus is less clearly identifiable as a drift associated event, although it may be. Some of the Neotropical and Ethiopian species of Ischnodemus are extremely similar, so much so that it is hard to imagine that they would not have diverged more than they have over such an immense period of time. The sperm reservoirs of all Neotropical Ischnodemus species are all of a highly apomorphic type and very similar to one another. In Africa, by contrast, many types of sperm reservoirs are present. It is possible that the ancestral Neotropical Ischnodemus could have arrived by over-water transport long after an extensive water gap had developed between Africa and South America. If so there has been time for much speciation and a great deal of morphological diversification to have taken place.

AFRICA: Distributions in Africa that appear to be best understood as related to the breakup of Gondwanaland are those already discussed relative to Australia and to South America. The plesiomorphic nature of such taxa as Capodemus and Macchiademus, now restricted entirely or largely to the southwest Cape Floral Kingdom, suggests that such taxa are the remnants of the "old" blissine fauna.

In summary then, it may be said that present blissine distributions and relationships strongly suggest that the Blissinae arose in tropical areas of Gondwanaland and that the Australian, Neotropical, and Ethiopian faunas show relationships that seem to be best explained by the breakup of Gondwanaland.

POST-DRIFT DISTRIBUTIONS

ORIENTAL REGION: One of the most striking features of blissine distribution is the paucity of *Ischnodemus* species in the Oriental Region. Only a few rather plesiomorphic species of *Ischnodemus* are present and the dominant groups within *Ischnodemus*, such as the *grossus* and *stali* groups with enlarged sperm reservoirs, are entirely absent. This is especially striking when one realizes the close relationship

of these very groups of Ischnodemus in Africa and South America. There are close relationships between Africa and Asia in other taxa of the Blissinae. Dimorphopterus gibbus, a widespread Oriental species, also occurs in West Africa. There are sister species of Talpoblissus in the two regions; two "groups" of Macropes are present in both areas and there are related species of Geoblissus and a number of closely related species of *Dimorphopterus*. All of this would seem to argue for a former close faunal connection between the two regions and to make it even more difficult to understand why the apomorphic Ischnodemus groups are absent from Asia. I believe this apparently anomalous situation is most reasonably understandable by interpreting the Talpoblissus, Geoblissus, and Dimorphopterus relationships between the two relatively continents as recent, probably Pleistocene events. These groups presumably originated in Africa and dispersed into Asia and the Palearctic during the Pleistocene periods when the xeric climates were less rigorous than at present. The appearance of the Ischnodemus situation may be exaggerated by taxonomy. There is an apparently plesiomorphic (undescribed, one micropter) blissine related to Ischnodemus in the mountains of Burma and the species of the genus Cavelerius could, of course, be considered to be only a group of Ischnodemus. This taken together with the presence in Asia of at least two (probably three) generalized Ischnodemus "groups" (noctulus, nigrocephalus and relatives, thoracicus) clearly shows that Ischnodemus species were present in Asia early. The lack of "advanced lines" seems to me to probably be due to the presence of a diverse *Macropes* fauna that ecologically replaces Ischnodemus to a large extent.

Ischnodemus-like ancestral "stocks" appear to have been present in Africa and South America for a long time. Nevertheless in both continents some of the more apomorphic Ischnodemus clades have been undergoing active speciation in relatively recent geological time. I believe that at least a considerable proportion of this speciation is best accounted for by Pleistocene events. Pluvial periods would have reduced the savanna to widely separated areas

where isolation would lead to speciation. Subsequently, dry periods would favor the spread of the savanna at the expense of the forest and allow the formerly limited savanna species to spread widely in both continents [see Moreau (1966) for Africa; Brown and Benson (1977) for South America].

MADAGASCAR: The present blissine fauna of this great island is somewhat of an enigma. On the one hand, there is considerable endemism at the generic level with radiation of some groups. On the other hand, the relationships of the fauna are almost entirely with Africa. Some of these African-Madagascan relationships are at a conspecific level and others of closely related sister species. Thus, at least some of the Madagascan species must have been acquired by dispersal across the Mozambique Channel. There are a number of lygaeid species in other subfamilies that occur in both Africa and Madagascar. Many of these tend to be "fugi-

tive" species associated with disturbed or early succession habitats. This suggests to me that there is considerable lygaeid faunal exchange at time Africa between present Madagascar. On the other hand, the degree of morphological differentiation of some of the endemic genera would at least suggest that a portion of the Madagascan fauna has been evolving on the island for a long time. However, if it is true that there has not been a direct land connection between Africa and Madagascar since the Triassic this would seem much too early for a group of insects that feed on monocot plants. Thus, the ancestors of these "older" faunal elements would appear to have reached Madagascar when the Mozambique Channel was much narrower or by stepping stones such as through the Comoro Islands. Unfortunately, as previously stated, we probably know too little about the blissine fauna of Madagascar to speculate further.

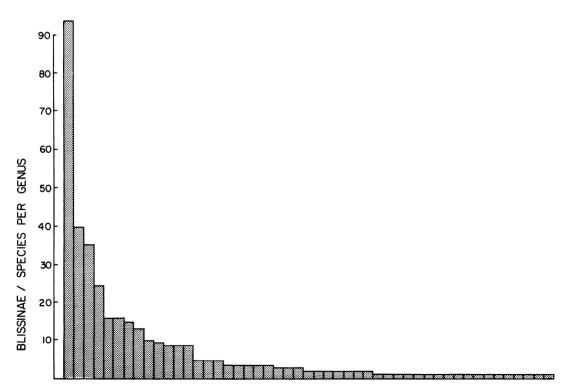


Fig. 17. Number of species per genus.

CHECK LIST OF THE BLISSINAE OF THE WORLD1

ARADACRATES

1. cochlear S. & W., 1969

ARADADEMUS

1 mirificus Slater, 1967

2. oculatus Slater, 1967

ATRADEMUS

1. allaudi Slater, 1967

2. capeneri (Slater), 1964 (Ischnodemus)

3. fusconervosus (Stål), 1855 (Micropus) synonym strigatus Walker, 1872

4. maritimus S. & W., 1973

AUSTRALODEMUS

1. elongatus S. & S., 1963

BARADEMUS

1. attenuatus Slater, 1967

BLISSIELLA

1. castanea (Slater), 1964 (Ischnodemus)

2. castanoides Slater, 1967

3. micans (Slater), 1964 (Ischnodemus)

4. nidus (Slater), 1964 (Ischnodemus)

5. pauliani Slater, 1967

BLISSUS

1. antillus Leonard, 1968

2. arenarius Barber, 1918 subspecies maritimus Leonard, 1966

3. barberi Leonard, 1968

4. bosqi Drake, 1940

5. brasiliensis Drake, 1951

6. breviusculus Barber, 1937

7. canadensis Leonard, 1970

8. hygrobius (Jensen-Haarup), 1920 (Mendocinia)

9. insularis Barber, 1918

10. iowensis Andre, 1937

11. leucopterus (Say), 1831 (Lygaeus)

synonyms: albipennis Dallas, 1852

albivenosus Fitch, 1856 (Micropus) apterus Fitch, 1856 (Micropus)

basalis Fitch, 1856 (Micropus)

devastator LeBaron, 1850 (Rhyparochromus)

dimidiatus Fitch, 1856 (Micropus)

femoratus Fitch, 1856 (Micropus)

Madagascar

Madagascar

Madagascar

Madagascar South Africa South Africa

South Africa

Australia

Madagascar, South Africa

Madagascar, Africa

Madagascar

Zaire, West Africa

Sudan

Madagascar

Puerto Rico

Eastern United States and Canada,

Southeastern United States

Texas

Argentina, Brazil

Brazil

Connecticut, Massachusetts, Maine

Alberta, Saskatchewan, Montana

Argentina

Southern United States, Neotropical

Iowa, Kansas, Missouri

Central & Eastern United States

1S. & W., Slater and Wilcox; S. & S., Slater and Sweet; S. & M., Slater and Miyamoto; S. & A., Slater and Ahmad; S.A.W., Slater, Ashlock, Wilcox; W. & S.,

Wagner and Slater; S. & H., Slater and Harrington; S. & Ash., Slater and Ashlock.

fulvivenosus Fitch, 1856 (Micropus)

6. nigrolimbatus S. & M., 1963

immarginatus Fitch, 1856 (Micropus) melanosus Riley, 1875 (Micropus) nigricornis Fitch, 1856 (Micropus) rufipedis Fitch, 1856 (Micropus) validus Blatchley, 1926 subspecies: hirtus Montandon, 1893 Eastern United States 12. minutus (Blatchley), 1925 (Ischnodemus) Florida synonym: pusillus Blatchley, 1925 (Ischnodemus) (preocc.) 13. mixtus Barber, 1937 California 14. nanus Barber, 1937 Kansas, Missouri? 15. occiduus Barber, 1918 Western United States, Canada 16. omani Barber, 1937 17. parasitaster (Bergroth), 1903 (Neoblissus) Argentina, Brazil, Uruguay 18. penningtoni Drake, 1941 Argentina 19. planarius Barber, 1937 Colorado, Kansas, Wyoming 20. planus Leonard, 1968 Grenada (West Indies) 21. pulchellus Montandon, 1893 Honduras, Panama, Costa Rica 22. richardsoni Drake, 1940 Peru, Argentina, Brazil 23. slateri Leonard, 1968 Puerto Rico 24. sweeti Leonard, 1968 Texas, Mexico 25. villosus Barber, 1937 California 26. weiseri (Drake), 1951 (Neoblissus) Argentina 27. yumana Drake, 1951 Arizona **BOCHRUS** 1. foveatus Distant, 1879 Vietnam, Assam, Burma, India synonyms: hoabinhensis Distant, 1918 tonkinensis Distant, 1918 2. poecilopterus Stål, 1861 Java **CAPODEMUS** 1. bispinosus S. & S., 1972 South Africa 2. darwini (Slater), 1964 (Ischnodemus) South Africa 3. distinctus S. & S., 1972 South Africa 4. elegiae S. & S., 1972 South Africa 5. herbosus S. & S., 1972 South Africa 6. hirsutus S. & S., 1972 South Africa 7. navis (Slater), 1964 (Blissus), new combination Rhodesia 8. pentameri S. & S., 1972 South Africa 9. rostratus (Slater), 1964 (Blissus) South Africa 10. rusticoides S. & S., 1972 South Africa 11. rusticus (Stål), 1865 (Blissus) South Africa 12. sabulosus S. & S., 1972 South Africa 13. stuckenbergi S. & S., 1972 South Africa 14. tenuatus S. & S., 1972 South Africa 15. variabilis S. & S., 1972 South Africa 16. wilcoxae S. & S., 1972 South Africa **CAVELERIUS** 1. antennatus S. & M., 1963 Assam, Burma 2. excavatus (Distant), 1901 (Macropes) India 3. illustris Distant, 1903 Burma, Malaya 4. minor S. & M., 1963 India 5. mishmiensis S. & M., 1963 Burma, India

Burma

7. obscuratus S. & M., 1963

8. saccharivorus (Okajima), 1922 (Blissus)

9. sweeti S. & M., 1963

10. tinctus (Distant), 1904 (Macropes), new combination

CAVELOBLISSUS

1. americanus S. & W., 1968

CHELOCHIRUS

1. atrox Spinola, 1839

synonym: fasciatus (Distant), 1901 (Macropes)

2. confertus S. & A., 1965

3. pirkimeroides S. & A., 1965

4. talpus (Walker), 1872 (Ischnodemus)

DENTISBLISSUS

1. corniger Slater, 1968

2. divisus (Walker), 1872 (Ischnodemus)

3. umbrosus Slater, 1968

4. venosus (Breddin), 1900 (Ischnodemus)

synonym: humboldti (Distant), 1903 (Macropes)

DIMORPHOPTERUS¹

1. annulatus (Slater), 1964 (Blissus)

2. anomalus Slater, 1974

3. atromaculatus (Distant), 1909 (Ischnodemus)

4. bicoloripes (Distant), 1883 (Blissus)

5. blissoides¹ (Baerensprung), 1859 (Micropus)

synonyms: japonicus¹ (Hidaka), 1959 (Blissus)

signoreti (Kuschakevitch), 1861 (Micropus) staphylinus (Jakovlev), 1874 (Ischnodemus)

6. brachypterus (Rambur), 1839 (Pachymerus) synonym: curtulus (Dohrn), 1860 (Micropus)

7. cornipes (Hesse), 1925 (Blissus)

8. cornutus Slater, 1974

9. doriae (Ferrari), 1874 (Blissus) variety: obscurus Reuter, 1888

10. erebus (Distant), 1909 (Ischnodemus)

11. fulgidus (Slater), 1964 (Blissus)

12. gibbus (Fabricius), 1793 (Acanthia)

synonyms: kyushensis (Hidaka), 1959 (Blissus)

sauteri (Bergroth), 1914 (Ischnodemus)

13. graminum (Lindberg), 1958 (Blissus)

14. hessei (Slater), 1964 (Blissus)

15. hirsutulus (Bergroth), 1916 (Blissus)

16. indicus Slater, 1974

17. latoides (Slater), 1967 (Blissus)

synonyms: obscurus (Slater), 1964 (Blissus) (preocc.)

18. latus (Distant), 1909 (Euhemerus)

19. lepidus S. & W., 1969

20. littoralis S. & W., 1973

21. nubicus (W. & S.), 1964 (Stenoblissus)

22. oblongus (Stål), 1865 (Blissus)

Nepal

Japan, Okinawa, Taiwan, Amami-

Oshima (Ryukyus) Pakistan, India

India

Paraguay, Brazil

Borneo, Java, Malaya

Sumatra

Borneo

Malaya, Sumatra, Java

Australia

Australia New Guinea

New Guinea, New Britain, "Ma-

laya," New Ireland

Africa

India

India

China, Japan, India, Thailand

Eastern Palearctic, Japan

Southern Europe, Africa

West Africa, Southwest Africa

Australia

Southern Palearctic

India

South Africa

India, China, Philippines, Java, New

Guinea, New Britain

Cape Verde Islands

South Africa, Rhodesia

Madagascar India

South Africa

Thailand, Vietnam, India, Ceylon

Thailand South Africa Egypt, Sudan South Africa

synonym: longirostris (Stål), 1874 (Blissus)

¹Subsequent to submission of the present paper Josifov, M. and Kerzher, I. M. have published a paper (Heteroptera of Korea II. 1978. Fragmenta Faunistica [Warsaw]) vol. 23, pp. 137-196 in which obsoletus Jakovlev is synonymized with blissoides; and thoracicus Jakovlev and japonicus Hidaka synonymized with spinolae.

23. obsoletus (Jakovlev), 1881 (Ischnodemus)¹

24. pallipes (Distant), 1883 (Blissus)

25. pilosus (Barber), 1958 (Caenoblissus)

26. rondoni S. & W., 1969

27. similis (Slater), 1964 (Blissus)

28. spinolae (Signoret), 1857 (Micropus) variety: geniculatus Horvath, 1882

29. sumatrensis Slater, 1974

30. syrtis S. & W., 1973

31. tenuatus (Slater), 1964 (Blissus)

32. thoracicus Jakovlev, 1881¹

33. typicus (Distant), 1909 (Esmun)

34. upembensis (Slater), 1964 (Blissus)

35. zuluensis (Slater), 1964 (Blissus)

EXTARADEMUS

1. collaris (Signoret), 1857 (Micropus)

2. collaroides S. & W., 1966

3. discalis (Barber), 1947 (Ischnodemus)

4. humerus S. & W., 1966

5. macer (Van Duzee), 1921 (Ischnodemus)

synonym: cahabonesis (Distant), 1893 (Ischnodemus)

umbratus (Distant), 1893 (Ischnodemus)

6. mundus S. & W., 1966

7. tumerosis S. & W., 1966

8. tylosis S. & W., 1966

EXTARAMORPHUS

1. magnatarsus S.A.W., 1969

GELASTOBLISSUS

1. rugosus S. & W., 1969

GEOBLISSUS

1. barchanorum (Kiritshenko), 1913 (Blissus)

2. hirtulus (Burmeister), 1835 (Blissus)

synonym: rotundatus Hidaka, 1959

3. magnofuscus S. & W., 1973

4. mekongensis S.A.W., 1969

5. niger (Slater), 1964 (Blissus)

6. putoni (Jakovlev), 1875 (Blissus)

7. siccus S. & W., 1973

HEINSIUS

1. explicatus Distant, 1901

synonym: anthropophagorum (Kirkaldy), 1908 (Macropes)

2. pallidus S. & S., 1963

Turkestan, Caucasus

Japan

Yap, Bismarcks, New Guinea, Aus-

tralia

Thailand

South Africa, Zaire, Senegal

Palearctic

Sumatra

South Africa

South Africa. Zaire. Sudan?.

Nigeria? Siberia

India

South Africa, Zaire, Chad

South Africa, Mozambique

Bolivia. Argentina, Venezuela.

Colombia

Brazil, Argentina

Cuba

Panama

Southwestern United States, Neo-

tropical

Mexico

Panama, Guatemala

Mexico

Vietnam

Madagascar

Turkestan

"Fr. Sudan," Egypt, Sudan, Cyprus,

Syria, India, China, Japan, Thai-

land, Borneo

Southwest Africa

Laos

South Africa

Southern Palearctic

Angola

Australia

Australia

HETEROBLISSUS

1. anomilis Barber, 1954

IPHICRATES

1. angulatus Slater, 1961

2. cervinellus Slater, 1961

3. gressitti Slater, 1966

4. lativentris (Bergroth), 1918 (Anisosoma)

5. lineatus Slater, 1961

6. malayensis Slater, 1961

7. montaguei (Distant), 1920 (Macropes)

8. neotenicus Slater, 1966

9. nigritus Slater, 1961

10. papuensis Slater, 1961

11. pseudolineatus Slater, 1968

12. rex Slater, 1966

13. spathus Slater, 1961

14. spinicaput (Scott), 1874 (Ischnodemus)

15. subauratus Distant, 1903

ISCHNOCORIDEA

1. elegans Horvath, 1892

synonym: picipes (Haglund), 1895 (Ischnocoridella)

ISCHNODEMUS

1. agilis (Spinola), 1852 (Micropus)

2. ambiguus S.A.W., 1969

3. antennatus S. & W., 1969

4. asciaformis S. & H., 1969

5. atricolor Berg, 1892

6. badius Van Duzee, 1909

7. basalis Walker, 1872

8. basilewskyi Slater, 1964

9. bequaerti Slater, 1964

10. bosqi S. & W., 1969

11. brevicornis (Stål), 1855 (Micropus)

synonym: curticornis Stål, 1874

12. brevirostris Bergroth, 1916

13. brincki Slater, 1964

14. brunnipennis (Germar), 1837 (Pachymerus)

15. canaliculus Slater, 1964

16. canus Slater, 1967

17. caspius Jakovlev, 1871 varieties: nigricornis Stichel, 1958

tetricus Stichel, 1958

18. congoensis Slater, 1964

19. conicus Van Duzee, 1909

20. consobrinus (Distant), 1918 (Macropes)

21. crassipes Slater, 1964

22. dentatus Wagner, 1963

23. diplachne S. & H., 1970

24. discolor (Walker), 1870 (Micropus)

25. falicus Say, 1831

synonym: punctatus (Provancher), 1872 (Rhyparochromus)

26. fallax S. & H., 1970

Argentina, Brazil

New Guinea, Solomons

New Guinea

Taiwan

Malaya, Philippines

Australia, New Guinea

Malaya, Sumatra

New Caledonia

New Guinea

New Guinea

New Guinea

Australia

Philippines

Australia, Tasmania

Japan

Ceylon

Ghana, Cameroon, Congo Republic

Argentina, Chile Thailand, Vietnam

Argentina

Zaire, Cameroon, Ghana, Guinea

Uruguay, Argentina

Southeastern United States

South Africa, Rhodesia

Cameroon, Congo Republic, Equa-

torial Guinea, Zaire

Zaire, South Africa

Brazil, Argentina, Uruguay

South Africa, Chad, Angola, South-

west Africa, Ruanda, Zaire

West Africa

South Africa

Southeastern United States

South Africa

Madagascar

Southern Palearctic

Zaire

Southeastern United States

India Zaire

North Africa

South Africa, Rhodesia, Botswana

Egypt

Eastern United States

Africa

27. formosensis S. & W., 1969

28. fulvipes (DeGeer), 1773 (Cimex)

synonyms: longus Walker, 1872

sallei (Signoret), 1857 (Micropus)

subspecies: schaffneri S. & W., 1969

29. fumidus S.A.W., 1969

30. gayi (Spinola), 1852 (Micropus)

synonym: flavitarsus (Reed), 1900 (Romicpus)

31. genei (Spinola), 1837 (Micropus)

synonym: championi Saunders, 1876

32. grossinigrus S. & W., 1969

33. grossus Slater, 1964

34. hesperius Parshley, 1922

synonym: brevicornis Parshley, 1922 (Preocc.)

35. inambitiosus B. White, 1879

36. inornatus S. & H., 1970

37. jaxartensis Reuter, 1885

38. lactipennis S. & W., 1969

39. linearis (Stål), 1855 (Micropus)

synonym: ochripoides Slater, 1964

40. lobatus Van Duzee, 1919

41. madagascariensis S. & W., 1970

42. mendax S. & H., 1970

43. missouriensis Froeschner, 1944

44. montanus S. & H., 1970

45. neotropicalis S. & W., 1969

46. nigripes Stål, 1874

47. nigrocephalus S.A.W., 1969

48. nigromaculatus S. & W., 1969

49. nigrostillatus Stål, 1858

50. nigrovenosus S. & W., 1969

51. noctulus Distant, 1901

synonyms: macrotomus Bergroth, 1894

nubilis Slater, 1964

52. notandus S. & W., 1969

53. oblongus (Fabricius), 1803 (Lygaeus)

synonym: variegatus (Signoret), 1857 (Micropus)

54. obversus S. & H., 1970

55. ocellaris S. & H., 1970

56. ochripes (Stål), 1855 (Micropus)

synonyms: ochropus (Stål), 1865 (Blissus) quadrispinosus Slater, 1964

57. oculatus Slater, 1967

58. parabasalis Slater, 1964

59. paramoides S. & W., 1969

60. parathoracicus S. & W., 1970

61. perplexus S. & H., 1970

62. praecultus Distant, 1883

synonym: atramedius Blatchley, 1926

63. proprius Slater, 1966

64. pseudotibialis S. & W., 1969

65. pulchellus S. & W., 1969

Argentina, Brazil

South America. Central America.

West Indies, Florida

Mexico Thailand

Chile

Southern Palearctic

Argentina

East Africa

Central United States

Brazil

Tanzania, Kenya

Southern Palearctic

Argentina, Brazil, Paraguay

South Africa, Rhodesia, Zaire

Southeastern United States

Madagascar

Tanzania, Zaire, Congo Republic

Central United States

Kenva

Paraguay, Argentina

Colombia, Panama

Laos

Brazil, Argentina, Bolivia

Brazil

Brazil

Rhodesia, Ceylon, Sumatra, Malaya,

Vietnam, Thailand, Java, Borneo

Venezuela, Bolivia, Brazil, Guyana,

Trinidad, Paraguay

South and Central America

Zaire, Zambia, Tanzania, Mozam-

bique, South Africa

Ethiopia, Kenya, South Africa

South Africa

Madagascar

South Africa, Rhodesia, Zaire, Tan-

zania

Argentina, Paraguay

South Africa, Mozambique

Northern Ethiopian Region

South and Central America, South-

ern United States

Brazil, Argentina

Brazil

Brazil, Paraguay

5. nigritus S. & W., 1973

Argentina 66. pullus S. & W., 1969 Southern Palearctic 67. quadratus Fieber, 1837 synonyms: atlanticus Lindberg, 1932 parallelus (Costa), 1841 (Pachymerus) 68. ranavalonus S. & W., 1970 Madagascar Florida 69. robustus Blatchley, 1926 70. rottensis Statz and Wagner, 1950 Oligocene (Germany) 71. rufipes Van Duzee, 1909 Southern United States synonyms: intermedius Blatchley, 1926 Palearctic 72. sabuleti Fallen 1826, (Lygaeus) synonyms: brunnipennis Rey, 1887 (preocc.) decuratus Herrich-Schaeffer, 1837 (Pachymerus) palustris Carayon, 1944 73. schoutedeni Slater, 1964 Malawi, Tanzania, Zaire, Ruanda 74. severus S. & W., 1970 Argentina 75. signoreti Berg, 1883 Argentina Malaya, Burma, China, Vietnam, 76. sinuatus S.A.W., 1969 Thailand, Borneo, New Guinea, Nepal 77. slossoni Van Duzee, 1909 Eastern United States 78. sordidus Slater, 1968 Western Australia Brazil, Argentina, Chile 79. spatulatus S. & W., 1969 80. stali (Signoret), 1858 (Micropus) Argentina, Uruguay 81. staliellus S. & W., 1969 Argentina 82. subflavus S. & W., 1969 Argentina 83. suturalis Horvath, 1883 Southern Palearctic 84. tenebrosus S. & H., 1970 South Africa, Rhodesia 85. thoracicus (Distant), 1909 (Macropes) Nepal, Vietnam, South Africa South America 86. tibialis Stål, 1858 87. tibialoides S. & W., 1969 Paraguay 88. torquatus S. & H., 1970 Ethiopia 89. transitius S. & W., 1969 Brazil 90. ulugurus Scudder, 1962 Tanzania 91. umbrosus S. & H., 1970 Zambia, Zaire 92. venustus Slater, 1964 Urundi, Ruanda, Kenya, Zaire, Congo Republic Zaire 93. wittei Slater, 1964 94. zavattarii Mancini, 1953 Ethiopia, Kenya, Sudan, Tanzania, South Africa, Zaire ?95. aleocharoides (Jakovlev), 1905 Palearctic (Dimorphopterus), new combination **LEMURIBLISSUS** 1. acuminatus Slater, 1967 Tanzania, Madagascar LUCEROCORIS 1. brunneus Slater, 1968 **Philippines** 2. nigrotibialis Slater, 1968 **Philippines MACCHIADEMUS** 1. acuminatus S. & W., 1973 South Africa 2. angustus S. & W., 1973 South Africa 3. capensis (Slater), 1964 (Ischnodemus) South Africa 4. diplopterus (Distant), 1903 (Blissus) South Africa

South Africa

MACROPES 1. africanus S. & W., 1973 Tanzania. Zaire 2. albosignatus Distant, 1918 Africa 3. alternatus S. & W., 1973 India 4. australis (Distant), 1901 (Ischnodemus) Australia, Philippines, Ceylon, New synonym: longurio (Bergroth), 1918 (Rhabdomorphus) Guinea, Malaya, Taiwan, Hainan Island Tanzania, Chad 5. bacillus (Gerstaecker), 1873 (Ischnodemus) India 6. brunneus S. & W., 1973 Burma 7. burmanus S. & W., 1973 Cambodia 8. comosus S.A.W., 1969 India 9. consimilis Distant, 1918 Pakistan 10. crassifemur S. & W., 1973 India, Burma, Ceylon, Thailand, 11. dilutus Distant, 1901 Vietnam variety: nesiotus Breddin, 1907 12. exilis S. & W., 1973 China, Vietnam Laos. Pakistan, India, 13. femoralis Distant, 1918 Sumatra, Nepal China, Taiwan 14. harringtonae S.A.W., 1969 Afghanistan 15. hoberlandti S. & A., 1967 **Philippines** 16. lacertosus Bergroth, 1918 India, China, Thailand, Vietnam, 17. lobatus S.A.W., 1969 Sumatra China 18. maai S. & W., 1973 19. maculosus S. & W., 1973 Malava Taiwan, China 20. major Matsumura, 1913 synonym: fossor Bergroth, 1914 21. minor S.A.W., 1969 Vietnam Zambia, Zaire, Senegal 22. nigrolineatus Distant, 1918 23. obnubilus (Distant), 1883 (Ischnodemus) Japan, China, Bonin Islands, Guam, Vietnam **Philippines** 24. philippinensis Distant, 1909 25. pilosus S.A.W., 1969 Laos, Thailand 26. praecerptus Distant, 1904 Burma, Laos 27. privus Distant, 1909 Ceylon, Taiwan, India synonym: abbreviatus Distant, 1918 28. pronotalis Distant, 1910 India, Vietnam Laos, Thailand 29. pseudofemoralis S.A.W., 1969

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31. raja Distant, 1909
32. rufipes Distant, 1911
33. simoni Distant, 1918
34. spinimanus Motschulsky, 1859
synonyms: centralis (Walker), 1872 (Ischnodemus)
           dentipes Motschulsky, 1858 (?)
           leucodermus Breddin, 1907
           singularis Distant, 1909
           sinhalanus Kirkaldy, 1907 (?)
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30. punctatus (Walker), 1872 (Ischnodemus)

35. subauratus Distant, 1904 36. sultanus Distant, 1901 East Africa 37. umbrosus S. & W., 1973 Cevlon 38. uniformis Distant, 1909 India, Laos 39. varipennis (Walker), 1872 (Ischnodemus) synonym: annamita Bergroth, 1894 40. yoshimotoi S.A.W., 1969

India, Malaya, Vietnam, Laos Cambodia, Vietnam, Laos, Thailand

India, Laos, Java, Vietnam, Thai-

Ceylon, India, Vietnam, Thailand,

Vietnam, Thailand

India, Pakistan

Philippines

Cambodia, Laos

land India

MERINADEMUS

1. baraoides Slater, 1967

MICAREDEMUS

1. antennatus Slater, 1967 synonym: decolor Slater, 1967

2. capitatus Slater, 1967

3. coatoni S. & W., 1973

4. congoensis (MS)

5. denticulatus (Slater), 1964 (Ischnodemus)

6. eleganoides Slater, 1967

7. elegans Slater, 1967

8. gillonae (MS)

9. kerzhneri S. & W., 1970

10. lemuriensis Slater, 1967 11. obscurellus Slater, 1968

12. pilosulus (Horvath), 1924 (Ischnodemus)

13. pilosus Slater, 1968

14. pusillus (Dallas), 1852 (Micropus)

15. quadratus S. & W., 1970

16. wilcoxae (MS)

PATRITIODEMUS

1. albomaculatus S. & A., 1969

2. clavatus S. & A., 1969

3. delicatus S. & W., 1968

4. dilutipes (Stål), 1858 (Ischnodemus)

synonym: punctiger (Stål), 1874 (Ischnodemus)

5. minutus S. & A., 1969

6. nigellus S. & A., 1969

7. singularis S. & A., 1969

8. unicoloris S. & A., 1969

PATRITIUS

1. alternatus S. & W., 1966

2. colombianus S. & W., 1966

3. cubensis Barber, 1947

4. fusconervosus (Stål), 1858 (Ischnodemus)

5. grossus (Haglund), 1868 (Papirius)

synonym: velutinus (Walker), 1872 (Ischnodemus)

6. laevus (Stål), 1858 (Ischnodemus)

7. longispadix S. & W., 1966

PIRKIMERUS

1. bellus S. & A., 1965

2. burmanus S. & A., 1965

3. chinai S. & A., 1965

4. esakii Miyamoto & Hidaka, 1960

5. japonicus (Hidaka), 1961 (Ischnomorphus)

synonym: davidi S. & A., 1965

6. javanus S. & A., 1965

7. nicobarensis Distant, 1909

8. ocellatus S. & A., 1965

9. papuensis Slater, 1968

10. parviceps Bergroth, 1918

Madagascar

Madagascar

Madagascar

South Africa

West Central Africa

South Africa, Mozambique

Madagascar

Madagascar

West Africa Madagascar

Madagascar

Madagascar

Madagascar Madagascar

Rhodesia, South Africa, Zaire.

Ruanda, Tanzania, Urundi

Madagascar

East Africa

Brazil Peru

Brazil

Brazil, Argentina

Argentina Peru Brazil

Boliva

Colombia Colombia

Cuba **Brazil**

Argentina, Paraguay, Uruguay, Bra-

Brazil, Argentina

Brazil

India Burma Malaya

Japan

Japan, China, Vietnam

Malaya, Java, Penang Island, Nic-

obar Island Vietnam

New Guinea

Philippines

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 11. philippinensis Slater, 1968 12. qadrii S. & A., 1965 13. sesquipedalis Distant, 1904 	Philippines India Burma, India, Borneo
PRAEBLISSUS 1. albopictus Barber, 1949	Mexico
PRAETORBLISSUS 1. gigas Slater, 1966 2. gradus S. & W., 1968 3. obrieni S. & Ash., 1976 4. wilcoxae S. & Ash., 1976	Peru Peru, Bolivia Costa Rica Peru
PROCELLADEMUS 1. consobrinus S. & W., 1966 2. venenatus S. & W., 1966	Bolivia Peru
PSEUDOBLISSUS new genus 1. trispinosus (Slater), 1967 (Blissus)	Madagascar
RAMADADEMUS 1. anomalous Slater, 1967 2. multispinosus Slater, 1967 3. sakalava Slater, 1967	Madagascar Madagascar Madagascar
RETICULATODEMUS 1. calcar S. & W., 1966 2. nitidus S. & W., 1966 3. orbiculatus S. & W., 1966 4. orbiculoides S. & W., 1968 5. umbrosus S. & W., 1966	Peru Argentina Bolivia, Colombia Brazil Mexico
RIGGIELLA 1. distinctus S. & A., 1965 2. planus S. & A., 1965 3. vianai Kormilev, 1949	Peru Brazil Argentina, Paraguay, Peru
SCANSIDEMUS 1. peregrinus S. & W., 1969 2. taprobanes S. & W., 1969	Burma Ceylon
SCINTILLADEMUS 1. gemmatus Slater, 1968	New Guinea, New Britain
SLATERELLUS 1. hackeri Drake & Davis, 1959	Australia
SPALACOCORIS 1. nigritus S. & A., 1964 2. philippinensis S. & A., 1964 3. rufusculus S. & A., 1964 4. sulcatus (Walker), 1872 (Ischnodemus) 5. sulcifer Stål, 1874	Java, Borneo Philippines, Borneo Sumatra Vietnam, Malaya, Java, Borneo Vietnam, Malaya, Java, Borneo
TALPOBLISSUS 1. cydnoides (Slater), 1964 (Blissus) 2. latus (Distant), 1918 (Blissus)	South Africa, West Africa India
TOONGLASA 1. forficuloides Distant, 1893	Mexico
XENOBLISSUS 1. lutzi Barber, 1954	Brazil

KEY TO GENERA OF BLISSINAE OF THE WORLD

1.	Dorsal surface of pronotum at least partially pruinose (fig. 4A-N)2
la.	Dorsal surface of pronotum completely shining or sub-shining completely lacking pruinose
	areas (except rarely a very narrow pruinose strip along posterior pronotal margin) (fig.
2.	40)
2a.	Fore coxal cavities closed (fig. 6K)6
3.	Anterior half of pronotum shining, posterior
	half pruinose (fig. 67) (Western Hemisphere)
3a.	Pronotum with various combinations of shining
	and pruinose but never with anterior half
	shining and posterior half pruinose (figs. 24, 26)
4.	Membrane completely white; posterior margin
	of pronotum tan, strongly contrasting with
	anterior dark area; body relatively elongate over 4½ mm. (fig. 29)Caveloblissus
4a.	Membrane dark brown with exception of a
⊤ а.	transverse sub-basal pale band; posterior
	pronotal lobe completely dark; body short,
	much less than 4 mm. in length (fig. 67)
5.	Dorsal surface of pronotum completely pru-
	inose; apical corial margin of macropters
	concave (Western Hemisphere) (fig. 24)
5a.	Dorsal surface of pronotum partially shining,
	usually extensively so; apical corial margin
	of macropters straight (fig. 26) (South Af-
,	rica)
6. 6a.	Fore femora mutic
7.	Anterior half of dorsal surface of pronotum
	shining, posterior half pruinose (figs. 27,
	28)
7a.	Pronotum either entirely pruinose, or with pru-
	inose and shining areas but never with ante-
	rior half completely shining and posterior half pruinose (figs. 43-51) Ischnodemus
8.	Membrane composed of numerous small re-
	ticulate cells (fig. 72) Reticulatodemus
8a.	Membrane not composed of reticulate cells .9
9.	Fore femora with 1-2 spines
9a.	Fore femora multispinose, at least 4 spines present
10.	Males with numerous short acute spines on
	abdominal venter and spurlike development
	of seventh connexivum (figs. 8A, 34)
	(Western Hemisphere)Extarademus

10a. Males lacking short acute spines on abdominal venter and without a spurlike projection of 11. Males with genal tusks that project well forward of apex of tylus (fig. 31) (Australian 11a. Males without genal tusks or with very short ones that are not visible from above ...12 12. Middle and hind femora mutic (South Africa) Ischnodemus 12a. Middle and hind femora spinose (Neotropical) 13. Body very broad and flat; pruinosity on dorsal surface of pronotum confined to collar area and a narrow strip across transverse impres-13a. Body relatively slender and elongate; dorsal surface of pronotum predominately pruinose or shining areas occurring as two broad bands, one in area of calli and second across humeri (fig. 61)Patritius 14. Fore coxal cavities open (fig. 6-I) 15 14a. Fore coxal cavities closed (fig. 6K)24 15. Fore femora multispinose (Western Hemi-15a. Fore femora mutic or at most with 1 or 2 spines (Eastern Hemisphere)17 16. Abdominal venter with a prominent stridulitrum on segments 3-6; metathoracic scent gland auricle simple and rounded (figs. 9G, 16a. Abdominal venter lacking a stridulitrum; metathoracic scent gland auricle strongly produced anteriorly (fig. 68) ... Praetorblissus 17. Anterior portion of tylus and juga bearing a series of short sharp thick spines (fig. 78) 17a. Anterior portion of tylus and juga lacking 18a. Fore femora with 1 or rarely 2 ventral spines 19. Scutellum with pruinose areas present laterally and/or anteriorly20 19a. Scutellum completely shining and non-pruinose 20. Macropters with apical corial margin straight; brachypters and wings reduced to minute pads (micropters), at most only covering anterolateral area of first abdominal tergum and widely separated from one another mesally; antennae terete (fig. 26)......

20a. Macropters with apical corial margin concave in part; brachypters with wings variable—if

	short and padlike then relatively broad and		22) (Madagascar)
	nearly in contact mesally; usually with sec-	29a.	Metathoracic scent gland auricle ovoid or ellip
	ond and third antennal segments clavate		tical (fig. 9B-E, G-I) never curving forward
	(figs. 32, 33)Dimorphopterus		as a scimitar-shaped arc; body often elong
21.	Fore tibiae broadly expanded and flattened		ate but not usually so30
	throughout, with a series of sharp spines	30.	Corium and clavus short with lateral area o
	running along almost entire margins of	50.	corium thickened and apical margin mark
			edly concave (fig. 23); usually less than 4
210	shaft (fig. 37)		mm. in length (Ethiopian, Madagascar
21a.	Fore tibiae conventionally terete (figs. 32, 33)		
22		20	
22.	Scutellum completely shining (fig. 54)	30a.	Corium not greatly shortened and withou
			strongly thickened lateral portion; apica
22a.	Scutellum at least in part pruinose (figs. 5-I-L)		corial margin straight; at least 5 mm. in
			length
23.	Macropters with apical corial margin straight;	31.	Scutellum completely shining, non-pruinose
	brachypters with wings reduced to minute		pronotal hairs elongate, upstanding and lat
	pads (micropters), at most only covering		erally directed anteriorly (Neotropical) (figs
	anterolateral area of first abdominal tergum		43-51)
	and widely separated from one another	31a.	Scutellum at least pruinose laterally; pronota
	mesally; antennae tereteCapodemus		hairs short, usually decumbent, sometimes
23a.	Macropters with apical corial margin concave		almost glabrous, never elongate or directed
	at least in part; brachypters with wings vari-		anteriorly32
	able—if short and padlike then relatively	32	Scutellum completely pruinose (Neotropical
	broad and nearly in contact mesally, usually	J	figs. 43-51)
	with second and third antennal segments	322	Scutellum pruinose only laterally
			Six mm. or more in length; labium extending
24	clavate	33.	
			well into mesosternum (Neotropical) (figs
	Fore femora spinose	22-	43-51)
25.	Hind femora spinose (figs. 64, 65) (Oriental)	33a.	Less than 4 mm. in length; labium extending
	Pirkimerus		only between forecoxae (South African
	Hind femora mutic	2.4	(fig. 20)
26.	Ovipositor produced conspicuously caudad	34.	Fore femora with only a large bifid spine ven
	(fig. 8H, I) (Australian)27		trally near distal ends and a terminal spine
26a.	Ovipositor not conspicuously produced caudad		distad (fig. 19); fourth antennal segmen
	(fig. 8C-G)28		markedly petiolate (fig. 19)
27.	Third antennal segment noticeably narrower	34a.	Fore femora either multispinose or with one to
	than segment 2 (fig. 38); forewing even in		four distinctly separated spines; fourth an
	brachypters extending over two or three ab-		tennal segment usually not markedly petiol
	dominal segments; lateral pronotal margins		ate
	not sharply carinate; ninth paratergite of fe-	35.	Bucculae extended far forward of apex of tylus
	male not divided into two distinct sclerites		as broadened flaplike lobes (fig. 18
	(fig. 8H)		(Madagascar)
27a.	Second and third antennal segments subequal	35a.	Bucculae not strongly produced forward o
	in thickness (fig. 21); forewing reduced to a		apex of tylus
	minute pad not extending posteriorly onto	36.	Scutellum with pruinose areas present laterally
	abdominal terga; lateral pronotal margins		prosternum pruinose before and between
	sharply carinate; ninth paratergite of female		coxae; membrane of fore wing thickened
	divided (fig. 8-I) Australodemus		and opaque (fig. 75) (New Guinea)
28	Membrane with veins extensively anastomos-		Scintillademus
~ 0.	ing (fig. 76) (Australian)Slaterellus	360	Scutellum and prosternum both completely
28a	Membrane veins not extensively anastomosing	Jua.	shining and non-pruinose; membrane o
_0a.	29		forewing thin and translucent (fig. 19
20	Metathoracic scent gland auricle curving for-		(Madagascar)
<i>∠</i> ヲ.	ward in a scimitar-shaped arc (fig. 9Q),	27	
	body extremely elongate and slender (fig.	31.	Apex of abdomen with a pair of large elongate
	oody extremely clongate and stender (fig.		projections present (fig. 79) (Neotropical

	Toonglasa		lacking a median longitudinal groove on
37a.	Apex of abdomen lacking a pair of elongate		anterior lobe (fig. 53); labium reaching well
	projections		onto mesosternumLucerocoris
38.	Fore femora multispinose (at least three or four	47.	Hind tibia not appreciably longer than first
	spines present)		metatarsal segment (fig. 35) (Oriental)
38a.	Fore femora with only one or two spines pres-		Extaramorphus
	ent51	47a.	Hind tibia much longer than first metatarsal
39.	All femora multispinose40		segment48
39a.	Middle and hind femora mutic	48.	Metathoracic scent gland auricle greatly en-
40.	Male hind femora much larger and thicker than		larged at distal end (figs. 12A, 69) (Neo-
	fore femora (fig. 25)		tropical)
40a.	Male hind femora not larger and thicker than	48a.	Metathoracic scent gland auricle variable in
	fore femora41		shape but not strongly enlarged at distal end
41.	Metathoracic scent gland auricle strongly curv-		(Eastern Hemisphere)49
	ing anteriorly in a crescent-like arc (fig.	49.	Apical corial margin deeply concave (fig. 70)
	12C) (Madagascar (fig. 71)		Pseudoblissus
		49a.	Apical corial margin straight50
41a.	Metathoracic scent gland auricle either curving		Membrane of nearly uniform texture
	slightly posteriorly, or straight, with a		throughout (figs. 55-58)
	broadened and slightly anteriorly produced	50a.	Membrane of forewing with basal area dull
	distal end42		pruinose, strongly contrasting with shining
42.	Body only moderately flattened; metathoracic		center portion of membrane (fig. 41)
	scent gland auricle not enlarged strongly at		
	distal end but curving slightly posteriorly	51.	Males with prominent jugal extensions visible
	(figs. 7K, 61)		from above (figs. 6E-H, 40) (Oriental, Aus-
42a.	Body greatly broadened and flattened; meta-		tralian)
	thoracic scent gland auricle nearly straight	51a.	Males without jugal extensions52
	for most of length but expanded and some-		Hind femora spinose (figs. 64, 65) (Oriental)
	what produced anteriorly at distal end (fig.		Pirkimerus
	12B)	52a.	Hind femora mutic
43.	Scutellum pruinose laterally; pronotum com-		Apical corial margin concave54
	pletely pruinose laterally and ventrally and		Apical corial margin straight57
	with a narrow pruinose strip dorsally across		Body elongate, parallel sided, length six times
	collar, ventral surface of head with short		maximum width (fig. 59) (Madagascar)
	acute projections present, mesosternum		
	lacking a median longitudinal furrow (fig.	54a.	Body robust, elliptical, not parallel sided,
	73) (Neotropical)		length much less than six times maximum
43a.	Scutellum and pronotum completely shining,		width55
	non-pruinose; no projections present on un-	55.	Fore femora with two ventral spines; apical
	derside of head, mesosternum with a deep		corial margin concave only along inner one-
	median longitudinal furrow (fig. 74) (Orien-		third (fig. 80) (Neotropical) Xenoblissus
	tal)	55a.	Fore femora with a single ventral spine; apical
44.	Fore tibiae fossorial, enlarged and strongly		corial margin deeply concave for entire
	toothed (fig. 30) (Oriental)45		length (Madagascar)
44a.	Fore tibiae non-fossorial, chiefly terete, some-	56.	Eyes laterally produced; abdomen ovoid; mem-
	times enlarged and swollen at distal ends.		branes of forewings only partially overlap-
	47		ping (fig. 36)
45.	Lateral half of corium not shining (at most	56a.	Eyes not laterally produced; abdomen ellipti-
	only area of radial vein shining) (fig. 30)		cal; membranes of forewings completely
			overlapping (fig. 52)Lemuriblissus
45a.	Lateral half of corium shining (fig. 77)46	57.	Metathoracic scent gland auricle lunately curv-
	Pronotum subcylindrical with a deep longitudi-		ing anteriorly (fig. 9Q) (Africa,
	nal median groove on anterior pronotal lobe		Madagascar)
	(fig. 77); labium not attaining mesosternum	57a.	Metathoracic scent gland auricle lobate,
	Spalacocoris		rounded (fig. 10A)58
46a.	Pronotum sub-flattened, not subcylindrical,	58.	Prothorax with pruinosity present on sternum

WORLD KEY TO KNOWN FIFTH INSTAR NYMPHS OF THE BLISSINAE

The following key is based upon 24 genera and approximately 102 species. This key must be approached in a very different manner than that used when one is working with keys to the adults. Kevs to adults include all the known genera or species, and if they are accurately prepared erroneous results should be obtained only when an undescribed species is involved, where the limits of variation are greater than was known at the time the key was prepared or where an inadvertent error was made in the construction of the key. This is not true of the key to fifth instar nymphs presented below. A great many genera and species of the Blissinae already "exist" for which no nymphs have been available for study. The key, therefore, is to be viewed more as a short hand guide to place an unknown nymph in some position relative to those already known, to indicate many things that it cannot be and, perhaps most importantly, to point up the kinds of characters and character states that appear to be most useful in the segregation of nymphs.

It is already evident that the morphology of the immature stages will ultimately be extremely valuable to phylogenetic analysis of the subfamily.

Slater and Wilcox (1973) have developed a letter coding for the various sclerotized areas of the abdomen. This system (see fig. 7N, O) has been used throughout the following key.

1.	TM 2 present
la.	TM 2 absent
2.	TM 3 present (sometimes reduced to a minute
	mesal spot)3
2a.	TM 3 absent
3.	TM 4 present and separate from SGA 4-5
3a.	TM 4 absent, or if present fused with SGA
	4-5
4.	Mesothoracic wing pads sordid yellow
	Blissiella castanea
4a.	At least distal third of wing pads dark brown
5.	Fore femora mutic, abdomen a unicolorous

	sordid yellow; TM 3 ovoid, well devel-
5.0	oped; TM 6 absent Blissiella micans Fore femora with one or two small sharp
Ja.	
	ventral spines on distal one-third; abdo-
	men variegated with red-brown and yel-
	lowish; TM 3 reduced to a minute mesal
	spot, TM 6 present as a transverse sclerite
	covering most of posterior half of tergum
	Ischnodemus sinuatus
О.	TM 4 present (sometimes reduced to a very small spot)
6a.	TM 4 absent or fused with SGA 4-5 8
7.	
٠.	formly dark red; SM 4 absent
_	Iphicrates nigritus
7a.	Abdomen a variegated red, yellow, and
	brown; SM 4 present, large and conspic-
	uousScintillademus gemmatus
8.	TM 5 present, distinctly differentiated9
8a.	TM 5 absent or fused with SGA13
9.	Fore femora mutic
7.	
Q ₂	Fore femora armed below with two or more
Ja.	spines
10	
10.	
	spines
10a.	Fore femora multispinose12
11.	
	nent black spots and dashes; TM 6 a nar-
	row transversely elongate bar
11a.	TPC row absent; TM 6 trianguloid
12.	
	not produced anteriorly; head, pronotum,
	scutellum and mesothoracic wing pads a
	uniformly sordid yellow or yellowish
	brown; legs yellow Iphicrates spathus
12a.	
	distinctly produced antero-laterad into a
	blunt point; head, pronotum, scutellum,
	and proximal one-third of mesothoracie
	and proximal one-third of mesothoracic wing pads dark red-brown, remainder of
	wing pads dark red-brown, remainder of
	wing pads shading to a contrasting bright
	yellowish brown; legs chiefly dark brown
	Iphicrates papuensis
13.	SM 4 present, distinct
13a.	SM 4 absent
14.	SM 5 present and distinct
14a.	SM 5 absent (sometimes a few small dots
	present) ¹
	p. 200m()

¹This character is somewhat variable. With only a small series it may be desirable to "run" both ways at this couplet.

15.	Fore femora armed below with only one to	26.	Abdomen with posteriorly directed spines
15a.	three short sharp spines		present laterally on terga 8 and 9
16.	TL 6 present	26a.	Abdomen lacking spines on terga 8 and 9
	Fore femora armed below with three short sharp apines; antennal segments 2 and 3	27.	Each fore femur armed below with a single spine
	brown	27a.	Each fore femur with two or more spines
17a.	Fore femora armed with two spines (rarely one, sometimes a "hair" spine) (if vestige	28.	present
	of third spine present then antennal seg- ments 2 and 3 yellow, not brown)18		nent contrasting white macula in mesal area
18.	Head, pronotum, scutellum, and wing pads	28a.	Wing pads completely dark brown, or with
	light yellowish brown		mesal area yellowish brown but without a contrasting white macula present29
18a.	Head, pronotum, scutellum, and wing pads	29.	Abdomen variegated with red and yellow;
19.	dark brown to black		TM 7 remote from anterior margin of tergum 7, TML 7 present and distinct;
10-	ish white Micaredemus pusillus		TPC row absent or reduced to minute spots
19a.	Abdomen with red, yellow, and tan markings, or with terga 2 and 3 tan and 4	29a.	Abdomen uniformly dark red; TM 7 covering
20	through 7 bright yellow		entire tergum, TML 7 absent or fused; TPC row present with spots prominent
20.	Antennal segments 2 and 3 yellow, strongly contrasting with dark brown fourth seg-		
	ment; abdominal segments 3-7 with red and yellow markings; labium relatively	30.	SGA sclerites greatly enlarged into a prominent ovoid or quadrate patch, dark and
	elongate, reaching between fore coxae,		strongly contrasting with remainder of ter-
	second segment attaining base of head (Madagascar Morojejy, 14°26′ S, 49°44′	30a	gal surface
	E)	30 a .	greatly enlarged into a prominent partch
20a.	Antennal segments 2 and 3 red-brown, not strongly contrasting with dark brown		(if appearing enlarged then nearly uni- colorous with remainder of tergal surface)
	fourth segment; abdomen yellow and tan,		37
	lacking red markings; labium shorter, not	31.	All femora multispinose; posterior TML spots
	reaching anterior margin of fore coxae, second segment remote from base of head		greatly enlarged, as large as or larger than SGA scleritesRamadademus sakalava
	(Madagascar Andranomalaza, 25°12'S,	31a.	At most only fore femora spinose; TML spots
21.	46°52′E)		not greatly enlarged, smaller than SGA sclerites32
21a.	TL 6 absent	32.	Fore femora multispinose
22.	TL 6 present (sometimes reduced to a few small dots)	320	Fore femora mutic or with only a single spine
22a.	TL 6 absent	32 a .	present33
23.	Fore femora mutic	33.	Fore femora mutic
23a.	Each fore femora armed below with one or	22.	
24.	more spines	33a.	Fore femora with a single ventral spine present
	acoxae	34.	TM 6 present as a very narrow transverse
	Labium shorter, not reaching metacoxae25		dash; tergum 3 reddish brown with a
23.	TM 6 absent, mesothoracic wing pads relatively small, not attaining posterior margin		strongly contrasting yellow patch on each anterolateral area
	of metanotum Ischnodemus proprius		
25a.	TM 6 present, distinct; wing pads conven-	34a.	TM 6 absent; tergum 3 nearly uniformly yel-
	tional, reaching over anterior margin of	25	lowish, tan or red-brown
	tergum 3	<i>5</i> 5.	Labium relatively short, remote from fore

	coxae, third segment barely attaining base	45.	Abdomen uniformly dull yellow
	of head; tergum 6 with contrasting red and		Atrademus maritimus
	yellow coloration	45a.	Abdomen with terga 1 and 2 white, contrast
	Extarademus humerus		ing with dull brown color of terga
35a.	Labium longer, reaching at least between fore		through 7 Ischnodemus asciaformis
	coxae; tergum 6 nearly uniformly deep	46.	Anterior margin of TM 7 and SG sclerites
	pink, creamy yellow or tan36		broadly margined with black which con-
36.	Labium relatively elongate, one-half of fourth		trasts strongly with dark yellowish brown
	segment reaching onto mesosternum;		color of remainder of sclerites; color deep
	tergum 6 deep pink		orange
	Extarademus mundus	46a.	TM and SG sclerites either uniformly colored
36a.	Labium shorter, at most attaining anterior		throughout or with only very narrow in
	margin of mesosternum; tergum 6 creamy		complete traces of black margins; color
	yellow or tanExtarademus macer		light yellow or pink
37.	TM 7 absent	47.	SG 5-6 sclerotization elliptical, nearly twice
37a.	TM 7 present (sometimes nearly unicolorous		as wide as long (longitudinally)
	with remainder of tergum; in some species		Capodemus sabulosus
	reduced to a narrow transverse dash on	47a.	SG 5-6 sclerotization rectangular, much less
	either side of midline near posterior mar-		than one and three-fourths times as wide
	gin)		as long
38.	TML row present, spots prominent	48	TML 7 and TMA 7 connected to form a
	Extarademus macer		completely sclerotized mesal area ante-
38a.	TML row absent39		riorly on tergum; SM 7 very broadly trun-
	Fore femora armed below with two small and		cate at anterior end
	one large tuberculate spines on distal one-		
	third, abdomen chiefly sordid yellow; usu-	48a	TML 7 and TMA 7 not connected, forming
	ally micropterus, wing pads not extending	Tou.	three distinctly separated sclerites ante-
	beyond metanotum		riorly on tergum; SM 7 oval, rounded, no
			broadly truncate at anterior end
39a	Fore femora with a single small spine below;		Capodemus pentameri
<i>-</i> ,	abdomen creamy white; macropterus,	40	TM 7 divided into two separate patches, one
	wing pads extending over anterior half of	77.	on either side of midline (sometimes nar-
	abdominal tergum 3 Heinsius pallidus		rowly fused posteriorly) 50
40	TMA 7 present as a distinct (sometimes very	40a	TM 7 not completely divided into two sepa-
	small) spot (or if appearing obsolete then	₹7 a.	rate sclerites
	abdominal terga 3-7 uniformly pink and	50	Fore tibiae greatly enlarged with 4-5 heavy
	antennal segment 4 three and a half times	50.	chines at distal and and a new of stary
			spines at distal end and a row of stour
40a	as long as segment 3)	50a	spines along shaft
	TMA 7 absent or fused	30a.	Fore tibiae not greatly enlarged and spinose
71.		51	Small analisa and an America for form
	robust, nearly three and a half times	31.	Small species, under 4 mm.; fore femora mu-
	length of segment 3	£1.	tic
/1a	Fourth entennel segment shorter rever more	ora.	Larger species, over 7 mm.; fore femora mul-
+1a.	Fourth antennal segment shorter, never more	50	tispinose Lucerocoris nigrotibialis
	than three times as long as segment 3	32.	Labium relatively elongate, attaining or ex-
42	Fore femora with a single very small sharp	50.	ceeding mesocoxae
44.			Labium shorter, not attaining mesocoxae .54
	ventral spine present		TML row present Dimorphopterus cornipes
12.			TML row absent Blissus leucopterus
	Fore femora mutic	54.	Body unicolorous, sordid yellow to white
	SM 6 absent		(except often a diffuse red in scent gland
	SM 6 present and well developed	E 4 -	area)
	TM 7 mesally divided into two separate		Abdomen red or light brown55
140	patches Ischnodemus sabuleti	33.	Wing pads sordid yellow; abdomen red
₩ā.	TM 7 a single undivided plate		Ischnodemus parathoracicus

55a.	Wing pads dark brown; abdomen light brown56	65a.	Abdomen variously colored, often variegated and banded, never uniformly pale yellow
56.	Fore femora with a single sharp ventral spine present Dimorphopterus brachypterus	66.	Pronotum uniformly dark chocolate brown to
56a.	Fore femora mutic		black Atrademus capeneri
57.	SM 6 broad, trianguloid, posterior margin in	66a.	Pronotum bicolored with anterior two-thirds
	contact with SM 7		dark brown to black, posterior one-third a
			strongly contrasting testaceous to white
57a.	SM 6 smaller, irregularly ovoid, remote from		
	posterior margin of sternum and not in	67.	Pronotum uniformly dark brown to black
	contact with SM 7		Macchiademus diplopterus
	Ischnodemus nigrocephalus	67a.	Posterior one-third of pronotum white,
58.	Abdomen with at least mesal area of terga 1		strongly contrasting with dark anterior
	and 2 opaque white or yellowish and		area (see Slater and Wilcox 1973, pp.
	strongly contrasting with darker mesal		100, 111-112) Macchiademus sp. I
	area of tergum 359		TL 7 absent
58a.	Abdomen with at least tergum 2 (although		TL 7 present (somtimes reduced)70
	sometimes light colored) not strongly con-	69.	Pronotum and scutellum dark chocolate
	trasting with tergum 3		brown
59.	TM 6 present (in some specimens reduced to	69a.	Pronotum and scutellum honey yellow to
	a few small spots)	70	light brown (Arizona)
	TM 6 absent	70.	TML spots large and conspicuous, especially posteriorly71
	Fore femora mutic	700	TML spots absent or very small and in-
ooa.	Fore femora armed below with 1 or more short sharp spines	70a.	conspicuous, or nearly unicolorous with
61	TM 7 large, quadrate, uniformly black 62		remainder of dorsal surface72
	TM 7 smaller, distinctly narrowed anteriorly,	71	Fore femora mutic
oru.	reddish or yellowish brown, sometimes	,	Dimorphopterus blissoides
	laterally bordered with black 63	71a.	Fore femora with a prominent sharp ventral
62.	Antennal segments 1-3 a unicolorous sordid		spine present Dimorphopterus zuluensis
	yellow; SM 6 present as a median triangu-	72.	Labium relatively elongate, at least attaining
	lar sclerite Ischnodemus fulvipes		posterior margin of mesocoxae73
62a.	Antennal segment 3 (and 4) nearly black,	72a.	Labium shorter, not attaining anterior margin
	strongly contrasting with sordid yellow		of mesocoxae74
	coloration of segments 1 and 2; SM 6	73.	Fore femora armed below with a single short
	absent		stout spine Dimorphopterus pilosus
63.	Antennae uniformly dark red-brown to black;		Fore femora mutic
	wing pads dark red-brown; TM 7 reddish	/4.	Fore coxal cavities open (fig. 6-I)
62.	brown	740	
оза.	Antennae with segments 1 and 2 sordid	/4a.	Fore coxal cavities closed (fig. 6K)
	yellow, 3 and 4 becoming red-brown;	75	
	wing pads honey yellow; TM 7 yellowish brown	13.	Antennae uniformly reddish brown
64	TM 7 distinctly bordered with black; terga	75a	Antennae with segment 1-3 yellowish brown,
01.	3-5 with yellow areas anterolaterally that	754.	segment 4 strongly contrasting red brown
	strongly contrast with mesal and posterior		(or if segments 1-3 darker brown, labium
	red areas		barely attaining metacoxae, not reaching
64a.	TM 7 not distinctly bordered with black;		onto abdomen)
	terga 3-5 nearly uniformly red with at	76.	TL 7 absent, fused or reduced to tiny dot
	most diffuse indistinct light areas anter-		
	olaterally not strongly contrasting with re-	76a.	TL 7 present, distinct and well developed
	mainder of tergum		
(=		77.	TM 6 absent
05.	Abdomen a nearly unicolorous pale yellow.	10	a according to the second of DE
		'Se	e generic discussion of Blissus.

	TM 6 present		TM 6 present and prominent (See Slater and Wilcox 1973, pp. 69-70)
	ovoid mesal patch, nearly unicolorous with remainder of sternum (S. Dakota)	87a.	Fore femora armed below with a single spine; TM 6 absent Dimorphopterus littoralis
78a.	SM 6 small but distinct, light brown, contrasting with yellow sternum, transversely rectanguloid (Nicaragua) Blissus sp.	88.	Fore femora multispinose, spines stout, of various lengths and scattered, often curved
79.	Each fore femur armed below with a large bifid spine	88a.	Fore femora mutic or with 1-2 spines or a bifid spine present; if occasionally 3-5
	Fore femora mutic		spines present, these slender, straight, of nearly equal length and placed in a straight row94
	(except diffuse areas where scent glands are visible)		TPC row absent
80a.	Abdomen not unicolorous, variously marked with red, yellow, or orange81	90	Abdomen nearly uniformly reddish orange,
81.	Mesothoracic wing pads uniformly dark chocolate brown Ischnodemus conicus	<i>7</i> 0.	sclerites only slightly darkened, not strongly contrasting with remainder of
81a.	Mesothoracic wing pads with at least distal two-thirds sordid yellow to testaceous	90a	tergum
01	82	να.	rites distinctly darkened and contrasting with tergal surface91
02.	Large species, 6 mm. in length; abdominal terga 3-5 tinged with red along posterior	91.	TM 7, TMA 7 and TML 7 fused into a single
82a.	margins Ischnodemus badius Smaller species, less than 3.5 mm. in length;	91a.	sclerite
	abdominal terga 3-5 yellow and white, not tinged with red along posterior margins,	02	meson but distinctly separated from ante- rior margin of TM 7
	red only mesally where scent glands are visible		Large species, over 6 mm. in length; all femora multispinose Patritius laevus
83.	Abdomen dark red to red-brown on at least terga 3-7, lacking light markings (although	92a.	Smaller species, under 5.5 mm. in length; only fore femora multispinose93
	sometimes becoming lighter along lateral margins)	93.	TML 7 and TMA 7 fused across meson but distinctly separate from sinuate anterior
83a.	Abdomen variously colored with at least some light markings present on terga 3		margin of TM 7
	and 4, or terga 3-7 unicolorous reddish orange, yellowish or gray without light	93a.	TML 7, TMA 7 and TM 7 fused into a single sclerite with a straight anterior margin
01	markings	04	
	Fore femora mutic		TPC row absent95 TPC row present, spots sometimes very small
o .u.		<i>></i> 1 u .	99
85.	Abdominal terga 1 and 2 light yellow mesally, strongly contrasting with dark red coloration of remainder of tergum	95.	Abdomen chiefly light brown with contrasting white areas mesally on tergum 1 and a longitudinal patch on preconnexival area
85 0			of terga 4 and 5; TM 7 nearly in contact
osa.	Only abdominal tergum 1 light yellow mesally, tergum 2 uniformly red-brown or red		with posterior margin of tergum 6
86	and white	95a.	Abdomen white, testaceous, or unicolorous
	coxae		tan, lacking contrasting white areas; TM 7 small, remote from posterior margin of
ooa.	Labium much shorter, at most attaining posterior margin of prosternum	96.	tergum 6
87.	Fore femora armed below with two spines;		sordid yellow; antennal segments 1 and 2 testaceous, much lighter than brown seg-

	ments 3 and 4 (some populations of capeneri have the head, pronotum, scutellum, and wing pads dark brown to black, but antennal segments 1 and 2 are light testaceous)Atrademus capeneri		black; SM 6 large, broadly in contact with both posterior and anterior margins of sternum; TML spots large and conspicuous on all terga, much larger than TPC spots
96a.	Head, pronotum, scutellum and wing pads dark brown; antennae unicolorous brown	106a.	Head and pronotum reddish brown; SM 6 smaller, not attaining anterior or posterior margins of sternum; TML spots small and
97.	Fore femora armed below with 1 or 2 spines		inconspicuous, only slightly larger than TPC spots
97a.	Fore femora mutic98	107.	Abdomen bright yellow; wing pads honey
98.	Larger species, length 4.80 mm. or more;		yellow Ischnodemus brevicornis
	TML row present as small but distinct spots on tega 4-6	107a.	Abdomen gray; wing pads dark brown
	Dimorphopterus gibbus	108.	Fore femora with 1 or 2 distinct sharp ventral
98a.	Smaller species, total length not over 2.76;		spines present109
	TML row absentBlissus mixtus	108a.	Fore femora mutic110
99.	TM 6 present100	109.	Abdominal terga 3-7 nearly uniformly tan;
99a.	TM 6 absent, or at most represented by a few		wing pads dark brown
	small dots108		
	Fore femora mutic	109a.	Abdomen banded red and yellow; wing pads
	Fore femora armed below with one or more spines		honey yellow (Natal, S. Africa)sp.
101.	TM 7 uniformly dark brown	110.	Abdomen nearly unicolorous reddish, yellow- ish or tan, lacking contrasting light or
101a.	TM 7 red-brown, bordered laterally with		dark patches111
	black	110a.	Abdomen with at least terga 3 and 4 marked
102.	Dorsal preconnexival area, at least on terga		with contrasting red or yellow patches 113
	3-5, with anterior half yellow, posterior	111.	TMA 7 and TML 7 separate, and not fused
	half contrasting red or orange		with TM 7; antennae unicolorous dark
100			brownDimorphopterus gibbus
102a.	Preconncexival area uniformly dull yellowish	111a.	TMA 7 and TML 7 fused at least across
	or reddish, sometimes with a narrow lon-		meson and also fused with TM 7 either
	gitudinal stripe present near lateral mar-		mesally or completely; first three antennal segments yellow to light brown112
103	gins	112	SM 6 present, large trianguloid; TMA 7,
103.		112.	TML 7, and TM 7 fused into a single
103a	Abdomen nearly uniformly gray, yellow or		large sclerite Ischnodemus proprius
1054.	brownish yellow104	112a.	SM 6 absent; TMA 7 and TML 7 fused
104.	Fore femora with three ventral spines present;		across meson but connected with TM 7
	labium relatively elongate, reaching well		only mesally, distinctly separate laterally
	onto mesosternum, third segment surpass-		(Machadodorp, TVL, South Africa)
	ing fore coxae; abdomen brownish yellow		
		113.	Head, pronotum, scutellum, and wing pads
104a.	Fore femora with a single spine below; la-		honey yellow; TMA 7 present and sepa-
	bium relatively short, at most barely ex-		rate from TM 7 and TML 7; TML and
105	ceeding fore coxae		TPC rows very large and conspicuous
105.	Pronotum with posterior one-fourth pale	112-	
	yellow, strongly contrasting with dark	113a.	Head, pronotum, scutellum and basal third of
	brown remainder of surface (See Slater and Wilcox 1973, pp. 100, 112)		wing pads dark brown to nearly black,
			remainder of wing pads becoming lighter brown; TMA 7 absent or fused with TM 7
105a	Pronotum unicolorous light or dark brown		or TML 7; TML and TPC rows conven-
	······106		tional, not large and conspicuous
106.	Head and pronotum very dark brown to		
	i start to		an invacing anapes

GENERA AND NUMBER OF SPECIES OF EACH PRESENT IN KEY TO FIFTH INSTAR NYMPHS

Atrademus 2 of 4 Barademus 1 of 1 Blissiella 2 of 5 Blissus 6+ of 24 Capodemus 8 of 15 Cavelerius 1 of 10 Dentisblissus 1 of 4 Dimorphopterus 11 of 35 Extarademus 4 of 8 Geoblissus 1 of 8 Heinsius 1 of 2 Heteroblissus 1 of 1 Inhicrates 4 of 15 Ischnocoridea 1 of 1 Ischnodemus 27 of 94 Lucerocoris 1 of 2 Macchiademus 4 of 5 Macropes 14 of 40 Micaredemus 5 of 16 Patritiodemus 1 of 8 Patritius 2 of 7 Praeblissus 1 of 1 Ramadademus 1 of 3 Scintillademus 1 of 1

Genera with nymphs known—24 Genera with nymphs unknown—22

DIAGNOSES OF GENERA

ARADACRATES SLATER AND WILCOX Figure 18

Aradacrates Slater and Wilcox, 1968, p. 439.

Type Species: Aradacrates cochlear Slater and Wilcox. Monobasic.

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, slender, linear. Metathoracic scent gland auricle elongate, narrow, slightly curving anteriorly, enlarged toward distal end. Fore femora short, incrassate, armed below on distal third with a very large bifid spine; middle and hind femora bearing several short, sharp spines. Tibiae very short, clavate, with several teeth present along shaft, hind tibiae bearing several small teeth along lateral margins. First tarsal segment of fore leg elongate, swollen, bearing a mat of

soft hairs below. Entire body shining, no pruinose areas present. Apical corial margin slightly concave along inner portion. Membrane thin, hyaline, differentiated from clavus and corium, latter thickened along radial vein. Fore coxal cavities closed. Abdominal segments two through five fused without visible sutures. Ocelli small. Antennae with second and third segments clavate. Bucculae extending forward of tylus as large expanded semielliptical dorsally excavated flanged plates. Claspers somewhat scimitar shaped, inner projection absent, outer projection small and trianguloid. Sperm reservoir greatly reduced, wings absent, central cuplike area represented only by a small protruding scalelike projection arising from ejaculatory duct. Ovipositor unknown.

ARADADEMUS SLATER Figure 19

Aradademus Slater, 1967, p. 5

Type Species: Aradademus mirificus Slater. By original designation.

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

DIAGNOSIS: Elongate, robustly linear. Metathoracic scent gland auricle elongate, strongly angled anteriorly to form an "L"-shaped structure (fig. 90) or elongate, straight and enlarged at distal end (fig. 9M). Fore femora armed below with a large bifid spine. Fore tibiae short, swollen. Entire body completely shining and non-pruinose. Apical corial margin strongly concave. Membrane much thinner than corium. Fore coxal cavities closed. Ocelli small. Antenniferous tubercles large and curved. Fourth antennal segment with a narrow nearly pediculate proximal end. Sperm reservoir reduced to a tapering scalelike median projection and a pair of minute elliptical wings far removed from central projection. Spermatheca with very short thick pump (fig. 2C).

KEY TO SPECIES OF ARADADEMUS

- la. Eyes large, occupying greater part of lateral

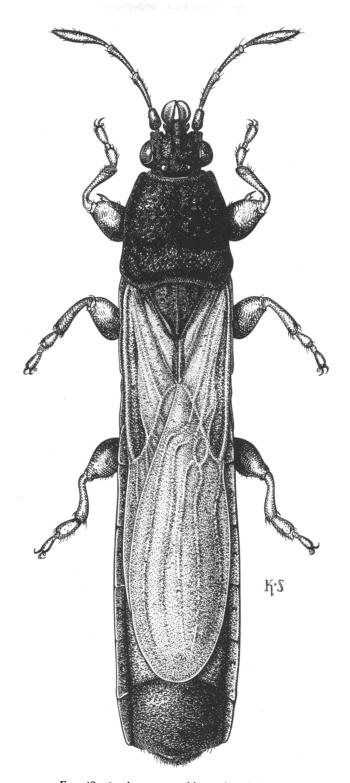


Fig. 18. Aradacrates cochlear, dorsal view.

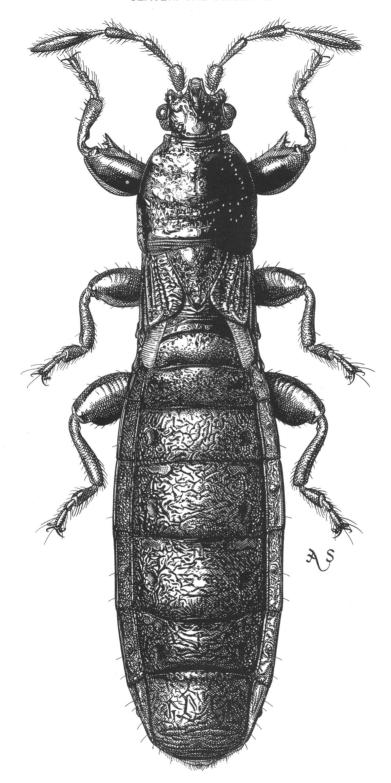


Fig. 19. Aradademus mirificus, dorsal view.

ATRADEMUS SLATER Figure 20

Atrademus Slater, 1967, p. 45.

Type Species: *Ischnodemus capeneri* Slater. By original designation.

DISTRIBUTION: Africa, Madagascar.

BIOLOGY: Species known to breed on several genera of Gramineae.

DIAGNOSIS: Elongate, slender, linear; metathoracic scent gland auricle short, rounded, earlike; fore femora usually with one or two spines, occasionally mutic; head and thorax completely shining above, below almost completely shining at most with a small pruinose area on venter before anterior coxae; scutellum with pruinosity usually present basally and sometimes laterally; apical corial margins of macropters straight; brachypters and micropters common; membrane translucent, much thinner than corium; fore coxal cavities closed; ocelli small; antennae terete or slightly clavate; sperm reservoir cup large, wings elongate, slender, straplike; ovipositor elongate. Spermatheca with pump very elongate, curved and swollen distally (fig. 2R).

KEY TO SPECIES OF ATRADEMUS

- 1. Fore femora mutic; labium relatively short, reaching posteriorly only between fore coxae (South Africa) (fig. 20).....
- la. Fore femur with one or more ventral spines, or if mutic then labium longer, extending at least to middle of mesosternum2

- 3a. Labium shorter, not or barely reaching posterior margin of mesosternum; usually brachypter-

AUSTRALODEMUS SLATER AND SWEET Figure 21

Australodemus Slater and Sweet, 1963, p. 53.

Type Species: Australodemus elongatus Slater and Sweet. Monobasic.

DISTRIBUTION: Australia.

BIOLOGY: Unknown.

DIAGNOSIS: Body very narrow and elongate, not flattened; metathoracic scent gland auricle small, rounded, earlike; fore femora with a single small spine present; head and pronotum of a somewhat dull appearance but without any indication of pruinosity either above or below, surface texture usually somewhat masked by presence of numerous flattened scalelike hairs: only micropters known; fore coxal cavities closed; ocelli small; antennae terete but very thick and stout; sperm reservoir reduced to a small median, distally rounded protrusion (fig. IEE, FF). Ovipositor elongate, paratergites eight and nine strongly produced caudad of segment eight (fig. 8-I). Spermatheca with a simple non-flanged bulb and a short narrow pump.

BARADEMUS SLATER Figure 22

Barademus Slater, 1967, p. 13.

Type Species: *Barademus attenuatus* Slater. Original designation.

DISTRIBUTION: Southern Africa, Madagascar.

BIOLOGY: Known to breed on *Ischaemun arcuatum* (Nees) Stapf. (Panicoideae, Andropogoneae).

DIAGNOSIS: Body extremely elongate, slender, parallel sided, non-flattened; metathoracic scent gland auricle scimitar-shaped (fig. 9Q) curving forward; all femora mutic; head and pronotum above and scutellum completely shining; no trace of pruinosity laterally or ventrally, even corium and clavus completely shining; apical corial margins slightly concave; membrane hyaline, strongly differenti-

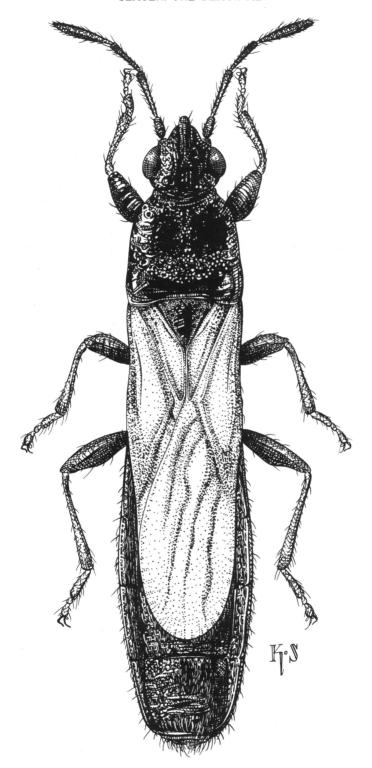


Fig. 20. Atrademus maritimus, dorsal view.

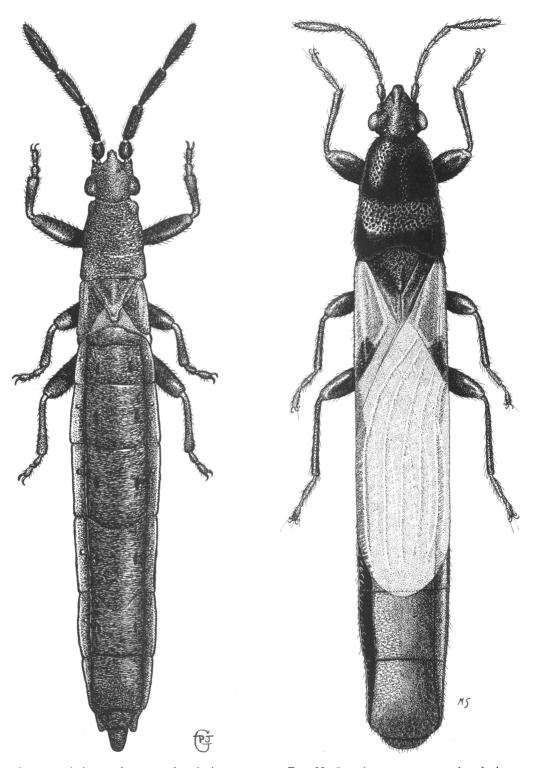


Fig. 21. Australodemus elongatus, dorsal view.

Fig. 22. Barademus attenuatus, dorsal view.

ated from corium; wing polymorphism present; fore coxal cavities closed; ocelli small; antennae terete; sperm reservoir greatly reduced, bulb minute, lacking wings; ovipositor elongate. Spermatheca with short simple pump (fig. 2G).

BLISSIELLA SLATER Figure 23

Blissiella Slater, 1967, pp. 17, 19.

Type Species: Ischnodemus castaneus Slater. Original designation.

DISTRIBUTION: Africa, Madagascar.

BIOLOGY: Known to breed on several genera of Gramineae.

DIAGNOSIS: Body thick, rather robust, elongate or stout, not flattened; scent gland auricle very small, rounded, button-like or earlike; all femora mutic; head, pronotum and scutellum completely shining: no pruinosity below, even corium and clavus completely shining; apical corial margin concave; corium considerably shortened even when an elongate membrane is present, thickened distally and heavily so along radial vein; brachyptery frequent; membrane nearly hyaline, strongly differentiated from corium; fore coxal cavities closed; ocelli small; antennae clavate; eyes large occupying most of lateral head surface, but not projected on a shelf; sperm reservoir with distally expanded bulb and small straplike wings (fig. 1Z, DD). Ovipositor elongate. Spermatheca with very short simple pump (fig. 2E).

KEY TO SPECIES OF BLISSIELLA

mm. (Africa)nidus (Slater

- 3a. Smaller species, total length under 5.0 mm...
- 4. Dorsal surface bearing prominent sub-decumbent nearly tomentose silvery hairs (fig. 5E); color generally dark brown; antennal segment 2 one-fourth longer than segment 3 and at least two-thirds length of segment 4 (Africa; Madagascar)castaneus (Slater)
- 4a. Dorsal surface appearing nearly glabrous, bearing only short, non-tomentose hairs (fig. 5F); color usually black, shining; antennal segments 2 and 3 subequal in length, segment 2 only slightly more than one-half length of segment 4 (Africa)micans (Slater)

BLISSUS BURMEISTER Figure 24

Blissus Burmeister, 1835, p. 290.

Neoblissus Bergroth, 1903, p. 253. NEW SYN-ONYMY.

Type Species: Opinion 705 (1964) of the International Commission of Zoological Nomenclature placed Blissus on the official list of generic names with Blissus hirtulus Burmeister as type species. Since I treat hirtulus in the genus Geoblissus, an explanation is necessary. Slater and China (1961) recognized that the genus Blissus as then understood was composite and that hirtulus the monobasic type was not congeneric with the economically important "Chinch Bugs" of North America. Accordingly, Slater and China (ibid.) appealed to the Commission to set aside hirtulus Burmeister as type species and designate the North American Chinch Bug Blissus leucopterus (Say) as type species. Unfortunately, both Dr. China and I believed the request to be so obviously desirable that we did not ask the hemipterological community to support the appeal. Although this appeal received a majority of the commission votes (including, I believe, those of all the entomologists), it failed to receive a sufficient number of votes. My discussion of this matter with Dr. China in 1964 indicated that a negative letter had been received by the commission from Dr. Eduard Wagner who was concerned that such action would affect a number of European species at that time placed in the genus Blissus. This was unfortunate as the European species involved are not congeneric with either

Blissus in the sense of leucopterus (Say) or hirtulus Burmeister. Subsequently this colleague wrote the Commission and sought to "rectify" possible effects of his letter, but no action has been taken by the Commission. I believe that the matter is still basically unre-

solved despite the action of the Commission, and continue to use *Blissus* in the sense of *leucopterus* (Say) although this is not in accord with the action of the Commission.

A further complication arises from the lack of distinguishing characters between *Blissus*

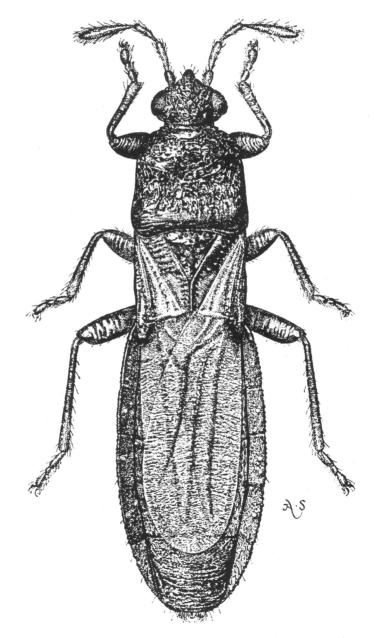


Fig. 23. Blissiella pauliani, dorsal view.

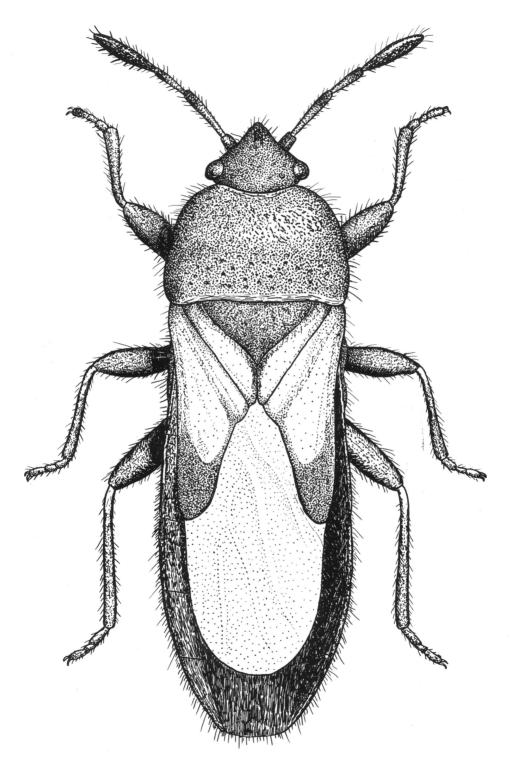


Fig. 24. Blissus mixtus, dorsal view.

(sensu leucopterus) and the South American myrmecophiles placed in the genus Neoblissus.

DISTRIBUTION: Western Hemisphere.

BIOLOGY: Some species serious pests of grain crops and lawn grasses. Breeding occurs on many species of Gramineae (see Slater, 1976).

DIAGNOSIS: Body short, stubby, thick; metathoracic scent gland auricle small, short, rounded and earlike (fig. 9K); all femora mutic; head, pronotum and scutellum above completely pruinose (only tylus shining); completely pruinose below on head and thorax; membrane comparatively thin and semitranslucent; wing polymorphism present, brachypters and micropters common; macropters with moderately concave apical corial margin; fore coxal cavities open; no marked sexual dimorphism; antennae terete or moderately clavate; ocelli small; sperm reservoir with large cup and elongate, slender, straplike wings; ovipositor elongate.

Discussion: Unfortunately no key to the species of this important taxon is available. *Blissus* has been under revision by Dr. D. E. Leonard for a number of years. For keys to the North American species see Leonard (1966, 1968).

The genus *Neoblissus* does not appear to have any characteristics of generic significance that it does not share with Blissus and is here synonymized. The species of *Neoblissus* are inquilines in the nests of ants where according to Drake (1951) they feed and breed on the grasses growing inside the chambers. Drake (ibid.) while retaining Neoblissus noted that the genus was distinguishable "largely by the very short wing pads with wide and subtruncate apex in the brachypterous form." If one wishes to follow the action of the International Commission on Zoological Nomenclature as discussed above, Neoblissus Bergroth is available for use for the Chinch bugs of the Western Hemisphere.

> BOCHRUS STÅL Figure 25

Bochrus Stål, 1861, p. 145.

Type Species: Bochrus poecilopterus Stål. Monobasic.

DISTRIBUTION: Southern Asia, Java.

BIOLOGY: Unknown.

DIAGNOSIS: Body broad, greatly flattened; metathoracic scent gland auricle very elongate, curving anteriorly throughout, expanded distally (fig. 13C); all legs multispinose; fore femora with only two or three distally located spines; hind legs much larger and more robust in male than in female; second tarsal segment very small; head and pronotum completely shining; traces of pruinosity present mesally along anterior margin or prosternum; scutellum pruinose laterad of a broad basally expanded central shining area; apical corial margin almost straight; membrane opaque, thickened; fore coxae closed; ocelli small; antennae terete; ovipositor dividing sternum six, elongate; a series of large calloused spots present on abdominal sternum. Sperm reservoir minute, apparently reduced to a small median distally rounded projection.

KEY TO SPECIES OF BOCHRUS

Pronotum bicolored, anterior half black, posterior half a strongly contrasting pale orange-yellow; legs with at least femora yellowish to ochraceous; male hind tibiae lacking a greatly enlarged basal spur......

la. Pronotum nearly uniformly black or very dark mahogany, never with anterior and posterior halves strongly contrasting (although sometimes with a narrow pale basal stripe); legs castaneous to nearly black; male hind tibiae with an enormous basal spur which is one-third to one-half length of tibiae (fig. 25)

CAPODEMUS SLATER AND SWEET Figure 26

Capodemus Slater and Sweet, 1972, pp. 211-212.

Type Species: *Blissus rusticus* Stål. By original designation.

DISTRIBUTION: Southern Africa.

BIOLOGY: Various species breed on Gramineae, Cyperaceae, and Restionaceae.

DIAGNOSIS: Body linear, moderately slender; metathoracic scent gland auricle elongately lobate, earlike, slightly curved posteriorly (fig. 9C); fore femora mutic or with a single minute ventral spine; tibiae unspecialized; head and

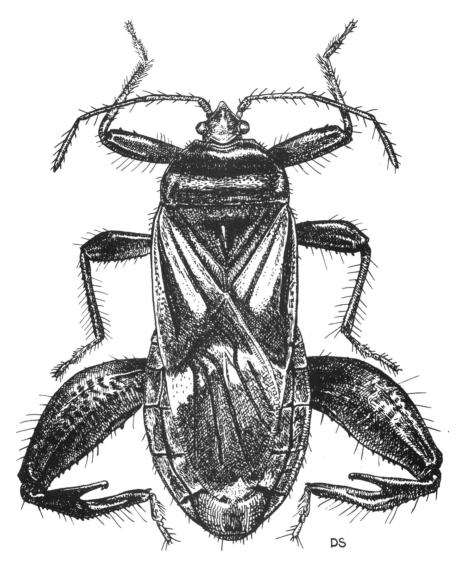


Fig. 25. Bochrus foveatus, dorsal view.

thoracic pruinosity variable, pronotum above usually, in large part, shining but frequently with pruinose areas present anteriorly and laterally in transverse impression; propleuron sometimes completely pruinose, sometimes with entire posterior lobe shining; apical corial margin straight, wing polymorphism frequent, often with extreme microptery; fore coxal cavities open; ocelli small; antennae terete; sperm reservoir with small cup and relatively narrow linear wings.

Blissus navis (Slater) is assigned to Capodemus but is quite isolated from the other species all of which form a closely related group.

KEY TO SPECIES OF CAPODEMUS

- 1. Labium very elongate, extending posteriorly beyond metacoxae rostratus (Slater)
- la. Labium shorter, never exceeding metacoxae, usually not extending caudad of mesocoxae

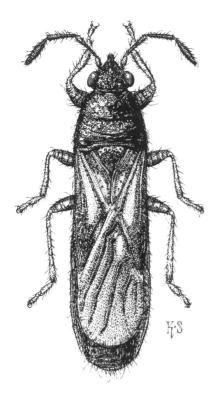


Fig. 26. Capodemus pentameri, dorsal view.

2.	Fore femora with one or two small spines
	present below on distal half (spine often
	minute)
2a.	Fore femora mutic
3.	Fore femora with two distinct spines present
	bispinosus Slater and Sweet
3a.	Fore femora with only a single small spine
	present
4.	Pruinosity on scutellum laterad of midline ex-
	tending nearly to posterior end of scutellum
4a.	Pruinosity on scutellum limited to anterior one-
	half or one-third6
5.	Acetabula shining; posterior margin of pleural
	area of prothorax with a narrow marginal
	shining stripwilcoxae Slater and Sweet
5a.	Acetabula pruinose; propleural area completely
	pruinose, including posterior margin
	stuckenbergi Slater and Sweet
6	Prothoracic pruinosity extending dorsally above
٠.	a dark shining bar near dorsal margin of
	propleural area onto lateral area of dorsal
	surface (fig. 4M)
60	
∪a.	Prothoracic pruinosity not present above shin-

	ing lateral bar, thus not present laterally on dorsal surface (fig. 5N, O)
7.	
7a.	Scutellum with pruinosity present at least on anterior third8
8.	Scutellar pruinosity laterad of midline extending posteriorly to or nearly to posterior end
Q ₀	of scutellum
	third to one-half of scutellum13
9.	inose adjacent to margin; mesosternum with a conspicuous longitudinal furrow present
9a.	Propleural area completely pruinose; meoster- num lacking a conspicuous longitudinal fur-
10.	row
10.	ends (fig. 6L)sabulosus Slater and Sweet
10a.	Brachypters with wing pads acuminate at distal
11.	ends (fig. 6J) (macropters unknown)11 Head relatively elongate, tylus exceeding distal
	end of first antennal segment; pronotal pru- inosity extending mesally across transverse impression to mesad of level of ocelli
lla.	Head relatively short, tylus usually not exceeding distal end of first antennal segment; pronotal pruinosity not strongly developed across transverse impression mesad of ocelli
12.	Eyes large, ovoid (fig. 7M)
12a.	Eyes smaller, subelliptical (fig. 7L)
13.	Prothoracic pruinosity not extending dorsally above shining bar (fig. 5N, O); posterior lobe of propleural area completely shining; acetabula shining (fig. 26)
13a.	Prothoracic pruinosity extending dorsad of shining bar onto lateral portion of dorsal surface (fig. 4M); posterior propleural area pruinose; at least fore acetabula pruinose.
14.	Meso- and meta-acetabula shining; dorsal sur-
	face densely clothed with elongate somewhat wooly pubescence
14a.	
15.	tively short and decumbent
	three or four striae visible (fig. 6J); scutellum deeply foveate laterally; brachyp-

CAVELERIUS DISTANT Figures 27, 28

Cavelerius Distant, 1903, p. 44.

Type Species: Cavelerius illustris Distant. Monobasic.

DISTRIBUTION: Southern Asia, Taiwan, Ryukyus, Japan.

BIOLOGY: Some species serious pests of sugar cane. Others taken on other grasses.

DIAGNOSIS: Body linear, moderately slender. Metathoracic scent gland auricle earlike, rounded but moderately elongate. Fore femora mutic. Pronotum dorsally with anterior half strongly shining, posterior half including humeral area pruinose. Lateral (below bar) and ventral surfaces of thorax and scutellum pruinose. Apical corial margin straight, membrane opaque but thin, noticeably differentiated from adjacent corium. Fore coxal cavities closed. Ocelli small. Antennae terete. Wing microptery, brachyptery, and submacroptery common. No noticeable sexual dimorphism. Spermatheca with large basal flange of bulb; pump short or elongate, always broadened noticeably at distal end.

DISCUSSION: This genus is very closely related to *Ischnodemus*. The large bulb and wings of the sperm reservoir (fig. III) reinforce the external similarities. Nevertheless the more apomorphic species (in particular the type species *illustris*) have the caudolateral margins of the pronotum produced strongly backward to terminate in a subacute point. The situation is somewhat analogous to that of *Patritiodemus* in that the more plesiomorphic species are similar to species of *Ischnodemus* (although with synapomorphy) but the more apomorphic species are strongly differentiated.

KEY TO SPECIES OF CAVELERIUS

1. Second antennal segment uniformly brown or black, strongly contrasting with pale first segment and unicolorous or nearly so with la. Second antennal segment white or light tan at least on proximal two-thirds, unicolorous with segment one; distal third of second and all of third and fourth segments dark or first three antennal segments light tan (if second segment largely dark then apex of abdominal connexivum seven acute) 6 2. Membrane tapering to a blunt point posteriorly, extending caudad only onto sixth abdominal tergum; membrane color in great part white with a large irregular central brown patch (Okajima) 2a. Membrane either extending onto seventh abdominal tergum, broad and non-tapering at tip and entirely brown except for small pale areas at base, adjacent to apex of corium and at apex of hemelytra, or reduced to short 3. Length of second antennal segment greater than interocular space; femora black or very dark nigrolimbatus Slater and Miyamoto 3a. Interocular space greater than length of second antennal segment; femora orange or yellow 4. Length of head greater than length of fourth antennal segment; tylus extending forward to distal end of first antennal segment (hemelytra frequently reduced to short "pads") (fig. 27)..... mishmiensis Slater and Miyamoto 4a. Fourth antennal segment longer than length of head; tylus extending forward only onto anterior third of first antennal segment5 5. Posterior margin of pronotum laterad of base of scutellum extended posteriorly as prominently produced lobes, these lobes frequently acute at apices; fourth antennal segment only slightly more than one and one-half times length of segment three ... illustris Distant 5a. Posterior margin of pronotum nearly evenly concave, not produced into prominent posteriorly directed lobes laterad of base of scutellum; fourth antennal segment long, more than one and three-fourths times as 6. First three antennal segments uniformly bright

orange-yellow, only fourth segment darkened

- - 7. Small species less than 5 mm. in length; inter-

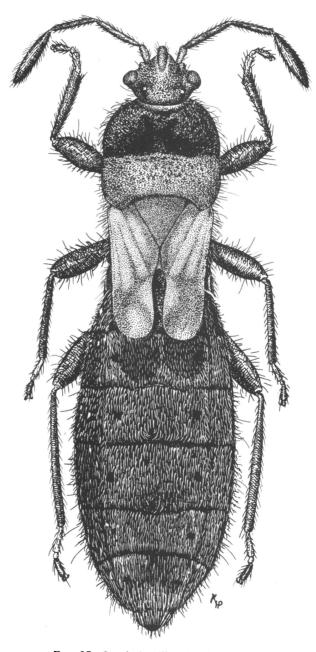


Fig. 27. Cavelerius illustris, dorsal view.

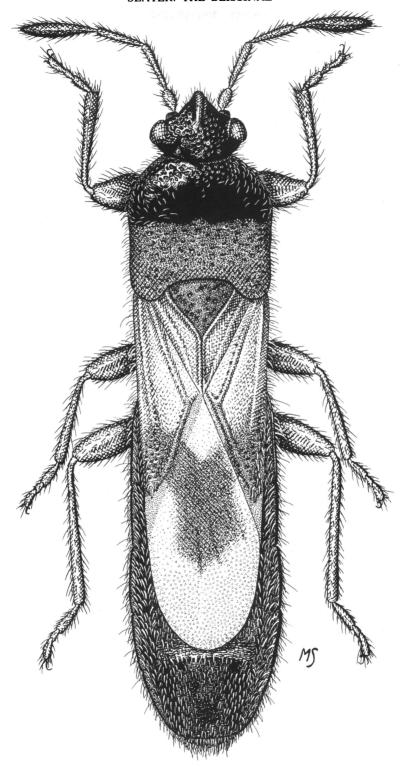


Fig. 28. Cavelerius tinctus, dorsal view.

- 8. Length of second antennal segment greater than or subequal to interocular width; fore femora dark brown; spiracles large, those on segment seven nearly one-third as great as total width across connexivum; subbasal lobe of paramere flat on distal surface............
- 8a. Interocular space greater than length of second antennal segment; femora bright yellowish; spiracles very small only about one-eighth of the total connexival width; subbasal lobe of paramere rounded excavatus (Distant)
- 9. Antennal segment 2 considerably longer than interocular space.....
- 9a. Interocular space greater than length of second antennal segment (fig. 28).....

CAVELOBLISSUS SLATER AND WILCOX Figure 29

Caveloblissus Slater and Wilcox, 1968, pp. 42-45.

TYPE SPECIES: Caveloblissus americanus Slater and Wilcox. Monobasic.

DISTRIBUTION: South America.

BIOLOGY: Unknown.

DIAGNOSIS: Body stocky (but elongate relative to sister group); metathoracic scent gland auricle short, rounded, earlike; all femora mutic; pronotum above with anterior one-half shining, posterior one-half pruinose; venter of head, sternum, posterior half of propleuron and scattered areas of anterior portion of propleuron pruinose; apical corial margin moderately concave mesally; membrane thinner textured than corium, translucent; fore coxal cavities open, ocelli small; no sexual dimorphism; antennae terete; sperm reservoir with large slender "cup" and elongate slender straplike wings. Spermatheca with large bulb and short straight pump that enlarges slightly at proximal end (fig. 2H).

CHELOCHIRUS SPINOLA Figure 30

Chelochirus Spinola, 1839, pp. 1-4.

Type Species: Chelochirus atrox Spinola. Monobasic.

DISTRIBUTION: Southeast Asia, East Indies. BIOLOGY: Unknown.

DIAGNOSIS: Body large, elongate, robust, broadened, and somewhat flattened. Metathoracic scent gland auricle large, elongate, usually strongly curved anteriorly in a lunate arc (fig. 10E) but sometimes recurved into a hooklike structure (fig. 10G). Fore femora enormously enlarged and incrassate, armed below with several huge spines or spurs. Fore tibiae shortened, thickened, toothed distally and apparently fossorial. First tarsal segment somewhat flattened, considerably enlarged. Body above strongly shining. Prothorax below either completely shining or shining anterior to acetabula and pruinose posterior to acetabula. Corium shining along radial vein and extensively along lateral margins. Membrane thickened and opaque nearly unitextured with corium. No sexual dimorphism evident and no wing reduction known. Ocelli very large and conspicuous. Antennae clavate at distal ends, short and very thick throughout (similar to those of Spalacocoris). Sperm reservoir reduced to a small median projection with tiny blocklike or straplike diverging wings present; spermatheca with bulb large, but basal flange reduced or absent; pump short, much broadened distally. Ovipositor reduced.

KEY TO SPECIES OF CHELOCHIRUS

- 3. Labium reaching mesocoxae, corium uniformly brick-red; fore femora lacking a serrated

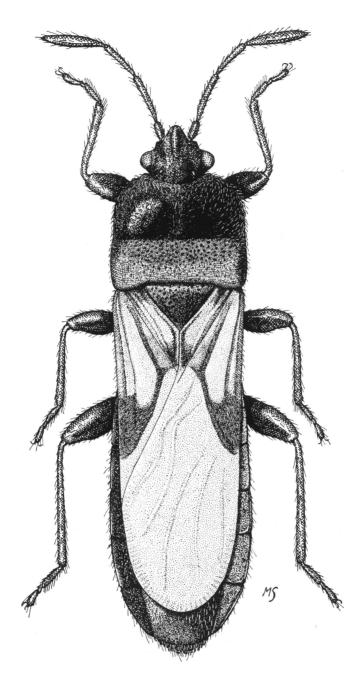


Fig. 29. Caveloblissus americanus, dorsal view.

flange distally on inner surface; legs black; metathoracic scent gland auricle curving pos-metacoxae; corium variegated with yellow

and black stripes; fore femora bearing an expanded serrate flange distally on inner surface; legs pale yellow; metathoracic scent gland auricle curving anteriorly (fig. 10E)

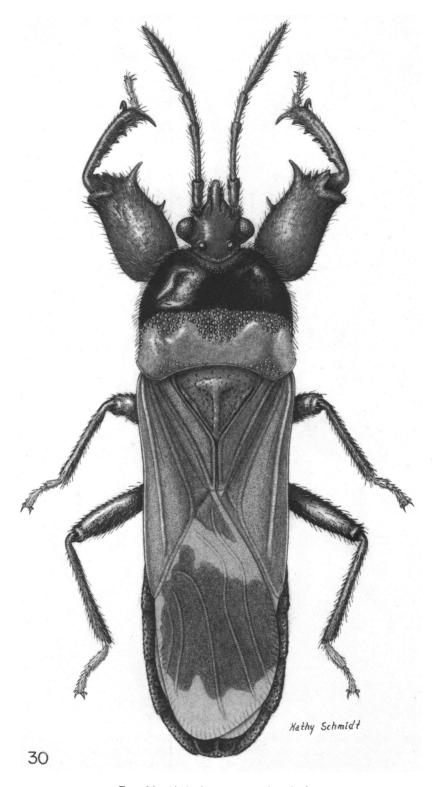


Fig. 30. Chelochirus atrox, dorsal view.

DENTISBLISSUS SLATER Figure 31

Dentisblissus Slater, 1961, pp. 481-482.

TYPE SPECIES: Ischnodemus venosus Breddin, 1900. By original designation.

DISTRIBUTION: Australia, New Guinea, New Britain, New Ireland.

BIOLOGY: Breeding records on wild sugar cane.

DIAGNOSIS: Body relatively broad but not strongly flattened; metathoracic scent gland auricle (fig. 11I) elongate and backward curving, either rounded or acute at distal end; all legs multispinose, middle and hind femora with very short stubby spines (almost absent in females); first tarsal segment very large, broadened with a "pad" of hairs below; head completely shining, pronotum in large part shining, usually with a broad pruinose band across center of disc and a narrower anterior band just behind "collar" and sometimes an additional pruinose band across basal area; almost completely pruinose below; scutellum pruinose but with a shining center stripe; both corium and membrane thick and opaque; apical corial margin straight for most of length, concave at base; fore coxal cavities closed; ocelli small; strong sexual dimorphism present, males with large bifid genal "tusks" (fig. 31); antennae somewhat clavate; sperm reservoir very small with an elongate basal stalk, wings short and broad, somewhat pear-shaped, broadly knobbed distally (fig. 1CC). Ovipositor elongate. Spermatheca with "double flanged" broad pump; bulb lacking a conspicuous basal flange (fig. 2P).

KEY TO SPECIES OF DENTISBLISSUS

- Forewing uniformly dark brown to black, with at most a minute light streak at base of membrane (fig. 31)umbrosus Slater
- la. Forewing with white areas present on clavus, corium and at least apex of membrane . . . 2

- 3a. Labium shorter, barely extending beyond fore coxae, segment two exceeding base of head by nearly one-half its length (ratio-length labium/interocular space average 2.64, range 2.40-2.84)venosus (Breddin)

DIMORPHOPTERUS STÅL Figures 32, 33

Dimorphopterus Stål, 1872, p. 44. Esmun Distant, 1909, p. 330. Euhemerus Distant, 1909, p. 331. Caenoblissus Barber, 1958, p. 186. Stenoblissus Wagner and Slater, 1964, p. 69.

Type Species: *Micropus spinolae* Signoret, 1857. By original designation.

DISTRIBUTION: Eastern Hemisphere.

BIOLOGY: Breeding reported on many species of Gramineae.

DIAGNOSIS: Body generally short and stout, sometimes elongate; metathoracic scent gland auricle short, rounded, earlike; fore femora mutic or armed below with one or two ventral spines; meso- and metafemora mutic; fore tibiae either terete or expanded, flattened and spinose; dorsal surface of head and pronotum shining or subshining, always lacking pruinosity; scutellum usually completely pruinose; apical coral margin usually concave; membrane thin, translucent at least in part; fore coxal cavities open; brachypters and micropters common; ocelli small; antennae generally moderately clavate; sperm reservoir with cup large and wings slender and straplike. Spermatheca variable often with elongate curving pump; bulb usually with well developed basal flange (fig. 2N, O).

KEY TO SPECIES OF DIMORPHOPTERUS

- 1. Fore femora with one or two spines (rarely three) present below on distal third2
- la. Fore femora mutic14
- 2. Fore femora with two spines present below .3

^{&#}x27;Measured ventrally from posterior margin of eye to apex of "tusk."

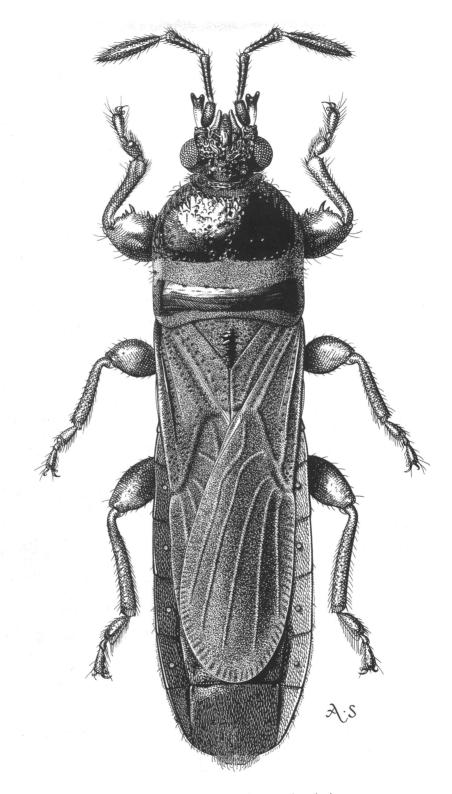


Fig. 31. Dentisblissus umbrosus, dorsal view.

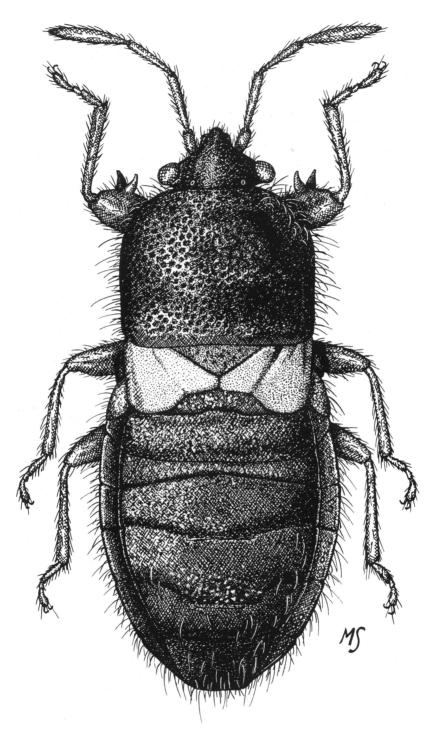


Fig. 32. Dimorphopterus cornutus, dorsal view.

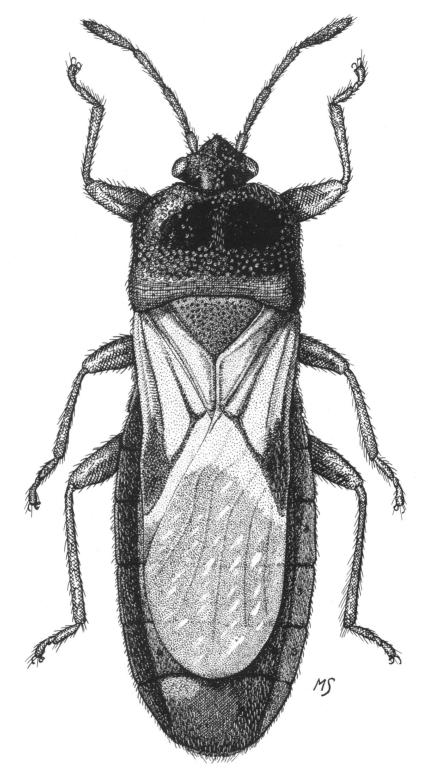


Fig. 33. Dimorphopterus gibbus, dorsal view.

2a.	Fore femora with only a single spine present ventrally (some specimens of zuluensis have		Second antennal segment less than 2.5 times length of segment 1
3.	two spines on one femur)4 Body elongate; fore femoral spines small, short, and simple (Africa)	11.	colorous with femora; membrane with large diffuse brown area adjacent to apical corial
			margin (Cape Verde Islands)
3a.	Body very short and stout; fore femoral spines		graminum (Lindberg)
	large, stout and "tusklike," inner spine	lla.	Tibiae yellow or light brown, much paler than
	sharply curved at apex to form a right-		dark femora, membrane uniformly opaque
	angled hook (Australia) (fig. 32)		white (Egypt, Sudan)
			nubicus (Wagner and Slater)
4.	Fore tibiae strongly expanded on distal half,	12.	Very small species, ♂ not over 2.7 mm., ♀
	somewhat flattened and strongly toothed to		3.0 mm.; macropters with clavus and cor-
4 -	give a fossorial appearance5		ium nearly uniformly bright yellowish tan,
4a.	Fore tibiae usually somewhat swollen distally,		lacking a distinct black macula apically on
	but rounded, not forming a ventrally flattened fossorial structure6		corium; brachypters with wing pads very small and distinctly separated mesally; color
5	Labium relatively short, at most extending be-		frequently reddish brown (Palearctic)
٦.	tween or slightly beyond mesocoxae, not		
	attaining posterior margin of metacoxae	12a.	Larger species, & always more than 2.8 mm.
	(South Africa)hessei (Slater)		in length, usually over 3.0 mm.; ♀ over
5a.	Labium elongate, extending posteriorly to or		3.3 mm., usually more than 3.5 mm. in
	beyond posterior margin of metacoxae		length; both macropters and brachypters
	(Australia)pilosus (Barber)		with hemelytra contrastingly black and
6.	Labium relatively short, extending posteriorly		white with a large conspicuous black mac-
	at most only between mesocoxae7		ula apically on corium; brachypters with
ба.	Labium elongate, extending to or almost to		hemelytra usually broadly in contact along
	metacoxae, always exceeding posterior mar-		midline; head and pronotum usually chiefly
7	gin of mesocoxae	12	black
/٠	mm.)8	13.	is usually black and strongly contrasting
7a.	Small, relatively short and stout species (less		with bright reddish or yellowish brown
	than 3.5 mm.) (Africa)similis (Slater)		tibiae; antennae longer, ratio of total anten-
8.	Labium extending well between mesocoxae; at		nal length to interocular width usually well
	least antennal segments 2 and 3 yellow to		over 3.0 (South Africa)
	brownish yellow; propleuron pruinose to		
	lateral shining line (Africa)	13a.	Size smaller, usually not over 3.0 mm.; fore
	zuluensis (Slater)		femora usually reddish brown and little
8a.	Labium shorter, not reaching mesocoxae; an-		contrasting with tibiae; antennae shorter,
	tennal segments 2 and 3 red-brown; pro-		ratio of total antennal length to interocular
	pleuron shining (Siberia) ¹		width usually less than 3.0 (Palearctic,
o	Propleural area of proportion looking a bond of	14	Ethiopian)brachypterus (Rambur)
7.	Propleural area of pronotum lacking a band of pruinosity above acetabula; head, pro-	14.	Labium elongate, extending to, or well between, or caudad of metacoxae15
	notum, and scutellum predominantly red-	14a	Labium relatively short, usually not exceeding
	brown (Africa) cornipes (Hesse)		mesocoxae, but if so not extending between
9a.	Propleural area with a distinct band of pru-		or beyond metacoxae20
	inosity above acetabula, or if pruinosity re-	15.	Metathoracic scent gland auricle acutely
	duced then head, pronotum, and scutellum		pointed at distal end16
. ~	predominantly black10	15a.	Scent gland auricle evenly and broadly
10.	Length of antennal segment 2 much more than	1.0	rounded at distal end17
	2.5 times length of segment 111	16.	Labium exceeding metacoxae, reaching poste-
			rior margin of metasternum (Oriental)lepidus Slater, Ashlock, and Wilcox
¹See	footnote p. 50.	16a.	Labium shorter, reaching only to anterior mar-

	gin of metacoxae (Oriental)		most of length, becoming concave only
17	Propleural area of prothorax lacking pruinosity		mesally near distal end of claval com- missure
17.	above acetabula (South Africa)	25.	Larger species, over 4.0 mm. in length; pro-
	fulgidus (Slater)		thoracic pruinosity confined to anterior por-
17a.	Propleural area with pruinosity present above		tion of sternum, not extending dorsad of
	acetabula18		acetabula (Oriental) indicus Slater
18.	Membrane of fore wing chiefly hyaline with a	25a.	Smaller species, less than 3.75 mm.; pru-
	contrasting opaque area anteriorly and		inosity extending dorsad of acetabula on
	broadly in contact with apical margin of corium (fig. 33); size 3.0 mm. or less (Ori-	26	prothorax
	ental)	20.	cupying entire apical angle and continuing
	rondoni Slater, Ashlock, and Wilcox		as a complete broad band along entire api-
18a.	Membrane uniformly opaque white throughout;		cal margin and across distal ends of clavi to
	size larger, 3.9 mm. or greater19		form a continuous arcuate stripe across
19.	Labium extremely elongate, extending midway		hemelytra; body relatively slender and
	to apex of abdomen (South Africa)		elongate, total length considerably more
	oblongus (Stål)		than three and a quarter times width across
19a.	Labium shorter, scarcely exceeding posterior	26-	pronotum (Oriental) typicus (Distant)
	margin of metacoxae (South Africa)	20a.	Darkened distal area of corium sometimes nar- rowly extending to and along cubital vein
20	Scutellum completely shining and non-pruinose		but never forming a complete broad arcuate
-0.	(Oriental) anomalus Slater		stripe across hemelytra; body relatively
20a.	Scutellum either completely pruinose or with		short and stout, total length less than three
	pruinosity present basally and laterad of		and a quarter times pronotal width 27
	shining median area21	27.	Propleuron with a distinct pruinose area dorsal
21.	Membrane with posterior portion hyaline or		to acetabula; scutellum completely pruinose
	subhyaline, strongly contrasting with opaque anterior postion lying adjacent to	270	(Oriental; Japan)bicoloripes (Distant)
	apical corial margin, or membrane almost	2/a.	Propleuron with pruinose area not extending as a broad band dorsally above acetabula;
	entirely hyaline22		scutellum with a median traingular shining
21a.	Membrane completely opaque throughout28		elevation (Oriental) erebus (Distant)
22.	Body very elongate and linear; macropters with	28.	Membrane fuscous brown with a broad white
	distance from apex corium to apex abdo-		lunate transverse vitta extending across
	men more than twice basal width of pro-		membrane just distad of distal end of cor-
	notum (brachypters unknown) (South Africa)tenuatus (Slater)	280	ium (Oriental) atromaculatus (Distant)
22a.	Body relatively broad and stout, macropters	20a.	Membrane generally pale or suffused with fuscous, but always lacking a conspicuous
	with distance from apex corium to apex		white vitta beyond disal end of corium
	abdomen subequal to or less than twice		29
	basal width of pronotum (some species fre-	29.	Size smaller, usually less than 3.5 mm. in
22	quently brachypterous)23		length; if larger (latoides) then membrane
23.	Pronotun appearing nearly glabrous with only		of hemelytra hyaline, opaque only ante-
	extremely short inconspicuous hairs present; large species, over 4.5 mm. (Oriental) (fig.	200	riorly adjacent to corium
	33)gibbus (Fabricius)	274.	length; if smaller over 3.5 mm. and with
23a.	Pronotum appearing hirsute with conspicuous		membrane of hemelytra opaque throughout
	often upstanding although frequently only		33
	semi-decumbent hairs present; not more	30.	Scent gland auricle sharply pointed at distal
	than 3.8 mm. in length (or if hairs short	20	end (Oriental)sumatrensis Slater
	and inconspicuous than smaller species not over 4.25 mm. in length)24	30a.	Scent gland auricle broadly rounded distally.
24.	Apical corial margin deeply concave for most	31	Pronotal hairs semierect; body shape subovoid
	of extent (Oriental)	51.	(Madagascar)
24a.	Apical corial margin straight or convex for	31a.	Pronotal hairs completely declivent; body form
			·

subparallel and relatively elongate (Africa) 32. Labium extending posteriorly well between mesocoxae; fourth segment longer than segment 3 and subequal to segment 2 (body length 3.15 mm.) upembensis (Slater) 32a. Labium not attaining mesocoxae; fourth labial segment equal in length to segment 3 and shorter than segment 2 (body length 3.8 33. Anterior acetabula pruinose (Palearctic)..... 34. Legs with at least femora dark brown to black, usually contrasting with pale tibiae, antennae nearly unicolorous dark brown (Palearctic)¹spinolae (Signoret) 34a. Legs and first, second and third antennal segments yellowish brown, fourth and sometimes third antennal segment contrasting dark red-brown (Palearctic)blissoides (Baerensprung)1

EXTARADEMUS SLATER AND WILCOX Figure 34

Extarademus Slater and Wilcox, 1966, pp. 62-63.

Type Species: *Macropes collaris* Signoret, 1957. Original designation.

DISTRIBUTION: South and Central America, southern North America, Cuba.

BIOLOGY: Breeding known to occur on several genera of Gramineae.

DIAGNOSIS: Body elongate, slender, non-flattened; metathoracic scent gland auricle elongate, tapering, curving forward, scimitarshaped (fig. 9N) or occasionally rounded at distal end; femora with one ventral spine; males of some species with a large ventral spine on hind femur; pruinosity variable, frequently with head above shining mesally, pruinose laterally, occasionally completely shining; pronotum dorsally varying from condition of maximum pruinosity where shining areas are present as a pair of large triangular calli patches and a complete humeral band, to completely shining; scutellum and lateral and ventral surfaces of prothorax completely pruinose; apical corial margin straight; membrane subhyaline or opaque but much thinner than corium; brachyptery common; fore coxal cavities closed; ocelli small:

¹See footnote p. 50. These authors believe *blissoides* as treated here is synonymous with *spinolae*.

most species strongly sexually dimorphic, males usually with more incrassate hind femora, patches of spines and tubercles on abdominal venter (fig. 8A) and sometimes head and pronotal modifications; antennae terete; sperm reservoir with small often stalked "cup" and elongate, slender, diverging straplike wings (fig. 1E, F); ovipositor elongate. Spermatheca with large bulb, short distally expanded pump and elongate tube.

KEY TO SPECIES OF EXTARADEMUS

- 1a. Pronotum shining over entire dorsal surface . . 4
 2. Tylus much broadened at anterior end to form a splayed-out truncate apex; males with a pair of sharp spines projecting from ends of connexiva seven tylosis Slater and Wilcox
- 3. Second antennal segment relatively elongate, always more than one and one-quarter times as long as interocular space (1.31-1.50); males with a pair of acute spines projecting from ends of abdominal connexiva; abdominal sterna 5, 6, and 7 swollen, tumid and armed with a series of acute tubercles.......
- 3a. Length of second antennal segment usually subequal to or only slightly greater than interocular distance (never more than one and one-fifth interocular distance); males lacking caudally projecting spines at end of connexiva; abdominal sterna with a series of small tubercular spines, but segment 5, 6, and 7 not tumid and swollen (fig. 8A).......
- Membrane with a large distinct median dark discal spot (Cuba)discalis (Barber)

 Membrane often suffused, but lacking a discal
- 4a. Membrane often suffused, but lacking a discal spot......5

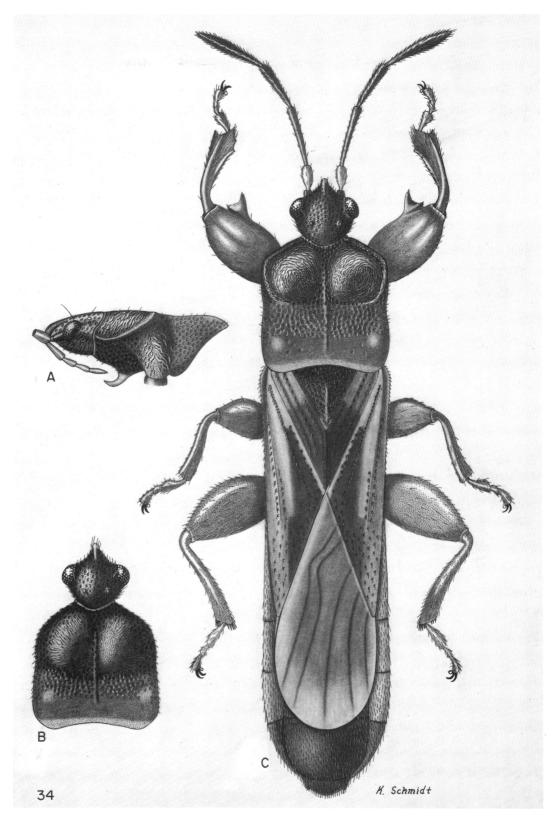


Fig. 34. A Extarademus humerus, male, dorsal view. B. Same, head and prothorax, lateral view. C. Extarademus humerus, female, head and pronotum, dorsal view.

- 5a. Labium more elongate, extending posteriorly almost to fore coxae, third labial segment considerably exceeding base of head6

- 7a. Males lacking a projecting spine from posterior margin of eighth sternum, latter with serrate edges and median bulge, but lacking a distinct spine ...collaroides Slater and Wilcox

EXTARAMORPHUS SLATER, ASHLOCK, AND WILCOX Figure 35

Extaramorphus Slater, Ashlock, and Wilcox, 1969, pp. 698-699.

Type Species: Extaramorphus magnatarsus Slater, Ashlock, and Wilcox. Monobasic.

DISTRIBUTION: Southeast Asia.

BIOLOGY: Unknown.

DIAGNOSIS: Elongate, sublinear, extremely flattened dorsoventrally. Metathoracic scent gland auricle strongly bent anteriorly, "L"shaped. Fore femora strongly incrassate, multispinose. Middle and hind femora enlarged, shortened, strongly thickened with a series of subbasal spines below; hind tibiae extremely short and stout, scarcely longer than greatly swollen and enlarged first tarsal segment. Body above and below completely shining, no pruinose areas present. Membrane much thinner than corium. Apical corial margin straight. Fore coxal cavities closed. Ocelli small. Abdomen truncate apically with connexivum seven posteriorly produced into blunt points with additional sharp toothlike spines present dorsally along posterior margin. Clasper with outer lobe place far distinct from base. Sperm reservoir with a dorsal sclerotized plate developed from modified cup and with a pair of prominent laterally projecting winglike structures present.

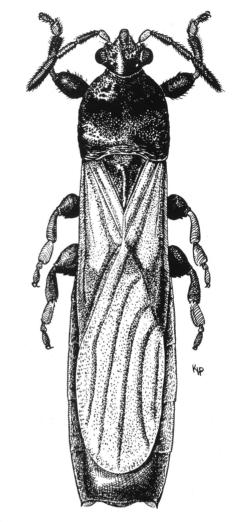


Fig. 35. Extaramorphus magnatarsus, dorsal view.

GELASTOBLISSUS SLATER AND WILCOX Figure 36

Gelastoblissus Slater and Wilcox, 1968, pp. 435, 438.

Type Species: Gelastoblissus rugosus Slater and Wilcox. Monobasic.

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

Diagnosis: Body stout, robust, elliptical. Abdomen very strongly, ovately expanded and rounded. Metathoracic scent gland auricle narrowly lobate, short and rounded. Fore femora

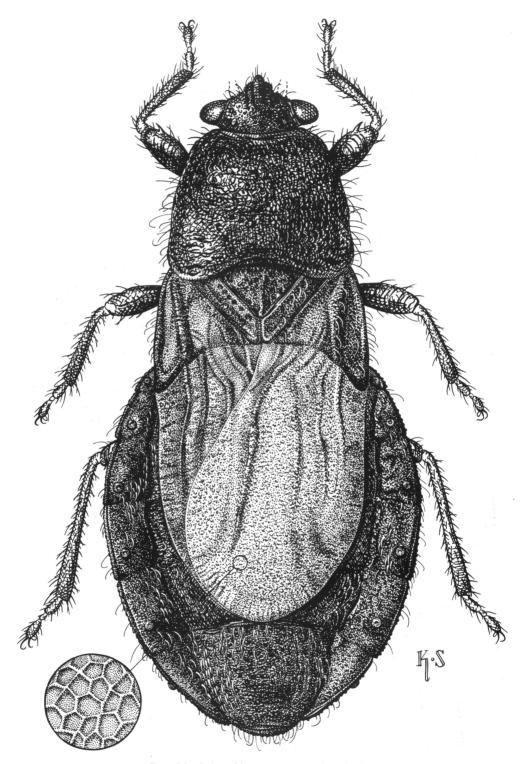


Fig. 36. Gelastoblissus rugosus, dorsal view.

moderately incrassate armed below on distal third with a single, very elongate, toothlike spine; middle and hind femora mutic. Entire body subshining above and below, completely lacking pruinose areas. Apical corial margin concave. Membrane thin, subhyaline, strongly differentiated in texture from corium; membrane composed of tiny reticulate cells. Fore coxal cavities closed. Ocelli small. Clavus and corium much shortened, membrane not covering seventh abdominal tergum thus submacropterous with membrane of one wing only partially covering membrane of other wing. Eyes laterally produced and rather transverse. Antennae and genitalia unknown.

GEOBLISSUS HIDAKA¹ Figure 37

Geoblissus Hidaka, 1959, pp. 269-270.

Type Species: Geoblissus rotundatus Hidaka. Monobasic.

DISTRIBUTION: Africa, Asia, Southern Eurasia, Japan, East Indies.

BIOLOGY: Reported as injuring sugar cane and on grasses of the genera *Elymus* and *Aristida*.

DIAGNOSIS: Body short, stout, thick; metathoracic scent gland auricle short, broadly rounded, earlike, with a short distal "hook"; fore femora mutic; fore tibiae broadly expanded and armed with rows of spines along margins; head and pronotum above and scutellum completely shining; prothorax pruinose below in front of and below coxae and behind coxae dorsad to lateral margins; body covered with elongate upstanding hairs; apical corial margins strongly concave; membrane hyaline, much differentiated from coriaceous corium; fore coxal cavities open; ocelli small; brachypters unknown; antennae clavate; sperm reservoir with well-developed cup and straplike slender wings.

KEY TO SPECIES OF GEOBLISSUS

- - ¹See discussion of type species of Blissus Burmeister.

- 2a. Scutellum usually lacking anterolateral spines.
- 3. Entire corium white to translucent, at most tinged with yellowish or tan on distal one-fourth (Africa)...siccus Slater and Wilcox
- 4. Relatively large species, ♀ 4.15-4.20 (♂ unknown); dark brown coloration on corium extending anteriorly half-way along cubital vein (Africa)niger Slater

- 5a. Prosternum pruinose before coxae and usually on propleuron posterior to acetabulum 6
- 6. Width of head much greater than half pronotal width; length of scutellum much more than half of scutellar width (30:49) (Oriental) (fig. 37).....

HEINSIUS DISTANT Figure 38

Heinsius Distant, 1901, p. 469.

Type Species: Heinsius explicatus Distant. Monobasic.

DISTRIBUTION: Australia. BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, linear, not flattened; metathoracic scent gland auricle rounded, lobate; fore femora with a single small ventral spine present; body lacking pruinosity above and below; body and appendages clothed with numerous conspicuous flattened scalelike hairs; apical corial margin concave; membrane much thinner than corium, lacking reticulate veins; corium shortened; antennae short, stout, segments rather nodular, segments three and four narrower than segments one and two; ninth and tenth female abdominal para-

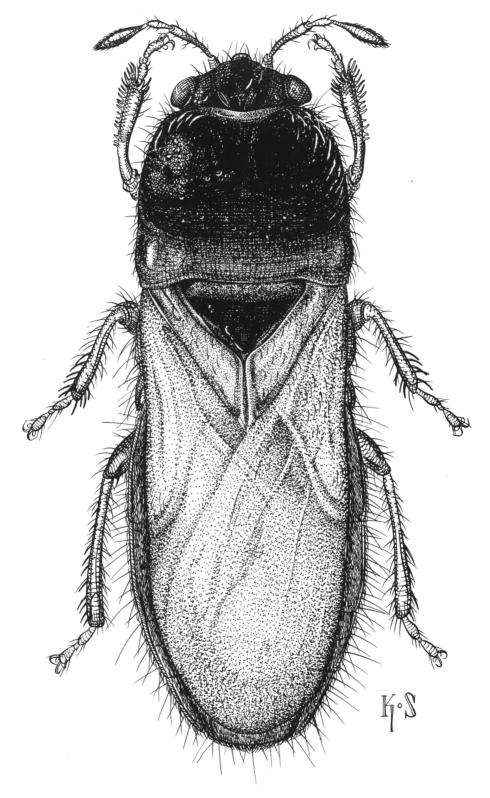


Fig. 37. Geoblissus mekongensis, dorsal view.

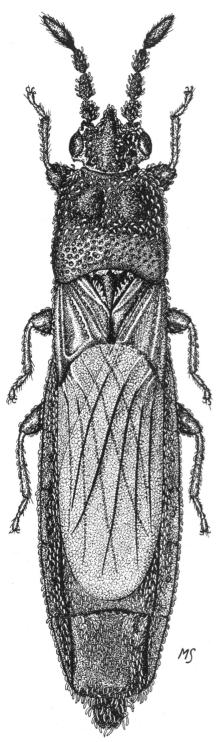


Fig. 38. Heinsius explicatus, dorsal view.

tergites elongated and extending conspicuously caudad of segment eight (fig. 8H); abdominal sterna three thru six each with only two trichobothria present; sperm reservoir reduced to a small distally acute scalelike process (fig. 1T, U). Spermathecal bulb appearing to be secondarily invaginated; with a short pump (fig. 2F); fifth instar nymph lacking conspicuous posterior sclerotization.

KEY TO SPECIES OF HEINSIUS

1a. Head width greater than one and one-fourth times interocular distance; antenniferous tubercles diverging strongly cephalolaterad from bases; veins of membrane chocolatebrown, contrasting conspicuously with pale membrane explicatus Distant

HETEROBLISSUS BARBER Figure 39

Heteroblissus Barber, 1954, p. 221.

Type Species: Heteroblissus anomilis Barber. Monobasic.

DISTRIBUTION: South America.

BIOLOGY: Unknown.

DIAGNOSIS: Body robust, stout but moderately linear. Metathoracic scent gland auricle short, rounded, earlike (fig. 9G). Fore femora incrassate armed below with one large and several small spines. Body above and below completely shining, lacking pruinose areas. Apical corial margin concave basally but straight for greater portion of length. Membrane thin, semihyaline, considerably thinner than adjacent corium. Fore coxal cavities open. Fore tibiae not swollen but with a series of small tubercles along shaft. Ocelli small. Extreme microptery common with wings reduced to tiny scalelike pads that do not reach abdomen. Abdomen with a rather large striated (apparently stridulatory) area present. Sperm reservoir variable sometimes with very large distally broadened bulb and wings broad basally and tapered distally (fig. 1AA). In some species bulb appear-

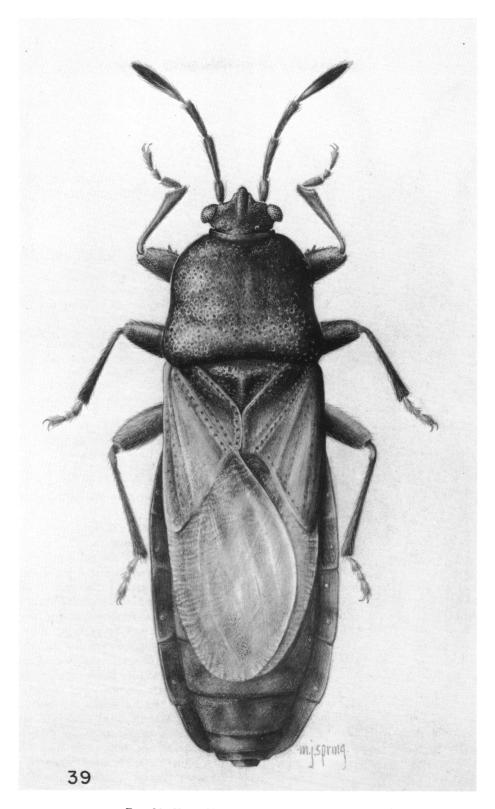


Fig. 39. Heteroblissus anomilis, dorsal view.

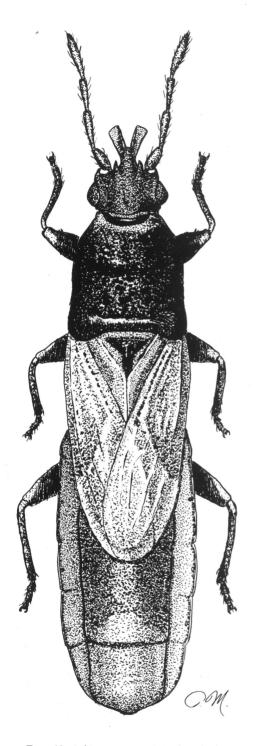


Fig. 40. Iphicrates gressitti, dorsal view.

ing to be completely membranous irregular sclerotized areas probably representing wing remnants. Ovipositor elongate. Spermathecal bulb very large, pump short and stout (fig. 2-I).

IPHICRATES DISTANT Figure 40

Iphicrates Distant, 1903, pp. 44-45. Anisosoma Bergroth, 1918, pp. 69-70.

Type Species: Iphicrates subauratus Distant. Monobasic.

DISTRIBUTION: Australia, New Caledonia, Solomon Islands through East Indies to Southern Asia, Taiwan, Japan, Philippines.

BIOLOGY: Poorly known, some evidence of breeding on bamboos and other grasses.

DIAGNOSIS: Body moderately elongate, linear, not strongly flattened. Metathoracic scent gland auricle little produced above adjacent body surface, usually straight, not strongly tapered to distal end, occasionally curved anteriorly and sometimes angled posteriorly (fig. 11B, C, D, F). Fore femora moderately incrassate usually armed below with two spines. Middle and hind femora usually mutic occasionally with a series of small spines present. Pruinosity variable. Dorsal surface of head and pronotum subshining, sometimes appearing granulose, with a partial or complete ring of pruinosity present anteriorly just behind collar. Lateral and ventral surfaces with variable pruinosity—from completely shining to completely pruinose. Scutellum laterally either pruinose or shining. Apical corial margin usually straight but sometimes strongly concave. Membrane much thinner than corium. Fore coxal cavities closed. Ocelli small. Marked sexual dimorphism present, males with bucculae much enlarged projecting forward in front of tylus and frequently distally expanded. Antennae terete or moderately clavate. Antenniferous tubercles hooked (fig. 6E-G). Juga of males frequently raised above head surface and elongated as a pair of projecting horns. No noticeable transverse pronotal impression present. Microptery uncommon but known to occur, submacroptery common. Tarsi sometimes two-segmented. Claspers conventional. Sperm reservoir with bulb stalked, margin thick, wings protruding at right angles to bulb, broad basally, distally terminating in an anvil-shaped apex (fig. 1BB) in some species much reduced. Spermathecal pump double flanged (fig. 2K), similar to condition found in *Macropes*.

KEY TO SPECIES OF IPHICRATES

- 1. Dorsal surface of each abdominal connexival segment bearing a brown patch on anterior one-third to one-half (only extreme micropters known with clavus and corium fused, membrane reduced to a small marginal flap and extending posteriorly only onto anterior portion of second abdominal tergum) (New Guinea)neotenicus Slater
- la. Dorsal surface of connexivum lacking differentiated dark brown patches (occasionally a small black spot present on posterolateral areas) (usually macropterous or submacropterus, if wings reduced then with clavus and corium distinct, membrane well developed and extending posteriorly at least well onto third abdominal tergum)

- 3a. Apical corial margin straight or only slightly concave along inner one-third; veins of membrane at most only slightly darker than ground color, not strongly contrasting . . . 6
- 4. Large species (9.20 mm.); labium extending only to anterior margin of fore coxae; bucculae produced forward as narrow, straight spikes, widely separated mesally for entire length (fig. 6H) (Philippines) . . rex Slater

- 6. Bucculae narrow and linear or converging, never strongly broadened and expanded dis-

- tally; if slightly expanded distally then juga attaining or exceeding apex of tylus 76a. Bucculae becoming strongly expanded distally
- 7a. Bucculae either rounded or truncate at apices
- 8a. Bucculae projecting forward as small slender subacute spikes, very broadly separated from one another for entire length9
- 9a. Middle and hind femora mutic; each fore femur bearing a large bifid spine below on distal one-third; membrane opaque, usually tan to dark brown (testaceous in some specimens) (New Guinea)nigritus Slater
- Juga attaining or exceeding apex of tylus, long and slender; apices of bucculae truncate (fig. 6F) (Malaya) malayensis Slater

- 11a. Bucculae nearly linear, never broadly in contact with one another along midline (Japan)
 spinicaput (Scott)
- 12. Bucculae strongly angulate from inner angles to lateral corners, or broadly rounded ...13

- 13a. Bucculae gradually expanded for greater portion of length, lateral margins strongly convex, dorsal surfaces concave, having a "scooped-out" appearance; each fore femur with three sharp ventral spines15
- 14. Labium relatively short, not exceeding fore coxae, second segment attaining base of head; pronotum uniformly shining, lacking a complete distinct pruinose band posterior

	to collar area; small species (3.64-3.75		
	mm.) (New Guinea)cervinellus Slater	20.	Each fore femur with three sharp spines below,
14a	Labium longer, nearly reaching posterior mar-		distal two spines divergent, each set on a
ı ıu.	gin of prosternum, second segment surpass-		stout tubercle; antenniferous tubercles very
	ing base of head by half its length;		long, acute, exceeding juga in length (New
	pronotum with a complete, narrow, but dis-		Guinea)papuensis Slater
		200	Each fore femur with two straight spines be-
	tinct pruinose band across area posterior to	20a.	
	collar; larger species (3.92-4.04 mm.) (Tai-		low; antenniferous tubercles short, stubby,
	wan) (fig. 40)gressitti Slater		slightly curved, much shorter than juga
15.	Bucculae with lateral margins terminating in an		(New Guinea) cervinellus Slater
	acute point, sometimes slightly recurved;	21.	Bucculae not surpassing apex of tylus, distal
	juga rounded and thick; pronotum black ex-		ends of bucculae angled mesocephalad to
	cept at humeral angles; basal width of pro-		terminate near apex of tylus22
	notum greater than median length	21a.	Bucculae exceeding tylus, if by only a short
			distance then their apices acute or subacute
15a.	Bucculae with lateral margins evenly rounded,		and not angled against lateral margins of
	not terminating in an acute angle; juga		tylus
	sharp and acute; pronotum with posterior	22.	Pronotum bearing a strip of pruinosity ante-
	one-third usually dark reddish brown; me-		riorly which is complete across midline;
	dian pronotal length subequal to width		scutellum pruinose basally and laterad of
	across humeri (Tasmania) spathus Slater		median elevation23
16	Bucculae meeting or nearly meeting along	22a.	Pronotum lacking dorsal pruinosity even ante-
10.	midline for some distance in front of apex		riorly; scutellum shining, lacking pruinose
	of tylus; juga very elongate, exceeding apex		area basally and laterally24
	of tylus, projecting cephalo-mesad (Ceylon)	23	Pronotum strongly bicolored with light
	subauratus Distant	23.	brownish yellow posterior half strongly
16.			contrasting with black anterior half; abdo-
ioa.	Bucculae usually separated from one another		men above uniformly bright yellow
	along meson, if in contact then with juga		subauratus Distant
	short, thick and not attaining apex of tylus	220	
		23a.	Pronotum not strongly bicolored, chiefly black,
17.	Veins of membrane broadly darkened, strongly		with a red-brown band confined to humeral
	contrasting with uniformly opaque white		area remote from transverse constriction;
	ground color of remainder of membrane		abdomen above red-brown with connexiva
	18		sometimes a lighter yellowish brown (fig.
17a.	Membrane of fore wing usually hyaline with		40)gressitti Slater
	veins nearly unicolorous19	24.	Antennae and femora (except at distal ends)
18.	Apical corial margin deeply and evenly (ar-		black, concolorous with head and pro-
	cuately) concave; pronotum nearly com-		notum; antenniferous tubercles projecting
	pletely shining with only a small patch of		anterolaterad as straight spinelike processes
	pruinosity present anterolaterad and remote		(fig. 6H)angulatus Slater
	from midline; three central veins of mem-	24a.	Antennae and femora bright red-brown, con-
	brane broadly "connected" with a dark area		spicuously lighter than black head and pro-
	along apical corial marginrex slater		notum; antenniferous tubercles curving
18a.	Apical corial margin shallowly sinuate, not		anterolaterad, not projecting as straight,
	deeply and arcuately concave; pronotum		acute spinelike processes (fig. 6F)
	with a complete pruinose band across		
	meson posterior to collar area; two central	25.	Bucculae projecting beyond apex of tylus and
	membranal veins separated from apical cor-		in contact with one another mesally either
	ial margin by a pale area		at distal ends or along entire mesal margins
			with exception of apical area26
19.	Bucculae concave along anterior margins and	25a	Bucculae barely projecting beyond apices of
	often in contact or nearly so (Philippines;		tylus, or if so projecting then separated
	Malaya)		from one another for entire length 27
19a.	Bucculae convex or nearly straight on anterior	26	Bucculae in contact with one another mesally
u.	margins, broadly separated along meson	20.	immediately before tylus, separate only dis-
	Barry, and a special strains and an arrangement		dis

tally; membrane translucent hyaline, lacking dark brown markingslineatus Slater 26a. Bucculae separate from one another for most of length, in contact along midline only at distal ends; membrane opaque white with veins and a diffuse vitta on inner half contrastingly dark brown.....montaguei (Distant) 27. Bucculae produced beyond apex of tylus as large rounded "scooplike" lobes, noticeablyspathus Slater 27a. Bucculae produced forward of apex of tylus either as acute or rounded projections but never strongly excavated on dorsal surfaces 28. A prominent bifid spine present on each fore femur Slater 28a. No bifid spine present on each fore femur... 29. Labium extending posteriorly onto anterior portion of mesosternum.....spinicaput (Scott) 29a. Labium not exceeding posterior margin of prosternum30 30. Bucculae conspicuously exceeding apex of tylus, usually by a distance subequal to length of juga papuensis Slater 30a. Bucculae very short, only very slightly exceeding apex of tylus, by much less than length of juga31 31. Labium relatively short, not attaining fore coxae, second segment not reaching base of head (ratio length segments $1 + 3 \times 100$ / interocular distance = 70).....cervinellus Slater 31a. Labium longer, attaining fore coxae, second segment reaching to or beyond base of head (ratio length segments $1 + 3 \times 100$ / interocular distance = 82-92).....

ISCHNOCORIDEA HORVATH Figure 41

Ischnocoridea Horvath, 1892, p. 260.

Type Species: Ischnocoridea elegans Horvath. Monobasic.

DISTRIBUTION: Western Africa.

BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, robust; metathoracic scent auricle rounded, earlike (fig. 9H); propleuron not excavated for reception of fore femora; fore femora multispinose; head and pronotum above completely shining, prothorax below narrowly pruinose to level above acetabula, dorsal portion of propleuron shining; scutellum with some pruinosity present laterally near base; apical corial margin straight; membrane broadly pruinose adjacent to apical corial margin, remainder strongly shining, membrane texture thickened nearly equal to that of corium: fore coxal cavities closed; ocelli small; antennae somewhat clavate; sperm reservoir small with cup holding slender, diverging, distally enlarged wings (fig. 1W), ovipositor elongate. Spermatheca with large bulb and straight, thick pump that is not double-flanged (fig. 2Q).

ISCHNODEMUS FIEBER Figures 43-51

Ischnodemus Fieber, 1837, pp. 337-338. Micropus Spinola, 1837, pp. 218-221. Thops Gistl, 1848, p. x. Romicpus Reed, 1900, pp. 66-67.

Type Species: Ischnodemus quadratus Fieber. Monobasic.

DISTRIBUTION: All major zoogeographic regions.

BIOLOGY: Most abundant on various species of Gramineae. Also known to breed on Cyperaceae, Juncaceae, Zingiberales, and Haemodoraceae.

DIAGNOSIS: Body moderately to very elongate, slender, linear, not strongly flattened. Metathoracic scent gland auricle variable but generally rounded and earlike, or similar in conformation but more elongate (figs. 10A, B, C; 9A, B, J). Fore femora mutic or with one or two short spines present on ventral surface. rarely with three or four spines present. Middle and hind femora always mutic. Pruinosity variable from completely pruinose to completely shining above and below. Apical corial margins straight. Membrane usually much thinner than corium, rarely thickened, opaque and similar in texture to adjacent corium. Fore coxal cavities closed. Ocelli small. Sexual dimorphism generally not evident. Microptery, brachyptery, and submacroptery common. Antennae terete or

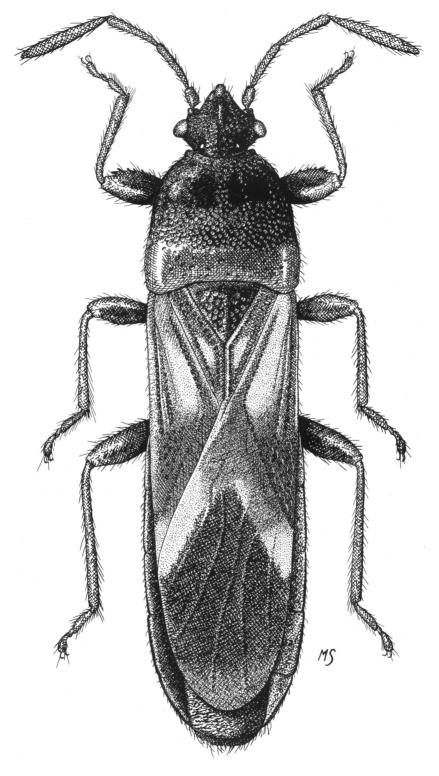


Fig. 41. Ischnocoridea picipes, dorsal view.

slightly clavate. Claspers generally of a plesiomorphic type. Sperm reservoir extremely variable from plesiomorphic with a large cup and straplike wings to reduced to a tiny cup and minute wings or a very large cup and extremely large flattened platelike wings that cover most of the membranous portion of cup (see fig. 1G-O, Q-S) (see also Slater and Harrington, 1970). Ovipositor variable in length.

DISCUSSION: Ischnodemus aleocharoides (Jakovlev) is a very unusual and anomalous species. It has apparently not been discussed in the literature since the original description by Jakovlev in the genus Dimorphopterus. Through the kindness of the British Museum, I have been able to examine a single micropterus female. The fore coxae are strongly closed indicating that it is not a species of Dimorphopterus. Lack of apomorphic features other than microptery and pruinosity loss would seem to relate it to my paraphyletic concept of Ischnodemus, where I place it provisionally. It is not included in the following key. However, it is not closely related to any other species of Ischnodemus thus far known. The head, entire prothorax (both above and below) and the scutellum are completely shining and lack any indications of pruinosity. The scutellum is very large, nearly as long as the pronotum, noticeably convex and coarsely punctate. The wings are reduced to small undifferentiated testaceous pads widely separated mesally and extending posteriorly only over the anterolateral corner of the second abdominal tergum. The fore femur is strongly incrassate and mutic. The metathoracic scent gland auricle is broadly splayed out distally to become somewhat mushroomshaped. It has many of the same highly modified features of the African Ischnodemus crassipes yet does not appear to be closely related, the scent gland auricle of the latter being much smaller and conventionally lobate. Also the pronotum of crassipes is strongly widened anterior to the well-developed transverse impression, whereas in aleocharoides the pronotum is nearly evenly cylindrical with the transverse impression at most scarcely discernible. Macropterous and male specimens must be examined to more adequately place this most anomalous species.

PHYLOGENETIC RELATIONSHIPS OF ISCHNODEMUS

Thus far it has proven impossible to find synapomorphic characters to place all of the species of *Ischnodemus* into a scheme of branching dichotomies. This is particularly true at the base of the cladogram. The scheme illustrated (fig. 42) appears to most accurately reflect not only our state of knowledge but also what is reasonably recoverable from a study of extant species alone. Certainly *Ischnodemus* is an old genus with many of the ancestral "lines" extinct. What does seem to emerge, however, are four reasonably definable clades within each of which a series of grades is evident

I conceive the ancestral *Ischnodemus* to be an only moderately elongate bug with a single spine on each of the fore femora, a completely pruinose head, thorax and scutellum, a wing without prominent color markings, a rounded lobate earlike metathoracic scent gland auricle, an ovipositor that reaches anteriorly to abdominal sternum five, and a labium extending onto the mesosternum.

GROUP I (FIG. 42A): The species which most closely approximate this condition are those which I call the oculatus group and include additional oculatus itself, two Madagascar species, canus and madagascariensis and two African species, diplachne and schoutedeni. These species seem to represent a verv close to the hypothetical plesiomorphic Ischnodemus. All of these species also have a plesiomorphic sperm reservoir with a rounded bulb and straplike wings (diplachne and schoutedeni have the wings curled at the tips, presumably a slightly derived feature). In canus, oculatus, and madagascariensis there is a modification of the fore femora in that madagascariensis has two fore femoral spines present, and in canus the fore femoral spines are absent.

GROUP II: A second group comprises only two species, the Oriental *noctulus* and Southwest Australian *sordidus*, which are discussed in some detail in the zoogeographic section. Both of these species have completely pruinose dorsal surfaces but show synapomorphies in the

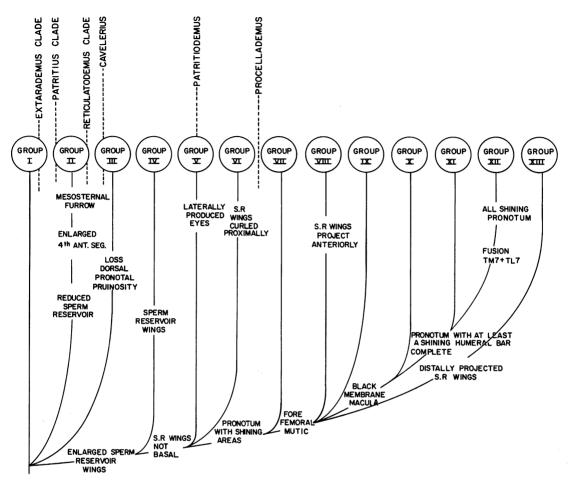


Fig. 42. Cladogram of "Groups" of *Ischnodemus* (continued on consecutive pages). A. Cladogram of species of *Ischnodemus* Group II. B. Cladogram of species of *Ischnodemus* Group III. C. Cladogram of species of *Ischnodemus* Group VII. E. Cladogram of species of *Ischnodemus* Group VIII. E. Cladogram of species of *Ischnodemus* Group X. G. Cladogram of species of *Ischnodemus* Group XI. G. Cladogram of species of *Ischnodemus* Group XII. I. Cladogram of species of *Ischnodemus* Group XIII.

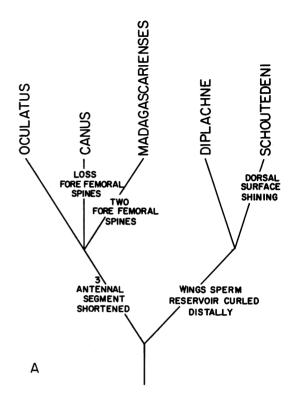
enlarged fourth antennal segment, the presence of a metasternal furrow, and particularly in the nature of the reduction of the sperm reservoir in which the bulb is very small and the wings reduced to extremely small "scales" arising from the extreme distal end of the bulb (sperm reservoir type IX of Slater and Harrington, 1973).

GROUP III (FIG. 42B): The third group is less homogenous than either of the two previously discussed. Essentially it consists of spe-

cies that have retained the pleisomorphic sperm reservoir but have partially or completely lost the dorsal pruinosity so that the head, pronotum, and sometimes even the scutellum are completely shining above. Some of these species also have a hyaline membrane. Within this group, however, I include such species as *genei* and *brincki*, which show incomplete loss of the dorsal pruinosity, and a number of species that have lost the fore femoral spine, including *genei*, *nigrocephalus*, *parathoracicus*, *thor-*

acicus, consobrinus, and crassipes. There is also a tendency for reduction of the sperm reservoir bulb, which is rather marked in such species as brevicornis, sinuatus, dentatus, brincki, and ulugurus. There is no strong apomorphic character that "holds" all of these taxa together. They do seem to form a relatively plesiomorphic group, but show a strong tendency toward pruinosity reduction and a grade tendency toward loss of fore femoral spines.

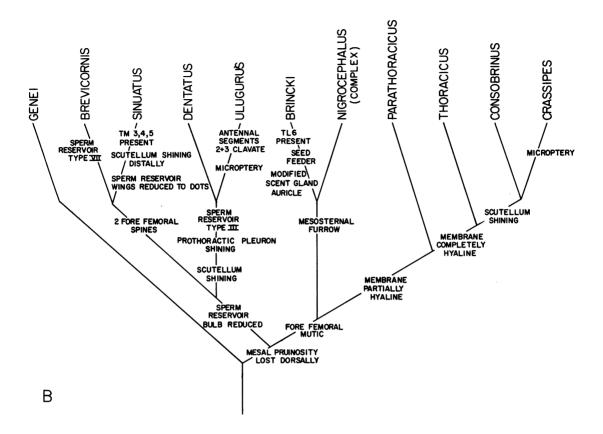
The three groups discussed above constitute the more generalized known species of Ischnodemus. They are all confined to the Eastern Hemisphere. The most important feature separating them from the remaining groups of Ischnodemus (discussed below) is that none of them have a highly derived sperm reservoir (except in the sense of reduction), whereas the remaining groups do have what we consider to be an important synapomorphic character in that the bulb of the sperm reservoir has become enlarged, with extremely wide, flattened, platelike wings (usually curled at the lateral ends) present. It should be noted that there are characters in the nymphal sclerites which may be very useful in eventually establishing better cladistic relationships in the three groups mentioned above, but unfortunately nymphs are known for so few species of these groups that speculation is probably unwarranted at the present time. It is my impression that a number of the major clades in the Blissinae have been derived from common ancestors of species within the above three groups. For example, it is probable that species of Cavelerius are closely related to such Ischnodemus species as genei; that species of Extarademus have arisen early, probably near a common ancestor with the oculatus group; that Patritius and its relatives have arisen from an ancestor not unlike that from which madagascariensis has arisen; that the clade that contains Reticulatodemus and its relatives has arisen from an ancestor similar to such species as Ischnodemus brevicornis and sinuatus. When one realizes that species such as these are rather plesiomorphic elements of the three major clades of the more "apomorphic blissines," one can see the important position of these



groups of *Ischnodemus* as possible sister groups of the more derived genera. Of the *Ischnodemus* species with enlarged platelike sperm reservoir wings there still are taxa which present some problems.

GROUP IV: There are two closely related species in Africa, congoensis and bequaerti, in which the wings of the sperm reservoir, while definitely flattened and platelike, are smaller and placed more basad than they are in other species. This may be the ancestral condition of this sperm reservoir type or it may be a secondary condition. The latter seems more likely, for also in Africa are such species as wittei, ocellaris, and canaliculus in which the sperm reservoir bulb is reduced, the wings have a somewhat "flapped" appearance proximally, and wittei, at least on external appearance, is quite similar to congoensis and bequaerti.

In South America there is a group of species which has been placed previously by us (possibly prematurely) as a separate genus Pa-

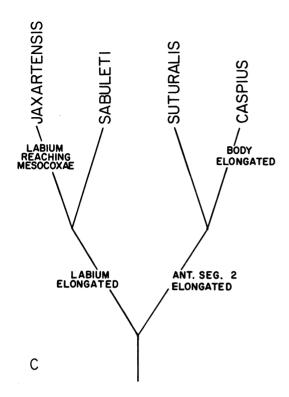


tritiodemus. These Patritiodemus species have very large, well-developed sperm reservoir wings but completely pruinose pronota and somewhat produced eyes. Within the "genus," mutic, simple spined, and enlarged bifid spined femora are present. It is this latter condition which led Slater and Ahmad (1971) to consider a separate genus to be involved. Some species of Patritiodemus are very similar to two other Neotropical species, transitius and proprius, which also have completely pruinose pronota and which are treated here as Group V.

GROUP VI (FIG. 42C): The only other *Ischnodemus* complex or group which has completely pruinose pronota is a Palearctic complex that includes the widespread species *sabuleti*. In these species the fore femora are always mutic, the nymphs have the Tm7 sclerite divided mesally (and Ta7 always present) in species whose nymphs are known, and very

importantly, the sperm reservoir wings are curled "under" along the proximal margins rather than laterally as they are in most other members of these enlarged sperm reservoir-wings-complexes. This seems to be a rather isolated group, and I would suggest has its closest relationships not with extant African elements but probably with the ancestors of the more generalized species of Group V.

All the remaining species of *Ischnodemus*, which constitute the majority of the species, are forms which have at least some shining areas on the dorsal surface of the pronotum. This is a character which is difficult to evaluate, as it occurs in a number of different conditions and appears to show grade levels of organization in different clades of *Ischnodemus* species. Therefore, it is sometimes difficult to separate convergences and parallelisms from cladistic relationships.

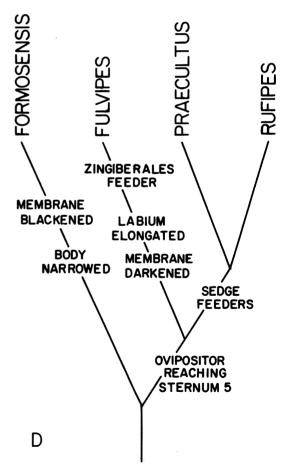


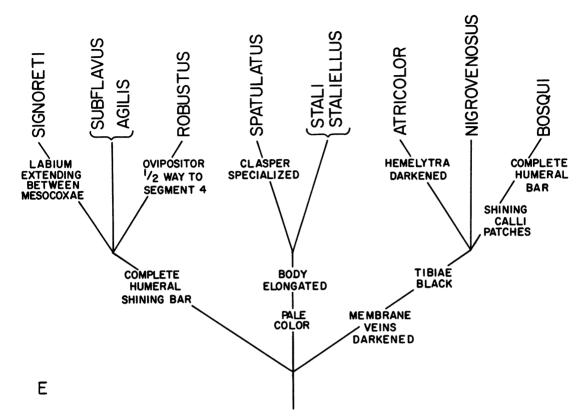
GROUP VII: There are only two species of those discussed in the preceding paragraph that have fore femoral spines. These are the southern African species ochripes and linearis, the former sometimes having four or five spines present on the fore femora. The sperm reservoir wings are very large and subquadrate in these two species, actually more similar to the type of sperm reservoir found in the majority of Neotropical and Nearctic species of Ischnodemus than that found in the other Ethiopian species.

The relationships of the Neotropical and Nearctic species with enlarged sperm reservoir wings are difficult to interpret. In all of them the sperm reservoir wings are large, relatively subquadrate similar to the condition found in the African species *ochripes* and *linearis*. They do appear to form a single phyletic lineage, although I have not been able to find a synapomorphic character of substance to separate them from other *Ischnodemus*. Within the Neotropics, however, several groups are present.

GROUP VIII (FIG. 42D): The fulvipes group seems to be a well-defined complex. In this group the lateral edges of the sperm reservoir wings project anteriorly, the eyes are produced laterad, and the clasper shape is rather distinctive. It is interesting to note that none of the species whose feeding habits are known feed upon grasses but upon other monocots such as sedges, zingiberales, and others. In addition to fulvipes itself, formosensis, praecultus, and rufipes belong to this group.

GROUP IX (FIG. 42E): The stali group may not be a monophyletic assemblage. I have included here such species as stali, staliellus, spatulatus, atricolor, nigrovenosus, bosqi, signoreti, agilis, subflavus, and possibly robustus. Most of the species are relatively elongate, pale





testaceous forms without a definite black macula on the membrane of the fore wing. They include species with a complete shining bar across the humeri such as *subflavus*, *signoreti*, and *agilis* and those in which there are three definite spots present such as *stali*, *staliellus*, and *spatulatus*. These species seem to be grass feeders and to be chiefly savanna insects.

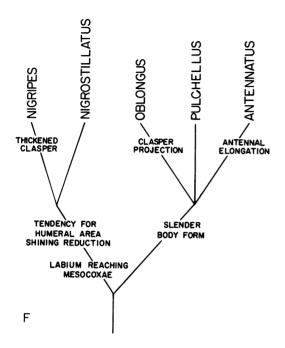
In the Neotropics there are a number of species that have a large black macula on the membrane. These species might seem to be held together by a superficial character. I believe there are two distinct phyletic lines represented.

GROUP X (FIG. 42F): The first of these maculate complexes I call the *oblongus* group. It contains five species. *Ischnodemus oblongus*, antennatus, and pulchellus constitute a very closely related complex and are unquestionably sister species. The other two species, nigrostillatus and nigripes, are somewhat more isolated, and it is possible that they represent a

more ancestral condition, since the humeral shining bar is often very much "reduced" and the labium extends to the mesocoxae. This group has an apparently synapomorphic character in the shape of the clasper and I do not argue strongly for the unity of the group. However, it is obvious that this group is quite distinct from the following which contains a number of species with a black membranal macula.

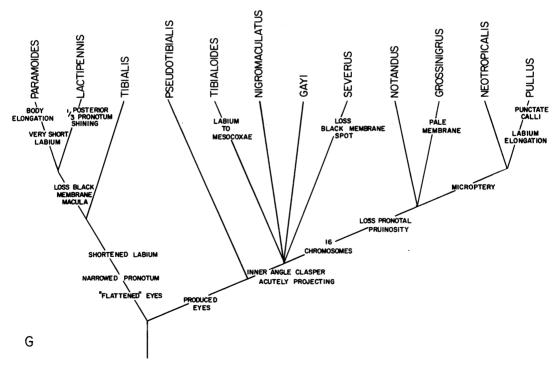
GROUP XI (FIG. 42G): This second maculate group I call the *tibialis* group. These species are characterized by a quite different body shape from the species of Group X, being for the most part elongate and slender, having a differently shaped clasper and laterally tapering wings to the sperm reservoir. They have a complete shining bar across the humeral area of the pronotum. To this complex belong several species which do not have the black macula. Thus, such species as *paramoides* and *lactipennis*, which lack the black macula, indicate that

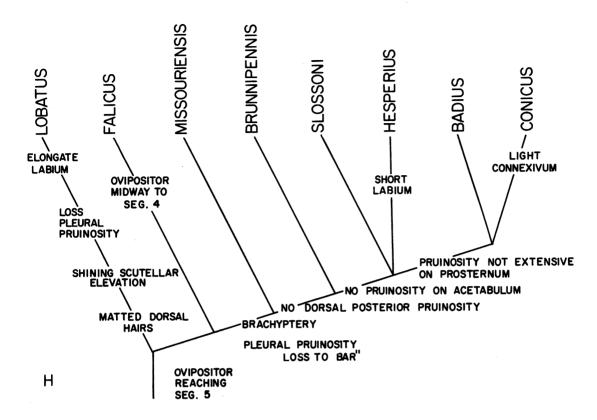
the other features used to hold the group together are of importance. The species belong-



ing to this complex, in addition to tibialis, are such closely related species as pseudotibialis, tibialoides and nigromaculatus, and somewhat more isolated species such as gayi, severus, paramoides, and lactipennis. It is also evident that within this group there has been anagenetic grade loss of the pronotal pruinosity, and thus such dorsally shining species as notandus, grossinigrus, and probably neotropicalis and pullus also belong here. These last four species are completely shining above but, particularly the first two, show close resemblance in most other characteristics to the other members of the tibialis group.

GROUP XII (FIG. 42H): The other *Ischnodemus* group found in the New World is essentially Nearctic. This constitutes the *falicus* group and makes up the great majority of the species found in North America. It includes *lobatus*, *slossoni*, *falicus*, *missouriensis*, *brunnipennis*, *hesperius*, *badius*, and *conicus*. These species are all characterized by dorsally shining pronota, for the most part elongate ovipositors and a fusion in the nymphal sclerites of Tm7 and Tl7. There seems to be no question but

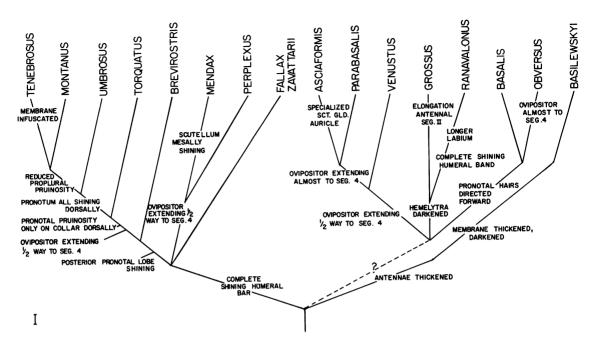




that these species constitute a very distinct, isolated group, the difficulty being in trying to establish its affinities. It is tempting to believe that the *falicus* group has arisen from a common ancestor with some of the most advanced species of the *tibialis* complex, such as *notandus* and *pullus*.

GROUP XIII (FIG. 42I): With the exception of ochripes and linearis the African species with enlarged sperm reservoir wings have these wings "shifted" distally. This African complex, which is "held together" by the synapomorphic character of distally placed sperm reservoir wings, constitutes a reasonably closely related clade. However, within this complex there are, of course, subgroups. There is a subgroup held together by the presence of a complete shining humeral bar, but the species are rather dissimilar and again show a grade anagenesis to a complete dorsal shining pronotal surface. This subgroup is composed of mendax, perplexus, zavattarii, fallax, brevirostris, torquatus, umbrosus, montanus, and tenebrosus. Such species as grossus, ranavalonus, basalis, and obversus form a closely related subgroup which probably also should include venustus, parabasalis, and asciaformis. This latter subgroup is the more plesiomorphic as indicated by the presence of three well-separated shining humeral spots.

In summary, the genus *Ischnodemus* appears to be a complex containing a number of African and Madagascan species that show many plesiomorphic conditions; a presumed clade in the Neotropics divisible into several distinct groups and a highly apomorphic African group subdivisible into two primary subunits. It is possible that a dichotomous cladogram may be produced of all these Ischnodemus species when sophisticated information on nymphs. host plants, chromosome numbers, and so forth, is available but I am not optimistic that this will be the case. It does not seem to me likely, as previously discussed, that complex speciation processes, such as are present in Ischnodemus, plus obvious grade and anagene-



tic changes in several phyletic lines, are capable of being reduced to simple branching dichotomies.

KEY TO SPECIES OF ISCHNODEMUS1

- la. Fore femora mutic14
- Vertex of head pruinose at least narrowly along midline, usually head shining dorsally only on tylus and frequently as a small dash or L-shaped mark adjacent to each ocellus (but ochripes sometimes with more extensive shining area present) ...3
- 2a. Vertex of head shining, at least mesally .. 8
- 3. Pronotum with conspicuous shining areas at least posteriorly across humeral region (figs. 43, 44, 45, 46, 47, 48, 49, 51) .4
- 4. Membrane of fore wing with a large dark brown to black central area strongly contrasting with pale apical portion; mesosternum lacking a deep troughlike median longitudinal furrow; calli not completely

'inambitiosus Buchanan White is unknown, the type apparently lost. It perhaps belongs to the tibialis complex.

- 4a. Membrane uniformly pale buffy yellow to sordid white with only veins darkened; mesosternum with a deep troughlike median longitudinal furrow; area of calli subtriangular, mostly shining (fig. 43); fore femora frequently with four spines present (South Africa) ochripes Stål
- 5. Fore femora with a pair of divergent spines present (Madagascar).....
- ... madagascariensis Slater and Wilcox 5a. Fore femora with a single spine present ... 6

- 7. Femora unicolorous sordid yellow; eyes large, protrudent, occupying major portion of lateral head surface (fig. 6D) (Madagascar).....oculatus Slater
- 7a. Femora dusky brown, becoming lighter distally; eyes smaller, not occupying major portion of lateral head surface (fig. 6C) (Africa).....

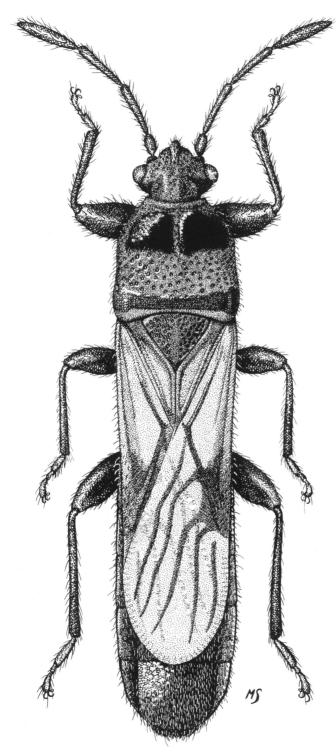


Fig. 43. Ischnodemus bosqi, dorsal view.

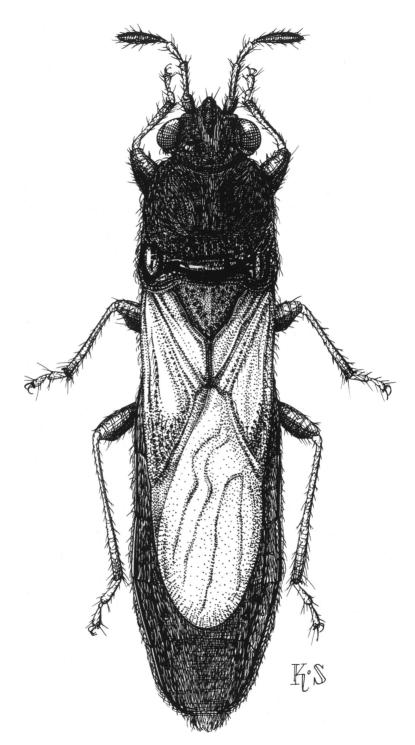


Fig. 44. Ischnodemus brevicornis, dorsal view.

8a.	shining, either patterned with pruinosity in	14a.	least posterior lobe pruinose, lacking shin-
	collar region (fig. 4N), in area of trans-		ing bar or spots (fig. 50)88
	verse impression and narrowly along pos-	15	Dorsal surface of pronotum usually with dis-
	terior margin (fig. 4J), or pronotum	15.	tinct pruinosity pattern i.e., the combina-
	completely pruinose except for shining		tion of anterior collar area always
			distinctly shining mesally, a pair of mes-
	areas in region of calli and/or as a sub- basal transverse bar across humeral area		ally separated very large shining calli
0	(fig. 43)		patches present and a broad complete
9.	Total length labium subequal to width of		shining bar across humeral area (fig. 4G)
	head across eyes (Africa) (fig. 44)	15.	
0-	brevicornis Stål	ısa.	Pronotum dorsally shining, or pruinose or
9a.	Total length labium greater than width of		patterned with shining and pruinose areas
10	head across eyes		but not in combination shown above, if
	Fore femora with a single spine present11		pattern similar then anterior collar area
IUa.	Fore femora with a conspicuous bifid or dou-		completely pruinose (figs. 4F, 43) or large
	ble spine present (fig. 7-I) (Orient)		shining calli patches contiguous at meson
	sinuatus Slater, Ashlock, and Wilcox		and not separated by a median pruinose
11.	Scutellum completely pruinose (figs. 43, 51);		strip (fig. 4J) (genei)
	legs completely or in large part dark	16.	Labium very elongate, reaching between or
	blackish brown (Africa)		beyond mesocoxae, second labial segment
	bequaerti Slater		attaining fore coxae; mesosternum with a
lla.	Scutellum completely shining or subshining;		deep troughlike median longitudinal fur-
	legs reddish brown or bright yellow		row; antennae black (Africa)
			brincki Slater
12.	Labium relatively elongate, reaching onto an-	16a.	Labium relatively short, extending only be-
	terior portion of mesosternum, second seg-		tween fore coxae, second labial segment
	ment surpassing base of head; antennal		at most reaching base of head; mesoster-
	segments 2 and 3 short, clavate, their		num lacking a deep median longitudinal
	combined length less than interocular dis-		furrow; antennae reddish brown to sordid
	tance (.3034 mm.); anterior pronotal lobe		yellow17
	black, lacking contrasting light coloration	17.	Lateral pruinosity of anterior prothoracic lobe
	in collar area (Africa)		extending dorsally well beyond lateral
	Scudder		shining bar and onto dorsal surface of pro-
12a.	Labium shorter, not reaching beyond fore		notum (fig. 4M); pruinose area present
	coxae, second segment remote from base		dorsally on head behind eyes and along
	of head; antennae with segments 2 and 3		area of juga adjacent to eyes; anterior pro-
	longer, terete, their combined length more		notal lobe mesally somewhat impressed
	than interocular distance (.5040 mm.);		but lacking a deep groove (Africa
	anterior pronotal lobe black with contrast-		ocellaris Slater and Harrington
	ing yellowish brown band present across	17a.	Lateral pruinosity of anterior prothoracic lobe
	collar area (North Africa)		not extending onto dorsal surface of pro-
	dentatus Wagner		notum beyond lateral shining bar (fig.
13.	Tylus reaching more than halfway to distal		50); head dorsally completely shining,
	end of antennal segment 1; length of la-		lacking distinct pruinose areas behind and
	bium greater than length of pronotum and		before eyes; anterior pronotal lobe with a
	more than twice interocular distance (Af-		deep, sharply defined median groove (Af-
	rica)schoutedeni Slater		rica)
13a.	Tylus not reaching more than halfway to dis-	18.	Dorsal surface of pronotum shining, with at
	tal end of antennal segment 1; total length		most narrow pruinose area along posterior
	labium less than length pronotum and less		margin or behind collar area, or narrowly
	than twice interocular distance (Africa)		along lateral margins of anterior lobe
			19
14.	Pronotum with shining areas present at least	18a.	Dorsal surface of pronotum in large part pru-
	on posterior lobe as shining bar or spots		inose, or if largely shining then a com-
	(figs. 43, 46, 48, 49, 51)		plete transverse pruinose band present

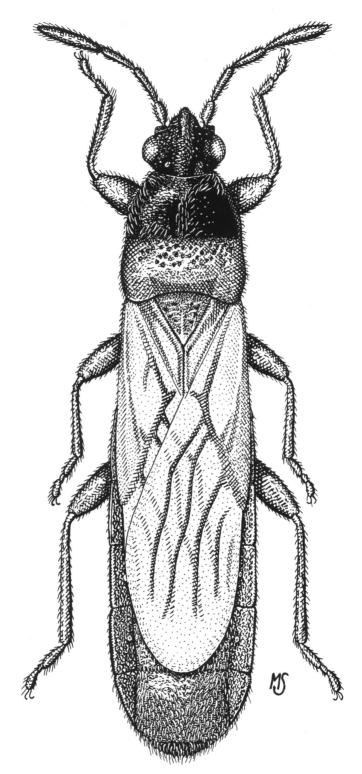


Fig. 45. Ischnodemus brunnipennis, dorsal view.

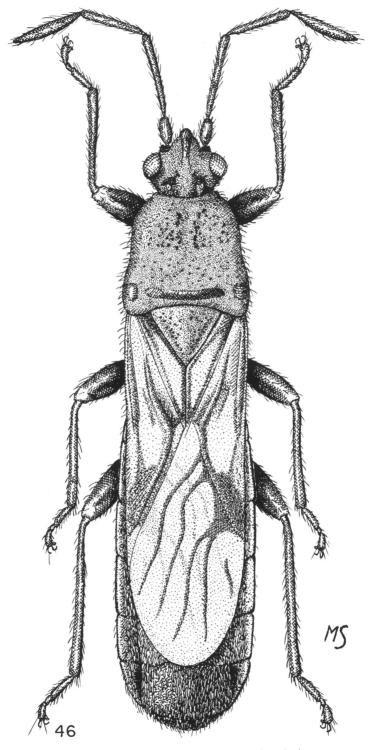


Fig. 46. Ischnodemus nigrovenosus, dorsal view.

additional material will become available.

	across transverse impression (fig. 4H)	25.	Labium barely attaining fore coxae, second segment not reaching base of head (Af-
19.	Prothorax nearly completely shining above and below, pruinosity limited to at most a	25a	rica)
	very narrow strip posteriorly on dorsum	254.	ond segment surpassing base of head by
	and a narrow strip anteriorly on sternum		half its length (South America)
	20		pullus Slater and Wilcox
19a.	Prothorax with extensive pruinosity present at	26.	Fore coxal cavities open (Orient)1 (see Slater,
	least on sternum between and anterior to		Ashlock, and Wilcox, 1969)27
	fore coxae (fig. 5T)	26a.	Fore coxal cavities closed
20.	Fore wing variegated; always macropterous,	27.	Membrane nearly uniformly smoky brown
	membrane well developed with a large		with diffuse white areas present at base
	conspicuous central macula present; dark-		and adjacent to distal end of corium; an-
	ened patches present along apical corial		tennal segment two dark brown to black;
	margins and apex of claval commissure		first labial segment attaining or nearly at-
	(South America)		taining base of head
20	notandus Slater and Wilcox	27-	fumidus Slater, Ashlock, and Wilcox
20a.	Fore wing lacking a conspicuous light and	2/a.	Membrane except veins completely white or
	dark variegated color pattern; frequently		hyaline; antennal segment two pale testa- ceous; first labial segment remote from
	brachypterous or micropterous species		base of head
21	with membrane greatly reduced21 Scutellum completely pruinose (fig. 5L),		ambiguus Slater, Ashlock, and Wilcox
21.	lacking a central shining area, or if ap-	28.	Labium elongate, reaching between meso-
	pearing somewhat shining near apex, then		coxae, or if labium slightly shorter then
	anterior pronotal lobe with a deep conspic-		head and pronotom covered with thick
	uous median groove		matted hairs
21a.	Scutellum either completely shining or at	28a.	Labium not reaching mesocoxae30
	least with a shining central area (fig. 5I, J,	29.	Head and pronotum densely clothed with
	M)24		thick matted, decumbent hairs, membrane
22.	Head and quadrate calli patches on anterior		opaque white (veins slightly darker);
	pronotal lobe concolorous (or if pronotal		scutellum with shining median elevation
	patches lighter than part of head, then		on distal third (fig. 5J) (North America)
	vertex of head also lighter); connexivum	200	
222	dark or yellow only on lateral areas23 Head darker than and contrasting with large	29a.	Head and pronotum nearly glabrous, lacking conspicuous matted hairs; membrane
22a.	quadrate patches on anterior pronotal lobe;		smoky, almost entirely infuscated;
	connexivum usually entirely yellow (North		scutellum completely pruinose (fig. 51)
	America) badius Van Duzee		(Orient)
23.	Relatively small species, & barely over 5		fumidus Slater, Ashlock, and Wilcox
	mm.; pronotum dark red-brown to black;	30.	Dorsal surface of pronotum with pruinosity
	length second antennal segment less than		present either anteriorly, laterally or poste-
	interocular space (South America) (fig.		riorly31
	47) neotropicalis Slater and Wilcox	30a.	Dorsal surface of pronotum completely shin-
23a.	Larger species, & over 6 mm; pronotum light		ing
	reddish brown; length second antennal	31.	Dorsal surface of pronotum lacking pruinose
	segment usually more than interocular		areas on anterior lobe (North America)
	space (North America)	310	Dorsal surface of pronotum with narrow pru-
24	Pronotum with conspicuous long upright	Jia.	inose areas laterally on anterior lobe and/
27.	hairs, those on anterolateral area directed		mose areas faterany on affection robe and
	forward; large species, total length	17.	fumidus and I. ambiguus really do not belong in
	8.64-9.70 mm. (South America)		demus as earlier defined. Their relationship may be
	grossinigrus Slater and Wilcox		apodemus. There is also an undescribed \(\varphi \) micropter
24a.	Pronotum glabrous or sparsely clothed with	from E	turma involved with a pruinose pronotum. For the
	short hairs, these directed posteriorly;	-	I prefer not to erect an additional genus in the hope
	smaller species under 7 mm 25	addition	aal material will become available

smaller species, under 7 mm......25

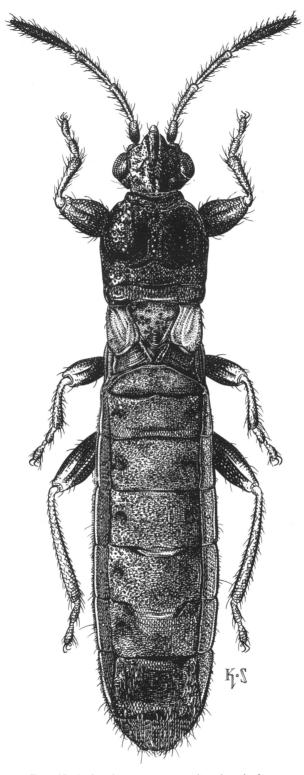


Fig. 47. Ischnodemus neotropicalis, dorsal view.

	or a pruinose transverse band posterior to shining collar area (fig. 4N)32	38a.	Scutellum nearly completely pruinose with at most a narrow shining elevation on distal
32.	Labium relatively short, not attaining anterior margins of fore coxae (Africa)	30	third (North America)
	torquatus Slater and Harrington	39.	ing at least partially over outer (lateral)
32a.	Labium longer, extending between fore coxae		surface of acetabula (fig. 5S), or if pru-
	but not reaching anterior margin of meso-		inose area appearing less extensive
	sternum (Africa)		(falicus) then macropterus species with la-
	umbrosus Slater and Harrington		bium short and second segment remote
33.	Head black, strongly contrasting with bright	20	from base of head40
	tan pronotum (Orient)nigrocephalus	39a.	Ventral prothoracic pruinosity confined to
220			median portion of prosternum and bor- dered by sinuous longitudinal lines at level
ssa.	Head and at least anterior pronotal lobe uniformly black		of lateral margins of coxal cavities (fig.
34	Membrane almost completely fuscous with		5T)41
٥,,	only apical portion pale (Africa)	40.	Species always macropterous or submacrop-
	tenebrosus Slater and Harrington		terous; ratio antennal segment 2 to 4 usu-
34a.	Membrane pale or hyaline with at most a		ally exceeding 0.78; antennal segments
	small dark spot adjacent to apical corial		1-4 usually concolorous, dark, often
	margin and/or with veins sometimes		wholly black; pronotal transverse impres-
	darker		sion dark and concolorous with anterior
35.	Membrane of hemelytra chiefly or entirely		lobe, both contrasting with lighter poste-
	hyaline, transparent, at most opaque only		rior pronotal lobe (North America)
25.	anteriorly between coria	40-	falicus (Say)
ээа.	Membrane of hemelytra entirely opaque white, often with darkened veins and/or	40a.	Species usually brachypterous or micro-
	spots		pterous; ratio of antennal segment 2 to 4 usually less than 0.78; antennal segments
36.	Membrane of hemelytra with anterior area		1-4 not of a single color (the change in
	adjacent to corium conspicuously opaque,		color may be abrupt or gradual and nearly
	strongly contrasting with hyaline texture		imperceptible, but segments 1 and 4 never
	of remainder of membrane (Africa)		concolorous); posterior pronotal lobe and
	parathoracicus Slater and Harrington		transverse impression usually light reddish
36a.	Membrane of hemelytra completely hyaline		brown contrasting with nearly black ante-
27	37		rior lobe (North America) (fig. 45)
37.	Scutellum pruinose except for extreme poste-	4.1	brunnipennis (Germar)
	rior end (fig. 5K); labium extending well	41.	Length of labium less than twice as great as
	onto mesosternum, second segment ex- ceeding base of head by more than half its		interocular space; veins of hemelytra pale
	length; mesosternum with a distinct me-		and indistinct, not contrasting with ground color; brachyptery type "pointed" (fig.
	dian furrow (Orient, Africa)		7E) (North America)
	thoracicus (Distant)		hesperius Parshley
37a.	Scutellum largely shining, pruinose only nar-	41a.	Labium more than twice as long as interocu-
	rowly along anterior and lateral margins;		lar space; veins of hemelytra indistinct,
	labium shorter, at most reaching anterior		elevated, usually light brown contrasting
	margin of mesosternum, second segment		with opaque white ground color; brachy-
	not exceeding base of head by half its		ptery type generally "narrowly lobate"
	length; mesosternum lacking a distinct me-		(fig. 7I) (North America)
	dian furrow (Orient) (Distant)	42	
38.	Scutellum usually almost completely shining,	42.	Pronotal calli with central area completely shining without interspersed dots of shin-
	narrowly pruinose across base, at least		ing and pruinose on calli (fig. 43) 43
	with a broad T-shaped shining median ele-	42a.	Pronotal calli entirely pruinose or with at
	vation (fig. 5-I) (Africa)		most intermixed irregular shining areas
	montanus Slater and Harrington		and pruinose areas interspersed, calli

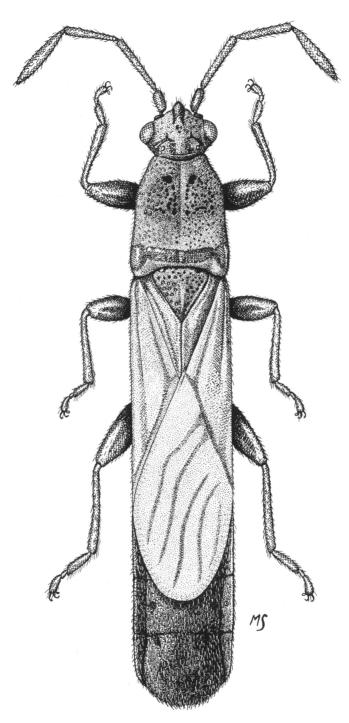


Fig. 48. Ischnodemus paramoides, dorsal view.

	never completely shining (fig. 4A-C, I).		femora pale
		49.	Labium short, at most slightly exceeding fore
43.	Shining pronotal calli area large, covering		coxae (South America) tibialis Stå
	most of anterior lobe, sometimes complete	49a.	Labium longer, reaching to or exceeding base
	or very narrowly separated across midline		of prosternum50
	(fig. 4E, J)44	50.	Fore femora very dark brown to nearly black
43a.	Shining pronotal calli area relatively small,		strongly contrasting with pale tibiae
	trianguloid, well separated by wide pru-		(South America)
	inose mesal area (fig. 4G)45		nigromaculatus Slater and Wilcon
44	Scutellum and vertex of head mesally pru-	50a.	Fore femora and tibiae nearly uniformly
	inose; membrane with veins dark brown,		bright yellow, concolorous; femora a
	contrasting with white ground color, a dif-		most very slightly infuscated, neve
	fuse brown spot present adjacent to apex		strongly contrasting with tibial coloration
	of corium (South America) (fig. 43)		5
	bosqi Slater and Wilcox	51	Labium relatively elongate, extending to o
110	Scutellum with broad shining median eleva-	51.	almost to mesocoxae, third segment ex
44a.	tion on distal two-thirds, head completely		ceeding posterior margin of prosternum
			(South America)
	shining; membrane unicolorous white (Pa-		tibialoides Slater and Wilcon
4.5	learctic)	510	Labium shorter, barely reaching onto meso
43.	Labium very elongate, reaching between or	Jia.	sternum, third segment not attaining poste
	beyond mesocoxae, second segment at-		
	taining fore coxae (S. Africa)		•
4.5	brincki Slater	•	America)
45a.	Labium short, reaching between fore coxae,	50	pseudotibialis Slater and Wilcon
	second segment just attaining base of head	52.	Hemelytra nearly uniformly dark chocolate
	(Africa)brevirostris Bergroth		brown, becoming narrowly light testa
46.	Pronotum in large part pruinose but with a		ceous along lateral corial margins and a
	shining glabrous area complete sub-basally		extreme base of hemelytra (Madagascar
	across humeri (fig. 4C, I), not divided		
	into three separate patches ¹ 47	52a.	Hemelytra with membrane white to smok
46a.	Basal area of pronotum with three distinct		gray, sometimes with veins light to dark
	shining regions, i.e., pruinose areas pres-		brown and/or a small diffuse brown spo
	ent separating central elongate patch from		present adjacent to apical corial margin.
	oval lateral patches (fig. 46)66		
47.	Fore wing variegated, membrane with a large	53.	Pronotum with entire basal one third to on
	conspicuous dark central macula which		half and anterior collar area strongly shin
	strongly contrasts with light ground color		ing, contrasting with remaining pruinos
	(in brachypters membrane macula some-		areas (fig. 4-I) (South America)
	times light brown and diffuse)48		lactipennis Slater and Wilcon
47a.	Fore wing usually not variegated, membrane	53a.	Pronotum almost entirely pruinose except nar
	lacking contrasting central macula, at most		row transverse band across humeri (fig
	with veins darkened and/or small spot		48)54
	present adjacent to apical corial margin, or	54.	Tibiae and femora nearly uniformly dark
	membrane nearly uniformly testaceous,		brown to black, (or if appearing lighte
	tan, or smoky brown52		then unusually elongate slender species
48.	Tibiae completely dark, concolorous with		total length more than six times width
	dark femora (South America)		(Neotropical)5
	gayi (Spinola)	54a.	Tibiae and usually femora pale to light brown
48a.	Tibiae pale, either strongly contrasting with		
	dark femora, or with both tibiae and	55.	Labium short, barely attaining fore coxae
			pronotum lacking a transverse impression
¹Al	though this character is usually constant and reliable		(South America) (fig. 48)
some s	pecies such as the African fallax group show consid-		paramoides Slater and Wilcon
erable	variation with several conditions present.	55a.	Labium longer, reaching onto mesosternum

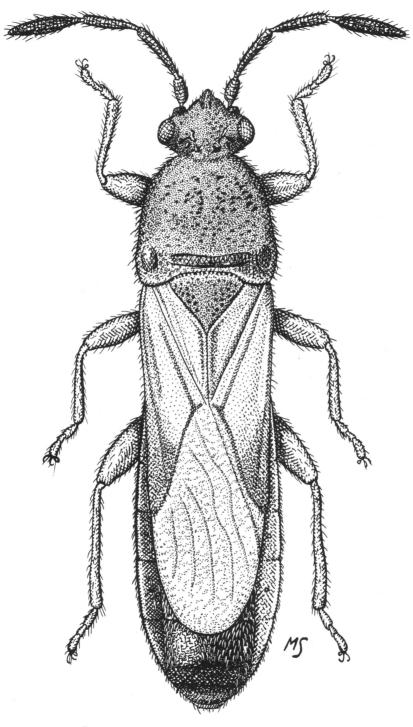


Fig. 49. Ischnodemus praecultus, dorsal view.

	pronotum with broad, shallow complete transverse impression (South America)	ground color (South America)	
56.		64a. Head relatively longer and pronotum broadd [ratio head] length to width greater than of	or
56a.	orly onto mesosternum	nearly equal to ratio pronotal length t width (78:75)]; antennae relatively slende	r;
57.	fore coxae	veins of membrane concolorous wit ground color (South America)	
٥,,	ond segment nearly attaining fore coxae	subflavus Slater and Wilco	X
57a.	(South America) signoreti Berg Labium shorter, remote from mesocoxae, sec-	65. Second antennal segment usually longer that interocular space, but never more that	
	ond segment not reaching fore coxae (Af-	1.25 times as long, sometimes with inte ocular space greater than length of secon	r-
58.	rica)parabasalis Slater Pronotal hairs directed anteriorly forward, at	antennal segment (Africa)	
	least on anterolateral area (Africa)	65a. Second antennal segment shorter, ratio	
58a.	Pronotal hairs directed posteriorly59	second segment to interocular space	
59.	Membrane testaceous with veins a contrasting	greater than 1.25 (Africa)	
	dark brown; apical corial margin lacking a brown spot; antennae relatively broad,	66. Membrane bearing a large black or dar	
	robust (North America) Blatchley	brown macula which contrasts strikingly with pale basal and apical areas 6	
59a.	Veins of membrane only slightly darker than	66a. Membrane unicolorous or nearly so, most fro	
	ground color and/or a diffuse brown spot present adjacent to apical corial margin;	quently pale testaceous but sometime dark smoky gray to nearly black, at mo	
	antennae slender, terete60	with differentiated spots adjacent to apic	al
60.	Labium relatively short, at most barely attain- ing anterior margin of fore coxae, second	corial margin, or with veins only contrastingly dark	n- 73
	segment not surpassing base of head61	67. Labium relatively short, not attaining anterior	
60a.	Labium longer, reaching at least between fore coxae, sometimes extending onto anterior	margin of fore coxae; clavus and corium nearly unicolorous light reddish brow	
	portion of mesosternum, second segment	(Africa) basilewskyi Slate	
61	surpassing base of head63 Mesal area of head vertex pruinose (Africa)	67a. Labium longer, at least reaching between for coxae; clavus and corium usually with	
01.	fallax Slater and Harrington	some light areas present6	8
	Mesal area of head vertex shining62 Scutellum completely pruinose (fig. 5L); eyes	68. Labium relatively elongate, extending nearl to mesocoxae, always reaching at lea	
02.	small relative to head (Africa)	midway from anterior mesosternum ma	
620	Southly with longitudinal chining area	gin to anterior margin of mesocoxa	
02a.	Scutellum with longitudinal shining area along median elevation (fig. 5I, J); eyes	(Western Hemisphere) (fig. 51)	
	relatively large (Africa)	68a. Labium shorter, sometimes extending ont	
63.	Membrane unicolorous white (South Amer-	mesosternum but always extending les than halfway from anterior margin of	
630	ica)	mesosternum to anterior margin of meso	0-
03a.	with a small diffuse brown spot present	coxae	
	adjacent to apical corial margin (Africa)65	band and frequently entirely black wit	th
64.	Head relatively short and pronotum narrow	When this character was used in the Slater and Wilco	ЭX
	[ratio of head length to width considerably less than ratio of pronotal length to width	(1969) key to species, there was an error in couplet 26. The	
	(62:88)]; entennes thick and start, voins	couplet should read: "Head relatively longer and pronotus	m

broader, i.e., ratio of head length. . . . " rather than ratio

of pronotal length.

(62:88)]; antennae thick and stout; veins

of membrane darker than membrane

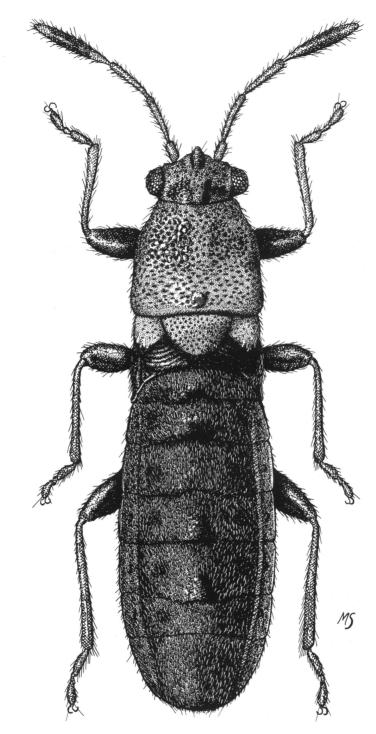


Fig. 50. Ischnodemus proprius, dorsal view.

	the exception of basal and distal ends; second antennal segment less than 1 1/2		Tibiae and femora nearly uniformly testaceous to light brown
	times as long as interocular distance	74.	Dorsal coloration including membrane a nearly uniformly dark chocolate brown to
6 0-	(South America)oblongus (Fabricius)		black; veins of membrane dark and not
оча.	All femora pale yellow or reddish, con-		
	colorous with pale tibiae; second antennal		strongly contrasting with dark coloration of remainder of membrane (South Amer-
	segment more or less than 1 1/2 times as		ica)atricolor Berg
	long as interocular distance, but if femora	74-	
	appear somewhat infuscated, then second	/4a.	Hemelytra in large part white to testaceous
	antennal segment more than 1 1/2 times as		with some dark markings; veins of mem-
	long as interocular distance70		brane prominently dark brown, strongly
70.	Labium relatively short, usually not exceed-		contrasting with pale coloration of re-
	ing posterior margin of fore coxae and not		mainder of membrane (South America)
	or barely attaining posterior margin of		(fig. 46)
	prosternum		nigrovenosus Slater and Wilcox
70a.	Labium usually extending onto anterior por-	75.	Pronotal hairs directed anteriorly forward, at
	tion of mesosternum, always exceeding		least on anterolateral area
	posterior margin of fore coxae and attain-	75a.	Pronotal hairs directed backward or upright
	ing posterior margin of prosternum72		77
71.	Second antennal segment more than 1 1/2	76.	Posterior margin of \circ fifth abdominal ster-
	times as long as interocular distance; com-		num (fourth visible) sinuate, segment con-
	bined length of second and third antennal		spicuously narrowed medially where
	segments more than 1 1/2 times width of		ovipositor sclerites extend anteriorly (fig.
	pronotum; antennae relatively slender		8G); pronotum usually tawny to pale red-
	(South America)		dish brown on posterior lobe, transverse
	antennatus Slater and Wilcox		impression and narrowly along collar; legs
71a.	Second antennal segment less than 1 1/2		usually pale yellow to tawny (Africa)
	times as long as interocular distance; com-		obversus Slater and Harrington
	bined length of second and third antennal	76a.	Posterior margin of ♀ abdominal sternum 5
	segments slightly greater than pronotal		straight or nearly so, segment not noticea-
	width, always less than 1 1/4 times width		bly narrowed medially (fig. 8E); pronotum
	of pronotum (South America)		dark blackish brown, only slightly paler
	pulchellus Slater and Wilcox		across posterior lobe, no light area present
72.	Labium just attaining posterior margin of		along collar; legs usually dark brown (Af-
	prosternum or at most with only distal half		rica) basalis Walker
	of fourth segment exceeding base of pro-	77.	Fore wing nearly unicolorous pale testaceous
	sternum; second antennal segment less		to opaque white (South America) 78
	than 1 1/2 times as long as interocular	77 a .	Fore wing not unicolorous white, with at
	distance; terminal third of membrane and		least veins darkened and/or a dark spot
	greater portion of clavus and corium		adjacent to corial apex
	white, not suffused with smoky brown	78.	Length of second antennal segment greater
	(South America)		than width of head across eyes; antennae
	formosensis Slater and Wilcox		pale tan to yellow with at most first seg-
72a.	Labium relatively more elongate with at least		ment darkened (South America)
	entire fourth labial segment exceeding	=0	stali (Signoret)
	base of prosternum; second antennal seg-	78a.	Length of second antennal segment less than
	ment at least (and usually more than) 1		width of head across eyes; antennae with
	1/2 times as long as interocular distance;		at least segments 1 and 2 dark brown to
	entire membrane, clavus and corium fre-		black (South America)
	quently suffused with dull smoky brown	70	staliellus Slater and Wilcox
	coloration (Western Hemisphere) (fig. 51)	79.	Labium relatively elongate, reaching well
72	Tibing and famore uniformly dork character		onto mesosternum, third segment attaining
13.	Tibiae and femora uniformly dark chocolate		or nearly attaining posterior margin of
	brown to nearly black		prosternum80

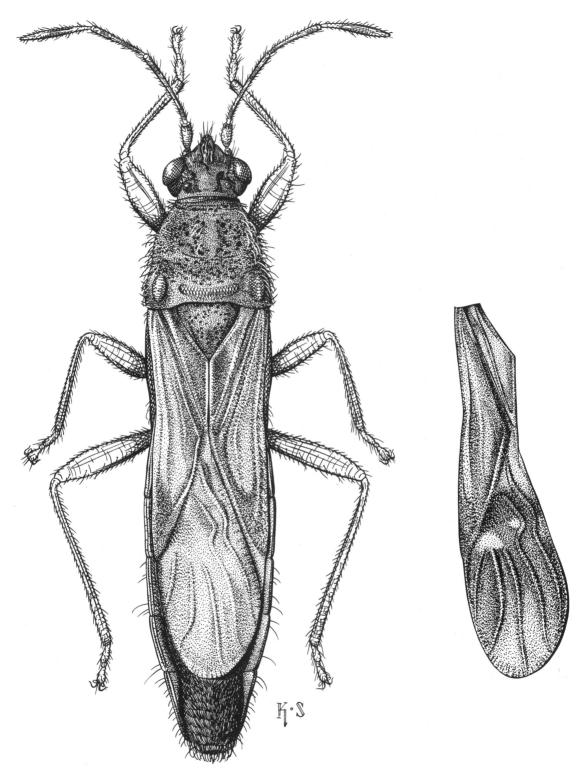


Fig. 51. Ischnodemus fulvipes, dorsal view.

79a.	Labium shorter, at most barely reaching onto mesosternum, third segment remote from
80.	posterior margin to prosternum82 Fore wing with ground color testaceous, veins of clavus and corium and distal third of corium brown (Africa)
	Fore wing in large part smoky or dark brown, membrane always nearly completely infuscated
	Scent gland auricle curving forward, "club-like" (fig. 10C); body wall with shelflike protrusion projecting out over scent gland area; antennal segment 2 shorter than segment 4 (Africa)
81a.	Scent gland auricle conventionally shaped; body wall not protruding out over scent gland area; antennal segment 2 longer than segment 4 (Western Hemisphere) (fig. 51)
82.	Length of second antennal segment usually somewhat greater than interocular space, but never more than 1 1/4 times as great, sometimes with interocular space greater than length of second antennal segment.
82a.	Second antennal segment longer, ratio length second segment to interocular space always greater than 1 1/485
	Membrane creamy white but with veins contrasting dark brown ¹ ; antennae dark redbrown to black (except at distal ends of segments 2-4) (South America)
83a.	Membrane white with veins at most only slightly darker, not strongly contrasting with ground color ¹ ; at least antennal segments 2 and 3 bright yellow-brown to light red-brown
84.	Labial length less than 1 1/4 times head width; ratio antennal segment 2/segment 4 more than 0.63 (Africa)

'This character is variable in praecultus with some specimens having dark brown veins. Total length can be used to separate praecultus from spatulatus. I. praecultus ranges from δ 4.44-5.64, \Im 5.58-6.48 and spatulatus δ 5.94-6.60, \Im 6.80-8.04. The labium is usually shorter in spatulatus, at most barely exceeding the fore coxae. In praecultus the labium extends well onto the mesosternum with the distal end of the third segment reaching the fore coxae, but in some specimens the labium barely attains the anterior margin.

- 85. Ratio antennal segment 2/interocular space more than 1.60; posterior margin of φ abdominal sternum 5 nearly straight, not noticeably abruptly narrowed medially, only tapering slightly inward from lateral margins (fig. 8F) (Africa) .grossus Slater
- 85a. Ratio antennal segment 2/interocular space less than 1.60; posterior margin of φ abdominal sternum 5 sinuate, segment abruptly narrowed medially at anterior extension of ovipositor (fig. 8G) 86
- 86. Dark coloration extensively developed on membrane (Africa) venustus Slater
- 87. Ratio total length labium/length antennal segment 2 more than 1.90 (Africa).......

 parabasalis Slater

- 88a. Labium shorter, at most extending nearly to mesocoxae, second segment not attaining anterior margin of fore coxae90
- 89a. Second antennal segment relatively short, much less than 1 1/2 times length of segment 3 (.50-.40); posterior pronotal lobe with contrasting creamy yellow area limited to a trianguloid patch adjacent to each humeral angle, not complete across midline (Palearctic)suturalis Horvath
- 90. Membrane bearing a central black or dark brown macula which contrasts with pale basal and apical or lateral areas 91
- 90a. Membrane unicolorous or nearly so, or testaceous with veins darkened, or strongly micropterous with membrane lacking or represented by a small flap93
- 91. Pronotum with area across humeri narrowly

	dull gray to black remainder of pronotal
	surface (Palearctic)
	sabuleti and quadratus
91a.	Pronotum completely dull gray to black 92
92.	
	femora (South America)
02 -	
92a.	Tibiae black or dark castaneous, unicolorous with dark femora (South America)
	nigripes Stål
93.	Pronotum with narrow bright yellow area
,	across humeri or ovoid yellow spots on
	posterolateral areas strongly contrasting
	with dull gray to black remainder of pro-
	notal surface94
93a.	Pronotum lacking strongly contrasting yellow
	area across humeri, either completely dull
	gray to black or with diffuse reddish brown areas posterolaterally not strongly
	contrasting with ground color of pronotum
	96
94.	Labium relatively elongate reaching meso-
	coxae, second segment surpassing base of
	head by two-thirds its length; antennae
	usually yellowish brown to reddish brown; legs light brown to yellow (Palearctic).
	jaxartensis Reuter
94a.	Labium shorter, remote from mesocoxae, sec-
94a.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing
94a.	Labium shorter, remote from mesocoxae, sec- ond segment usually barely surpassing base of head; antennae and femora except
94a.	Labium shorter, remote from mesocoxae, sec- ond segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark
	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
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95. 95a.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
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95. 95a. 96.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95. 95a.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95. 95a. 96.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95. 95a. 96.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)
95. 95a. 96.	Labium shorter, remote from mesocoxae, second segment usually barely surpassing base of head; antennae and femora except proximal and distal ends frequently dark brown to black (Palearctic)

- 98. Body and appendages nearly uniformly brown to black; labium relatively elongate, at least fourth segment extending onto mesosternum for its entire length (Australia)sordidus Slater

- 99a. Labium longer, extending at least to posterior margin of prosternum (South America)transitius Slater and Wilcox
- 100. Fourth antennal segment considerably longer than segment 3; fore wings reduced to small pads laterad of scutellum, not nearly in contact on midline, no membrane flange present (South America) (fig. 50)
 proprius Slater

LEMURIBLISSUS SLATER Figure 52

Lemuriblissus Slater, 1967, p. 11.

Type Species: Lemuriblissus acuminatus Slater. Monobasic.

DISTRIBUTION: Madagascar, E. Africa.

BIOLOGY: Unknown.

DIAGNOSIS: Body robust, moderately elongate, subelliptical, non-linear. Metathoracic scent gland auricle short, rounded, earlike. Fore femora armed below with a single short stout spine, middle and hind femora mutic.

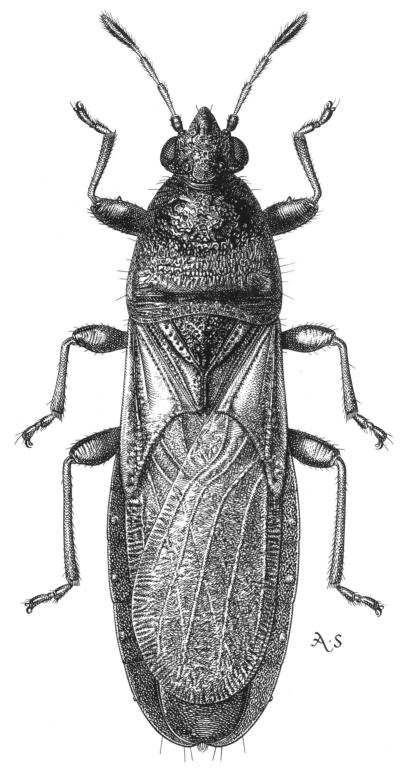


Fig. 52. Lemuriblissus acuminatus, dorsal view.

Entire body surface shining completely lacking pruinose areas. Apical corial margins strongly, deeply and evenly concave. Membrane thin, semitranslucent, much differentiated from surface of adjacent corium. Fore coxal cavities closed. Ocelli small. Antennae slender, prominently clavate. Genitalia unknown.

LUCEROCORIS SLATER Figure 53

Lucerocoris Slater, 1968, p. 281.

TYPE SPECIES: Lucerocoris nigrotibialis Slater. By original designation.

DISTRIBUTION: Oriental Region: Philippines.

BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, linear, nonflattened, subcylindrical. Metathoracic scent gland auricle strongly curved anteriorly forming a nearly right angle curve. Fore femora short, strongly incrassate, armed ventrally with numerous short, stout spines. Fore tibiae short, expanded, toothed at distal end. Middle and hind femora mutic. Head and pronotum above completely shining, non-pruinose. Scutellum pruinose but with median elevation shining. Apical corial margin straight. Membrane thick, nearly unitexturous with adjacent corium, its distal third strongly shining but proximal twothirds dull appearing subpruinose giving a twotextured appearance to membrane. Fore coxal cavities closed. Ocelli very large. Antennae short, stout, thick, second segment much shortened, shorter than segment one, subclavate. Claspers short, lobate, rounded. Sperm reservoir unknown. Ovipositor somewhat platelike, not completely dividing abdominal sternum seven.

KEY TO SPECIES OF LUCEROCORIS

- -brunneus Slater

MACCHIADEMUS SLATER AND WILCOX Figure 54

Macchiademus Slater and Wilcox, 1973, pp. 98-99.

TYPE SPECIES: Blissus diplopterus (Distant). By original designation.

DISTRIBUTION: South Africa.

BIOLOGY: Breeds on grasses chiefly of the genus *Ehrharta*. One species very destructive to wheat. Also known to breed on grasses of the genus *Pentaschistus* and *Pennisetum*. One species breeds on *Juncus*.

DIAGNOSIS: Body moderately elongate, sublinear. Metathoracic scent gland auricle slender, rounded, tapering to distal end. Fore femora incrassate with one or two spines present distally on ventral surface. Body lacking pruinosity both above and below except sometimes on metasternum. Apical corial margin straight. Membrane in large part transparent to translucent-hyaline, thinner than adjacent corium. Fore coxal cavities very narrowly open. Ocelli small. Antennae slender, terete or very slightly enlarged at distal end of segments two and three. Clasper generalized. Sperm reservoir unique with bulb large and ellipsoidal and with a median sclerotized band extending completely or partially through center of dorsal surface (see Slater and Wilcox, 1973), wings subtriangular or deeply incised, curving strongly ventrad so as to be scarcely visible in dorsal view. Ovipositor extending anteriorly to posterior margin of abdominal sternum five. See figure 2L for morphology of the spermatheca.

KEY TO SPECIES OF MACCHIADEMUS

- 2. Corium with distal half dark brown to black,

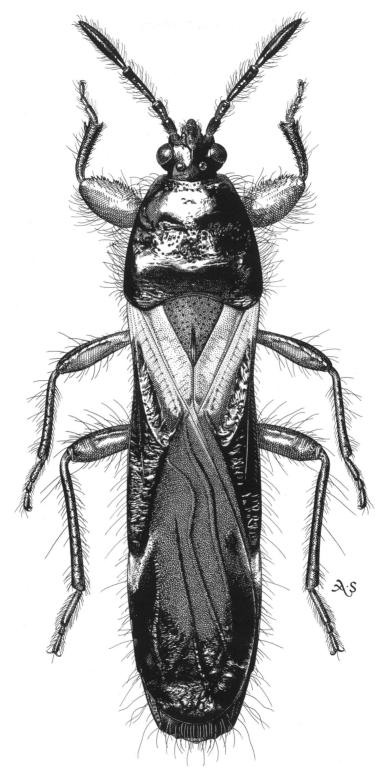


Fig. 53. Lucerocoris nigrotibialis, dorsal view.

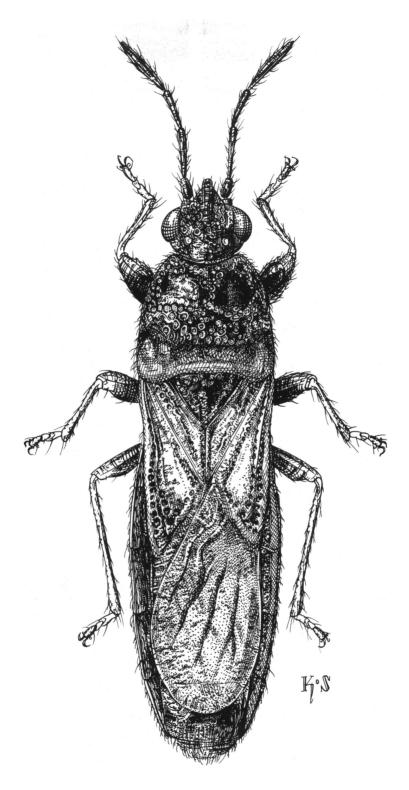


Fig. 54. Macchiademus diplopterus, dorsal view.

- 3. Labium elongate, extending well between or beyond mesocoxae, first labial segment attaining or nearly attaining base of head...
 acuminatus Slater and Wilcox
- 3a. Labium not or at most barely attaining mesocoxae, first labial segment remote from base of headnigritus Slater and Wilcox
- 4. Metasternum completely pruinose; abdominal sternum including connexival area completely dark chocolate brown to black....

MACROPES MOTSCHULSKY Figures 55-58

Macropes Motschulsky, 1859, p. 108. Rhabdomorphus Bergroth, 1918, pp. 68-69.

Type Species: *Macropes spinimanus* Motschulsky. Fixed by Distant 1904.

DISTRIBUTION: Africa, southern Asia to Australia.

BIOLOGY: Several species associated with bamboos but grasses of the genera *Sporobolus* and *Saccharum* also reported as host plants.

DIAGNOSIS: Body elongate, sublinear, varying from slender to robust. Metathoracic scent gland auricle either narrow and straplike (fig. 9E, I) or lobate (figs. 13B, 9D, F, P). Fore femora enlarged, incrassate, multispinose, usually lying closely appressed to a lateroventrally excavated anterior portion of prothorax. Head and prothorax shining, completely nonpruinose both above and below. Apical corial margin straight. Membrane varying from thin and subhyaline to thickened and nearly of same texture

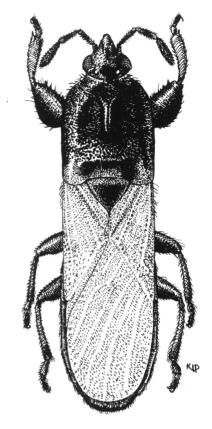


Fig. 55. Macropes uniformis, dorsal view.

as adjacent corium. Fore coxal cavities closed. Ocelli small. Sexual dimorphism and wing reduction not or little evident but submacroptery common. Some species with male fore femora more strongly incrassate than those of females. Antennae either terete or with segments two and three distally enlarged and subclavate. Fore tibiae usually somewhat swollen and armed with terminal or subterminal spines. Tarsi short, swollen, second segment generally relatively narrow and small. Head small. Claspers frequently scimitar-shaped with outer knob frequently placed far from base of clasper (fig. 2B), inner projection often obsolete. Sperm reservoir with a generally small rounded or elliptislender cup. wings and straplike. Spermathecal pump usually double flanged. i.e., separated into two parts by a transverse "suture" (fig. 2D, J).

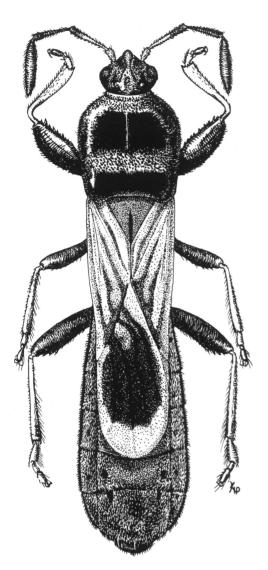


Fig. 56. Macropes raja, dorsal view.

KEY TO SPECIES OF MACROPES¹

- 1. Membrane either completely shining and somewhat translucent, or dull anteriorly and contrasting shining distally (fig. 7A, B)2

bacillus and sultanus not included.

- 2a. Membrane with anterior dull area extending posteriorly at least halfway along apical corial margins, posteriorly shining 13

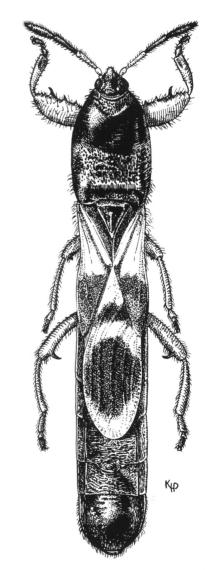


Fig. 57. Macropes varipennis, dorsal view.

 7a. Scutellum with shining non-pruinose area coffined to a narrow, elevated mesal stransometimes broadened basally	3a.	Membrane with exception of veins uniformly
darkened then lacking a contrasting whi opaque anterior area		pale or translucent throughout, or if slightly
opaque anterior area 4. Distal fourth of corium brown; membrane fore wing milky white, not markedly tran lucent; antennal segments 2, 3, and 4 blas or dark chocolate brown; \$\delta\$ with a proment "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		
 Distal fourth of corium brown; membrane fore wing milky white, not markedly tran lucent; antennal segments 2, 3, and 4 blas or dark chocolate brown; ♂ with a proment "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		anagua antoniar aras
fore wing milky white, not markedly tran lucent; antennal segments 2, 3, and 4 bla or dark chocolate brown; \$\delta\$ with a proment "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		opaque amerior area4
lucent; antennal segments 2, 3, and 4 blas or dark chocolate brown; 3 with a proment "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce	4.	
nent "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		fore wing milky white, not markedly trans-
nent "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		lucent: antennal segments 2, 3, and 4 black
nent "button-like" median protrusion on a dominal sternum 4; calli strongly pubesce		or dark chocolate brown: & with a promi-
dominal sternum 4; calli strongly pubesce		of dark chocolate brown, o with a profile
4a. Distal fourth of corium usually testaceous, un colorous with remainder of corium, if a pearing somewhat infuscated (African) the antennal segments 2, 3, and 4 brown, no black; calli shining, nearly glabrous (excein pilosus); no sternal protrusion on abdotinal sternum 4; membrane strongly transl cent		
 4a. Distal fourth of corium usually testaceous, un colorous with remainder of corium, if a pearing somewhat infuscated (African) the antennal segments 2, 3, and 4 brown, in black; calli shining, nearly glabrous (excessin pilosus); no sternal protrusion on abdotinal sternum 4; membrane strongly translent. 5. Pronotum relatively short and broad, ratio pronotal length to width less than 0.8 total length less than 4.0		
colorous with remainder of corium, if a pearing somewhat infuscated (African) the antennal segments 2, 3, and 4 brown, in black; calli shining, nearly glabrous (excessin pilosus); no sternal protrusion on abdotinal sternum 4; membrane strongly translent		
colorous with remainder of corium, if a pearing somewhat infuscated (African) the antennal segments 2, 3, and 4 brown, in black; calli shining, nearly glabrous (excessin pilosus); no sternal protrusion on abdotinal sternum 4; membrane strongly translent	4a.	Distal fourth of corium usually testaceous, uni-
pearing somewhat infuscated (African) the antennal segments 2, 3, and 4 brown, in black; calli shining, nearly glabrous (excessin pilosus); no sternal protrusion on abdous inal sternum 4; membrane strongly translecent		
antennal segments 2, 3, and 4 brown, in black; calli shining, nearly glabrous (excein pilosus); no sternal protrusion on abdormal sternum 4; membrane strongly translicent		
black; calli shining, nearly glabrous (excein pilosus); no sternal protrusion on abdorinal sternum 4; membrane strongly transl cent		
in pilosus); no sternal protrusion on abdorinal sternum 4; membrane strongly translecent		
inal sternum 4; membrane strongly transl cent		
inal sternum 4; membrane strongly transl cent		in pilosus); no sternal protrusion on abdom-
cent		
 Pronotum relatively short and broad, ratio pronotal length to width less than 0.8 total length less than 4.0		
pronotal length to width less than 0.8 total length less than 4.0	-	Durantum relatively short and broad ratio of
total length less than 4.0	Э.	
5a. Pronotum relatively more slender and elongate ratio of pronotal length to width more the 0.88 (in some specimens the ratio is less than 0.88, but these are relatively elongate total length over 5.0 (range 5.08-7.20). 6. Pronotum with posterior lobe light yellowing brown, the light coloration extending and riorly midway across transverse impression usually to anterior margin of impression		
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riorly midway across transverse impression usually to anterior margin of impression		
usually to anterior margin of impression		
6a. Pronotum red-brown or yellowish brown acrohumeri, lighter coloration at most reachin posterior margin of transverse impression. 7. Scutellum with a broad, shining area covering greater part of surface, pruinose areas confined to basal strip and narrow sub-basa areas. 7a. Scutellum with shining non-pruinose area confined to a narrow, elevated mesal strip sometimes broadened basally		
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posterior margin of transverse impression 7. Scutellum with a broad, shining area covering greater part of surface, pruinose areas confined to basal strip and narrow sub-basa areas 7a. Scutellum with shining non-pruinose area confined to a narrow, elevated mesal strip sometimes broadened basally 8. Median length of pronotum considerabing greater than width across humeri (fig. 5) 8a. Median length of pronotum subequal to or letter than width across humeri 9. Pronotum wider across calli than acroshumeri; interocular space more than 1 1 times as great as length of second antennous segment; evaporative area of metapleur coarsely punctate; corium uniformly test ceous (fig. 55) 9a. Pronotal width across calli subequal to width	6a.	
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fined to basal strip and narrow sub-basareas	/٠	
areas		greater part of surface, pruinose areas con-
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fined to a narrow, elevated mesal str sometimes broadened basally		areas
fined to a narrow, elevated mesal str sometimes broadened basally	7a.	Scutellum with shining non-pruinose area con-
8. Median length of pronotum considerab greater than width across humeri (fig. 5		fined to a narrow, elevated mesal strip
 Median length of pronotum considerabe greater than width across humeri (fig. 5		sometimes broadened basally 10
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than width across humeri	8a.	Median length of pronotum subequal to or less
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segment; evaporative area of metapleuro coarsely punctate; corium uniformly test ceous (fig. 55)		
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9a. Pronotal width across calli subequal to wid		ceous (fig. 55)uniformis Distant
	9a.	
		across humeri; interocular space less than 1

136

1/4 times as great as length of second antennal segment; evaporative area of metapleuron impunctate or nearly so; apical area of corium with brown infuscations..... Slater and Wilcox 10. Body very elongate, total length of body always more than seven times length hind tibia; legs generally uniformly bright yellowsubauratus Distant 10a. Body somewhat less elongate, total length of body less than seven times length hind tibia; femora generally fuscous to very dark 11. Pronotum with humeral width greater than median length, lateral pronotal margins usually slightly but evenly and distinctly tapering from humeral angles to anterior margin; ratio of head width to pronotal width less than 0.55.....pseudofemoralis Slater, Ashlock, and Wilcox 11a. Pronotum with mesal length as great as or greater than humeral width; lateral pronotal margins nearly parallel-sided from humeral angles to area of calli; ratio of head width to pronotal width 0.55 or greater12 12. Antennal segment 4 twice or more than twice length of antennal segment 2; pronotum appearing to taper gradually from humeral angles to area of calli femoralis Distant 12a. Antennal segment 4 less than twice length of antennal segment 2; pronotum appearing nearly parallel-sided from humeral angles to area of calli..... harringtonae Slater, Ashlock, and Wilcox 13. Dull area of membrane confined to extreme anterior portion, not extending more than midway along apical corial margin (fig. 7B)privus Distant 13a. Dull area of membrane extending posteriorly adjacent to apical corial margin at least almost to distal end of corium (fig. 7A)... 14. Smaller species, less than 4.72 mm. long; radial vein of corium shining but area laterad (to margin) contrastingly dull Maculosus Slater and Wilcox 14a. Large elongate species, over 6.5 mm.; lateral area of at least anterior three-eighths of corium including radial vein completely 15. Entire lateral area of corium including radial vein completely dark brown..... alternatus Slater and Wilcox 15a. Lateral area of corium at least in part white or pale testaceous, or if appearing dark then

middle and hind femora multispinose...16

16. Membrane with anterior dark area and large

	median dark macula dull, lunate transverse pale vitta present just beyond apices of
	coria and distal pale area contrastingly shin-
16.	ing
roa.	Membrane with dull area confined to anterior
	area and along apical corial margin, median dark macula shining as are lunate pale vitta
	and apical pale area
17	Eyes relatively sessile, not strongly produced
17.	above head surface; inner (median) vein of
	corium usually pale anterior to posterior
	end of claval commissure; anterior end of
	membrane between coria usually dark; lu-
	nate transverse white vitta across membrane
	often obsolete and reduced to lateral spots
	albosignatus Distant
17a.	Eyes prominently elevated above head surface;
	inner vein of corium usually darkened for
	some distance anterior to posterior end of
	claval commissure; anterior portion of
	membrane usually pale or at least with pale
	spots; pale lunate vitta on membrane com-
	plete nigrolineatus Distant
18.	1 7 1
10.	Satelli Illiani Slater, Ashlock, and Wilcox
18a.	Scutellum shining on median carina and adja-
10	cent areas on distal fourth
17.	taining lateral margins, latter broadly pale
	punctatus (Walker)
19a.	Dark membranal macula broadly attaining lat-
	eral marginsnigrolineatus Distant
20.	Hemelytra predominately light brown or
	yellow, at most with veins of membrane
	and distal third of corium darkened, never
	with a variegated contrasting black or
	brown and white pattern on hemelytra (oc-
	casional specimens of burmanus and exilis
	show an obscure darkening on the mem-
	brane mesally but not a sharply contrasting
	macula and the veins in the area in such cases are always much darker and strongly
	contrasting)21
20a	Hemelytra dark brown or with a variegated
204.	black or brown and white pattern usually
	including a large central dark membranal
	macula30
21.	Radial vein dark brown, shining throughout
	and coalescing distally with shining apical
	corial margin; membrane including veins
	corial margin; membrane including veins uniformly dull brown throughout
21	
21a.	Radial vein generally nearly unicolorous with
	adjacent areas of corium and shining only
	on anterior two-thirds or if appearing shin- ing throughout then membrane including
	veins not unicolorous dull brown22

22. Labium short, not extending posteriorly to fore coxaerufipes Distant

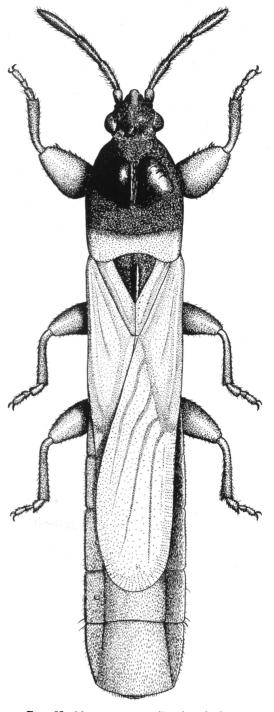


Fig. 58. Macropes australis, dorsal view.

22a.	Labium more elongate, always extending be-		fourth sternum (fig. 58)
	tween fore coxae and usually attaining pos-		australis (Distant)
	terior margin of mesosternum23	28.	Distal fourth of corium dark fuscous, strongly
23.	Pronotum strongly produced laterally anterior		contrasting with pale testaceous proximal
	to transverse impression, conspicuously		area; ♂ with a prominent "buttonlike" pro-
	wider across calli than across humeri; or if		trusion mesally on abdominal sternum 3
	only slightly wider across calli in macrop-		obnubilis (Distant)
	ters, then large species, over 8.1 mm	28a.	Entire corium uniformly light testaceous, lack-
	24		ing dark fuscous coloration apically; ರೆ
23a.	Pronotum nearly parallel sided from humeri to		lacking a median sternal protrusion29
	area of calli with width across calli sube-	29.	Body relatively short and stout, only four
	qual to that across humeri; or if somewhat		times as long as wide consimilis Distant
	wider across calli then small species, under	29a.	Body relatively elongate and slender, at least
	7.6 mm25		five times as long as wide
24.	Broad rather flattened species; pronotal calli		pronotalis Distant
	smooth and glabrous with three foveate de-	30.	Pronotum with a deep non-punctate median
	pressions present; apex of tylus exceeding		groove running longitudinally on midline
	apex of first antennal segment		between calli; hind femora generally con-
			spicuously armed with small spines (except
24a.	Elongate, more linear, not conspicuously flat-		in dilutus) (raja complex)31
	tened; pronotal calli without foveate depres-	30a.	Pronotum lacking a deep median non-punctate
	sions, shallowly punctate, often		longitudinal furrow between calli, some-
	conspicuously pubescent; apex of tylus at		times with one or two rows of indented
	most attaining apex of first antennal seg-		punctures in the area forming a shallow
25	ment		groove or with a fine impressed line on
25.	Veins of membrane light to dark brown, con-		either side of meson, or if appearing
	trasting with yellowish to testaceous ground		grooved, then & hind femora with a
	color of membrane, or if membrane with		swollen protrusion midway along ventral
	brown macula then veins nearly unicolorous		margin (see <i>spinimanus</i>); with or without
250	brown		spines on posterior femora (spinimanus
23a.	Veins of membrane transparent or testaceous,	21	complex)
	unicolorous or nearly so with ground color	31.	Middle and hind femora mutic
26	of membrane	210	
20.	distal third of corium nearly uniformly		Middle and hind femora spinose32
	chocolate brown	32.	Middle and hind tibiae light yellow to testa-
	exilis Slater and Wilcox		ceous, strongly contrasting with dark femora (fig. 56)raja Distant
26a	Legs nearly uniformly pale yellow to light	320	All tibiae dark brown to black, unicolorous or
Lou.	brown; distal third of corium testaceous at	32a.	nearly so with dark femora (posterior tibiae
	least on lateral half (some specimens of		sometimes lighter brown on distal half
	burmanus have the distal third of corium		33
	darkened but in such cases the pronotum is	33	Clavus and at least anterior third of corium
	wider than long)	55.	nearly uniformly opaque white
27.	Stout, robust species, body length less than		
	five times humeral width; labium reaching	33a.	Clavus and/or anterior third of corium with at
	onto anterior margin of mesosternum, sec-		least veins contrasting dark brown34
	ond segment nearly reaching anterior mar-	34.	Membrane with a large, round, dark median
	gin of fore coxae; ♂ with "button-like"		macula not attaining lateral margins and
	protrusion mesally on sternum 4 (third visi-		separated from basal dark area by a broad,
	ble) burmanus Slater and Wilcox		transverse, pale, lunate vitta adjacent to
27a.	Elongate, slender species, body length at least		apex of corium (the vitta sometimes inter-
	5 2/5 times humeral width (usually six or		rupted with diffuse narrow brown areas)
	more times); labium shorter, not exceeding		
	fore coxae, second segment barely surpass-	34a.	Membrane dark coloration not a distinct,
	ing base of head; & lacking protrusion on		black, round spot, entire basal two-thirds

	membrane black, or with at most a small light area at extreme base adjacent to distal end of corium and a small diffuse light area in center of disc; dark membranal area broadly in contact with lateral margins
35.	Antennae relatively short, length of segment 2 equal to or barely greater than interocular distance
	comosus Slater, Ashlock, and Wilcox
	Antenna relatively long, length of segment 2 at least 1.33 times interocular distance 36
36.	Large, robust species (8.28-10.92 mm.); pronotum noticeably wider across area of calli than across humeral angles; length of pronotum subequal to basal width; metathoracic scent gland auricle short, broad, very slightly curving anteriorly
36a.	Smaller, less robust species (7.56 mm.); distance across pronotal calli subequal to width across humeral angles; length of pronotum greater than basal width; scent gland auricle narrow, linear, strongly curving anteriorly
37.	Labium very short, remote from fore coxae; & with extremely elongate, curving spines distally below on hind femora (fig. 57)
37a.	Labium extending caudad to or beyond fore coxae; hind femoral spines, if present, very short and stout
38.	
38a.	Anterior fourth of corium with some pale markings or areas present
39.	Membrane chiefly black from base to apex, dark coloration extending to apical and lateral margins along entire membrane except in area of ovoid white spot at distal end of corium
39a.	Large dark macula on membrane, for the most part remote from apical and lateral margins (sometimes attaining lateral margin across subapical third)
40.	Length of body greater than 7.0 mmpraecerptus Distant
40a.	Length of body less than 6.5 mm
41.	Scutellum with anterolateral angles produced into upward-tipped, shining tubercles (sometimes very inconspicuous); antennae with at least segments 1 and 2 pale yellow

- 42a. Larger species, over 6.5 mm. (range 6.48-6.90 mm.); ♂ lacking swollen protrusion on hind femur; legs usually bright yellow...
 ...yoshimotoi Slater, Ashlock, and Wilcox

MERINADEMUS SLATER Figure 59

Merinademus Slater, 1967, p. 16.

Type Species: Merinademus baraoides Slater. Monobasic.

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

DIAGNOSIS: Body extremely elongate. slender, linear, not strongly flattened. Metathoracic scent gland auricle elongate, broadening to a strongly rounded distal end. Fore femora strongly incrassate armed below on distal third with a single large, elongate, acute spine. Fore tibiae short, stout, broadening distally. All legs short and stout. Body above and below completely shining, no pruinosity present. Apical corial margin strongly concave. Membrane thin, semitranslucent. Fore coxal cavities closed. Ocelli small. Submacropterous. Antennae with segments two and three short and conspicuously clavate.

MICAREDEMUS SLATER Figure 60

Micaredemus Slater, 1967, p. 29.

Type Species: *Micaredumus elegans* Slater. By original designation.

DISTRIBUTION: Madagascar, Africa.

BIOLOGY: Breeding known on several genera of grasses.

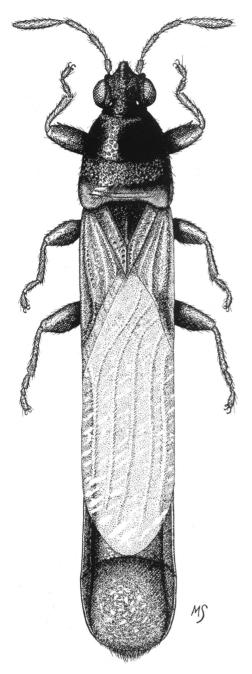


Fig. 59. Merinademus baraoides, dorsal view.

DIAGNOSIS: Body elongate, moderately broad, not strongly flattened. Scent gland auricle strongly, lunately curving forward (fig. 10F). Fore femora with one, two, or rarely

more small spines. Body completely shining above and below, even clavus and corium shining. Apical corial margin straight. Membrane hyaline, well differentiated in texture from corium. Fore coxal cavities closed. Ocelli small. Brachyptery and submacroptery frequent. Antennae terete or with segments 2 and 3 nar-Claspers slender, rowly clavate. projection obsolete, outer knob sub-triangular. Sperm reservoir minute, reduced to a small median scooplike projection (fig. 1GG), wings absent. Ovipositor elongate. Spermatheca with bulb lacking a basal flange; pump thick distally, strongly curved, elongate and tapering to a narrow proximal end (fig. 2M).

KEY TO SPECIES OF MICAREDEMUS
1. Fore femora armed below with two or more sharp spines
la. Fore femora armed below with a single sharp
spine
2a. Fore femora armed with two spines 6
3. Membrane of fore wing with a central longitudinally ovoid diffuse brown macula (West
Africa)gillonae Hamid and Slater
3a. Membrane translucent to transparent hyaline, lacking a central brown macula4
4. Fore femora armed with three spines; labium
extending onto mesosternum, second seg-
ment surpassing base of head by two-thirds
of its length (South Africa)
4a. Fore femora armed with four or five spines
(occasionally reduced to three spines on
first femur); labium shorter at most extend-
ing to fore coxae, second segment barely
exceeding base of head
5. Small species (3.12-3.52 mm.); male fore femoral spine configuration unique (fig. 7F)
(female unknown); pronotum wider across
anterior lobe than across humeri
(Madagascar)
quadratus Slater and Wilcox
5a. Larger species (5.0-6.0 mm.); fore femoral
spines in conventional row on ventral sur-
face; pronotum parallel sided for most of
length, not wider across anterior lobe than
across humeri (South Africa)
denticulatus Slater
6. Labium elongate, extending caudad at least
midway onto mesosternum (Madagascar)
(fig. 60)pilosulus (Horvath)

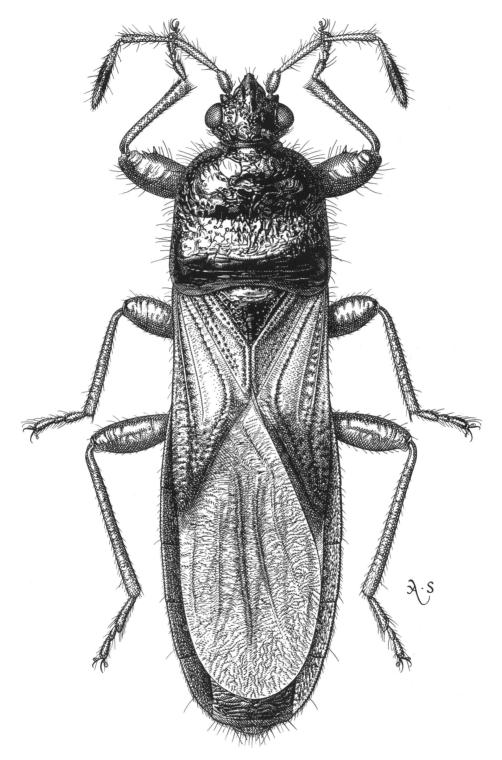


Fig. 60. Micaredemus pilosulus, dorsal view.

slightly beyond fore coxae	n inter-
7a. Length of antennal segment 3 less than interocular distance	rica) 1 Slater
8. Clavus pale testaceous to transparent hyaline, strongly contrasting with dark scutellum. 9. Prontoum densely clothed with decumbent silvery hairs, posterior lobe bright reddish brown, strongly contrasting with black anterior lobe (Madagascar)pilosus Slater	16
8a. Clavus chiefly chocolate brown to black, not strongly contrasting with dark scutellum. 9. Prontoum densely clothed with decumbent silvery hairs, posterior lobe bright reddish brown, strongly contrasting with black anterior lobe (Madagascar)pilosus Slater 16a. Pronotal length greater than width; scute longer than wide; femora chocolate to brown; tylus extending to or surpassing terior end of first antennal segment (Madagascar)	raceous
9. Prontoum densely clothed with decumbent silvery hairs, posterior lobe bright reddish brown, strongly contrasting with black anterior lobe (Madagascar)pilosus Slater	agascar) s Slater
` U , .	to dark sing an- segment
coming dark brown across humeri, sparsely clothed with short semi-decumbent hairs	Wilcox
(Africa)	
10a. Legs and first three antennal segments bright Type Species: Ischnodemus dilutipes	s Stål.
yellow or ochraceous	
11. Labium short, barely attaining fore coxae, second segment extending just beyond base of head; fore femoral spines divergent, set on small tubercle (Madagascar)	ıt, lin-
obscurellus Slater ear. Metathoracic scent giand auricle ust	
lla. Labium longer, reaching anterior margin of mesosternum, second segment extending well beyond base of head; fore femoral spines parallel, not set on tubercle (East	with a cluding
Africa)wilcoxae Hamid and Slater below. Apical corial margins straight. M	
12. Head deeply concave between eyes in area of brane thinner than adjacent corium. Fore c	
base of tylus (fig. 5A); anterior pronotal lobe convex on inner half, but laterally sharply flattened and depressed to give a ledgelike appearance to outer third (Madagascar)eleganoides Slater cavities closed. Ocelli small. Antennae ter Eyes conspicuously laterally produced on stalks. Labium very short not reaching coxae. Claspers conventional. Sperm reservables	n short g fore servoir
12a. Head normally convex between eyes (fig. 5B); pronotum convex, curving evenly downward from meson to lateral margin with no evidence of a flattened shelflike lateral third (Madagascar)elegans Slater	a with
13. Head deeply concave between eyes in area of KEV TO SPECIES OF PATRITIONEMI	MUS
base of tylus (ng. 5A); anterolateral prono-	
tal margin flattened and shelflike 1. Fore femora armed with at least one short s (Madagascar)	
13a. Head normally convex between eyes in area of la. Fore femora mutic	5
base of tylus (fig. 5B); anterolateral area of pronotum not flattened and shelflike14 2. Fore femoral spine simple, small and conspicuous; small species little over	
14. Length third antennal segment greater than in-	ane of
terocular space (Madagascar) hemelytra with a pale spot near base	

with an acute accessory spine extending at right angles to it; large robust species, over 7.0 mm. long; no pale spot on membrane of hemelytra4 3. Labium reaching base of prosternum; hemelytra lacking a diffuse dark central area 3a. Labium not quite attaining anterior margins of fore coxae; hemelytra with a diffuse darkalbomaculatus Slater and Ahmad 4. Pronotum and abdominal tergum unicolored, dark chocolate brown; antennal segments two and three slender; claspers without distinct inner lobes; male gonopore very small; spermathecal pump narrow and elongate 4a. At least part of pronotum and abdominal tergum pale brown, antennal segments two and three distinctly clavate; clasper with well-developed inner knob; male gonopore large; spermathecal pump wide and short relative to 5. Antennae and legs black or very dark chocolate brown unicoloris Slater and Ahmad 5a. Legs and at least basal segments of antennae light yellow to reddish brown 6 6. Membrane strongly suffused with dark brown on basal half; inner knob of clasper little produced: scutellum distinctly broader than 6a. Membrane pale on basal half as well as distally; inner knob of clasper strongly produced; scutellum as long as basal width7 7. Males less than 4.5 mm. long; second antennal segment subequal to length of scutellum; base of sperm reservoir lacking an accessory "yokelike" sclerite

PATRITIUS DISTANT Figures 61, 62

7a. Males over 6.0 mm. long; second antennal seg-

..... Slater and Ahmad

ment considerably longer than scutellum; base of sperm reservoir with a distinct "yokelike" accessory sclerite.....

..... Singularis Slater and Ahmad

Papirius Stål, 1865, p. 122. (Preoccupied).
Patritius Distant, 1901, p. 468 (new name for Papirius).

Type Species: *Papirius grossus* Haglund 1868. First included species.

DISTRIBUTION: South America, Cuba.

BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, linear, non-flattened. Metathoracic scent gland auricle elongate, slender, tapered distally, curving either anteriorly or posteriorly (fig. 7J, K). All legs multispinose. Males frequently with enlarged hind femora. Thoracic pruinosity variable from completely pruinose above and below to completely shining above, frequently chiefly pruinose above with sub-basal shining area and large shining "patches" or bands in area of calli. Membrane usually somewhat thinner than corium. Fore coxal cavities closed. Ocelli small. Usually with short genal tubercles or protrusions. Antennae terete, relatively thick. Sperm reservoir with elongate, slender, twisted wings, bulb slender (fig. 1C, D).

DISCUSSION: Patritius as currently delimited contains a number of rather dissimilar species. My interpretation of the cladistic relationships are as in figure 63. This cladogram includes four as yet undescribed species (each known from a single specimen). These undescribed species are important phylogenetically as they include the two most plesiomorphic species and also the most apomorphic. The plesiomorphic species resemble some species of *Ischnodemus* and indicate that Patritius is derived from an Ischnodemus clade with pruinosity still present on the dorsal pronotal and scutellar surfaces and before the development of enlarge platelike wings on the sperm reservoir. Within Patritius apomorphies are generally similar in nature to those that occur in other blissine phyletic lines, i.e., reduction of pruinosity, modification of scent gland auricle shape, sexual dimorphism of the posterior femora etc.

KEY TO SPECIES OF PATRITIUS

- 3. Either antennal segment 2 or 3 considerably longer than any individual labial segment (53:61)....colombianus Slater and Wilcox

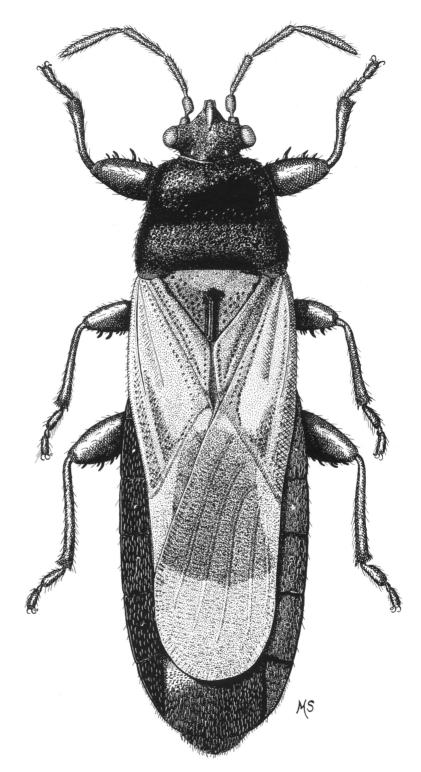


Fig. 61. Patritius colombianus, dorsal view.



Fig. 62. Patritius new species, dorsal view.

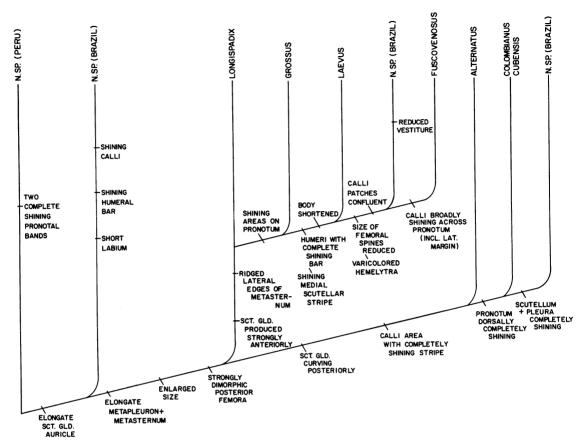


Fig. 63. Cladogram of *Patritius* species.

- 3a. Either second or third antennal segment subequal to or shorter than any individual labial segment cubensis Barber
- 4. Pronotum with a complete broad shining band across area of calli and a second band across area of humeri fusconervosus (Stål)
- 4a. Area of calli with at most a pair of shining triangular areas, never with a complete broad shining band; basal pronotal area either with or without shining area5
- 5. Posterior lobe of pronotum completely pruinose; lacking shining sub-basal areas between humeri longispadix Slater and Wilcox
- 5a. Posterior lobe of pronotum with a narrow shining area between humeral angles6
- 6. Shining area between humeri separated into three distinct patches, central elongate, lateral ovoid with a narrow but distinct pruinose area separating them......grossus (Haglund)

6a. Shining pronotal area between humeri complete, without a narrow pruinose area separating lateral and central patcheslaevus (Stål)

PIRKIMERUS DISTANT Figures 64, 65

Pirkimerus Distant, 1904, pp. 21-22. Ischnomorphus Hidaka, 1961, pp. 255-256.

Type Species: Pirkimerus sesquipedalis Distant. Monobasic.

DISTRIBUTION: Southern Asia, Japan, Philippines to New Guinea.

BIOLOGY: Collecting records from bamboo.

DIAGNOSIS: Body slender, elongate, often subcylindrical; metathoracic scent gland auricle unique, ovoid (fig. 13A); fore femora strongly incrassate, mutic or armed with one to four

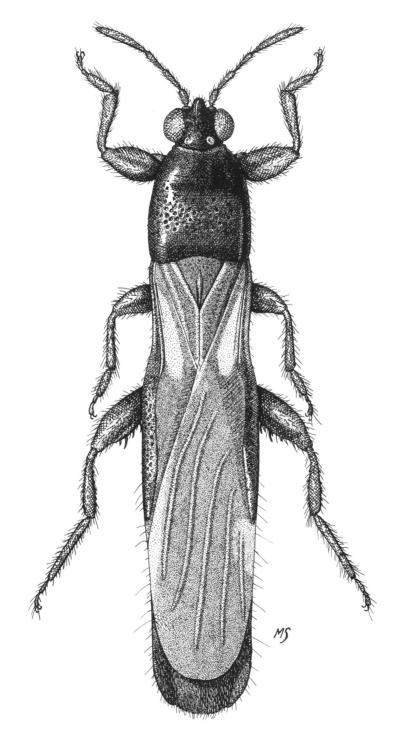


Fig. 64. Pirkimerus sesquipedalis, dorsal view.

small spines; hind legs enlarged, thickened, hind tibiae usually thickened, shortened and bearing a series of spines and teeth, hind femora toothed ventrally; corium elongate, tapering.

Head and pronotum above shining, polished, at most pruinose as an anterior collar, propleuron pruinose or shining; scutellum pruinose except along midline, hemelytra pruinose but often with a shining lateral stripe and radial vein. Fore coxal cavities closed. First tarsal segment enlarged, flattened with a "pad" of hairs below, first hind tarsal segment very elongate. Ocelli very large. Eyes frequently dimorphic, when so then larger in females. Labium frequently very short. Antennae in apomorphic species stout, clavate. Claspers small and stout lacking lateral "knob," sometimes blocklike; sperm reservoir with bulb reduced to a slender median projection, wings variable from absent to moderately large diverging lobes; spermatheca with short stout pump, tube variable from very short to elongate and completely coiled. Ovipositor much reduced, not laciniate.

DISCUSSION: This genus, as may be seen by reference to figure 15, is one of the most "advanced" of all the genera of Blissinae. The species are held together by the strongly apomorphic character of the unique ovoid and highly modified metathoracic scent gland auricle (fig. 13A). Within the genus (fig. 66) there are two major components: a plesiomorphic element containing four closely related species, japonicus, gadrii, javanus, and burmanus which are characterized by relatively elongate nearly terete antennal segments, slender elongate hind tibiae, similar eyes in both males and females, elongate hairs on the body surfaces and generally a pruinose non-shining corium. The peculiar species papuensis (fig. 65) from New Guinea is somewhat intermediate between these generalized forms and the advanced species discussed below. Iphicrates papuensis is a remarkable insect because of the highly modified abdominal segments (see Slater, 1968) but retains some of the generalized features of *iap*onicus and its allies, particularly the elongate non-spinose hind tibia. The antennae, however, are shortened and clavate and thus more similar

to the advanced forms of Pirkimerus. Unfortunately, the male is not known so one cannot tell whether or not papuensis has dimorphic eyes. The more apomorphic species of Pirkimerus are quite unusual in the strongly flattened eves of the males in contrast to the large globose eyes of the females and particularly in having the hind tibia very much shortened, thickened, usually covered with numerous short spines. Within this complex, as the cladogram (fig. 66) indicates, there is a progressive reduction in size and an increase in the narrowness and the cylindrical shape of the body until one reaches such minute species as nicobarensis, parviceps and philippinensis. Within the genus Pirkimerus, as within many other lineages in the Blissinae, there is a loss of pruinosity which is evident in this advanced group. A reduction sequence is apparent, from P. bellus, which has well-developed pruinosity on the propleuron almost up to the dorsal surface, through P. chinai, which has propleural pruinosity reduced but still evident dorsal to and posterior to the acetabula, to the small advanced forms, where if prothoracic pruinosity is present at all it is confined to the extreme anterior collar area and to the center anterior to the fore coxae. The accompanying cladogram (fig. 66) is a hypothesis of the probable cladistic relationships of the known species. Pirkimerus esakii is not included as I have not been able to reexamine specimens.

KEY TO SPECIES OF PIRKIMERUS

- 2a. Corium with a narrow shining glabrous longitudinal stripe at least on basal half in area

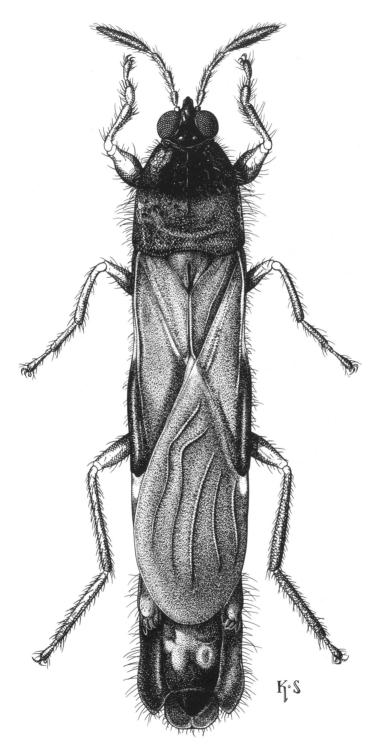


Fig. 65. Pirkimerus papuensis, dorsal view.

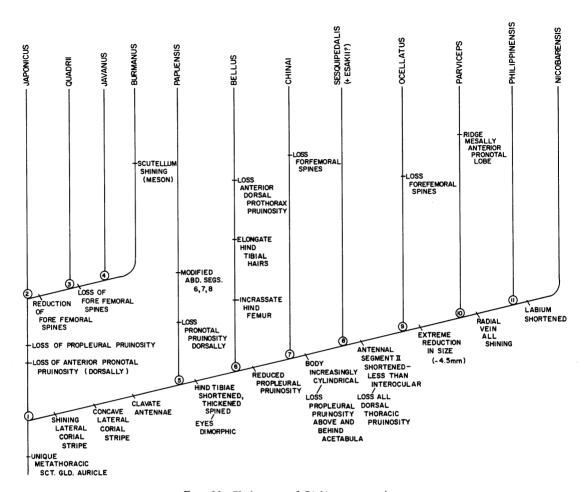


Fig. 66. Cladogram of Pirkimerus species.

of radius; hind tibiae short and thickened, bearing a series of prominent spines and tubercles; eyes strongly sexually dimorphic, in δ forming a nearly even curvature with lateral head margins6 3. Fore femora mutic4 3a. Fore femora with one to three small spines on ventral surface5 4. Scutellum completely pruinose, lacking a shining glabrous median carina on distal half (fig. 5L); basal half of corium and all of clavus uniformly bright yellow; femora light yellow; posterior margin of pronotum very shallowly concave..... javanus Slater and Ashlock 4a. Scutellum possessing a glabrous, shining median carina distally (fig. 5K); corium and

...... burmanus Slater and Ashlock

5. Fore femora with ventral spines confined to distal third; hind tibiae of ♀ less than six times as long as first antennal segment (♂ unknown) qadrii Slater and Ashlock

- 5a. Fore femora with ventral spines located centrally, one on inner face and one opposite on outer face; hind tibiae of females more than seven times as long as first antennal segment japonicus Hidaka
- Membrane of fore wing with a discrete brown central macula separated from lateral and

	apical margins by distinct white areas
	esakii Miyamoto and Hidaka
6a.	Membrane with brown area extending broadly
	to lateral and apical margins at least on
	distal third7
7.	Prothorax completely pruinose below, lacking
	shining areas except on acetabulae
	bellus Slater and Ashlock
70	
/a.	Prothorax shining or subshining below, lacking
•	pruinose areas8
8.	Corium with narrow area laterad of shining
	radial vein contrastingly pruinose brown
	parviceps Bergroth
8a.	Corium with area laterad of radial vein shining
9.	Pronotum with a narrow pruinose band present
	adjacent to anterior margin (fig. 64)
	sesquipedalis Distant
00	Pronotum completely shining, lacking anterior
9a.	
10	pruinose band
10.	Fore femora armed below on distal third with a
	single sharp spine (very short and in-
	conspicuous in ocellatus)11
	Fore femora mutic12
11.	Large species, over 8.0 mm. long; fore femora
	with a long sharp curving spine below on
	distal third
Ha.	Smaller species, under 6.0 mm.; fore femora
	with a short, inconspicuous spine below
	ocellatus Slater and Ashlock
12	Labium relatively short, not or barely attaining
12.	
	anterior margin of fore coxae, second seg-
	ment scarcely reaching base of head; prono-
	tal length and width subequal
	nicobarensis Distant
12a.	Labium longer, nearly reaching posterior mar-
	gin of prosternum, second segment exceed-
	ing base of head by more than half its
	length; pronotum much longer than wide
	philippinensis Slater
	1 11

PRAEBLISSUS BARBER Figure 67

Praeblissus Barber, 1949, p. 141.

Type Species: Praeblissus albopictus Barber. Monobasic.

DISTRIBUTION: Mexico.

BIOLOGY: Intercepted at quarantine stations on orchids.

DIAGNOSIS: Body short, stubby, thick; metathoracic scent gland auricle small, short, rounded and earlike; all femora mutic; anterior half of pronotum above and head above and below strongly shining; posterior half of pronotum above and all of thorax below completely pruinose; wing polymorphism present, brachypters and micropters known; macropters with only moderately concave apical corial margin; fore coxal cavities open; no marked sexual dimorphism; antennae terete or moderately clavate; ocelli small; sperm reservoir with large cup and elongate, slender straplike wings; ovipositor elongate.

PRAETORBLISSUS SLATER Figure 68

Praetorblissus Slater, 1966, pp. 3-11.

Type Species: Praetorblissus gigas Slater. Monobasic.

DISTRIBUTION: South and Central America. BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, robust, linear or sometimes slightly sub-elliptical often broadened. Metathoracic scent gland auricle large, almost spatulate, distally often curving anteriorly. All femora usually multispinose. Dorsal pruinosity absent, sometimes with pruinose areas present on ventral surface of head, mesally between and before coxae on prosternum and as a narrow strip across anterior margin of mesosternum and mesopleuron. Apical corial margin straight or slightly concave on inner third. Membrane thick, opaque, not strongly differentiated from adjacent corial surface. Eyes sessile. Ocelli small. Extreme microptery common with wing pads reduced to minute scalelike pads that do not reach abdomen. Antennae terete. Fore coxal cavities open. Claspers slender, lacking a conspicuous inner projection. Sperm reservoir small with a nearly membranous bulb and slender wings.

KEY TO SPECIES OF PRAETORBLISSUS

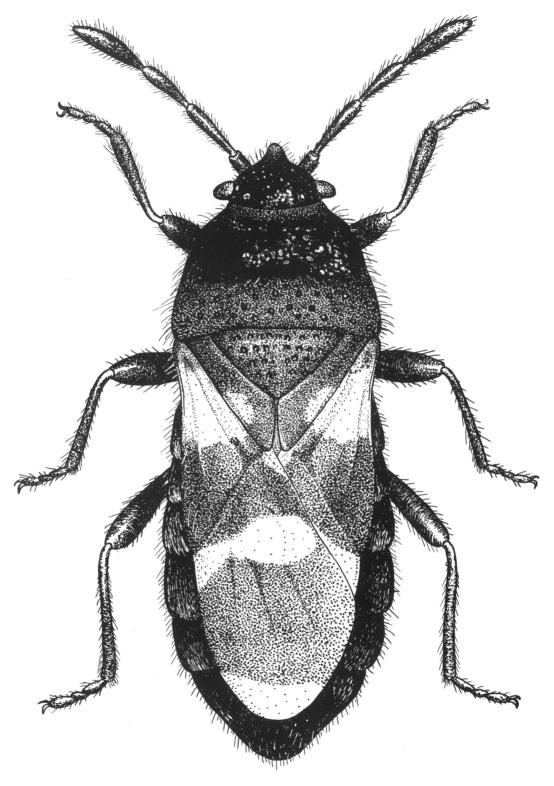


Fig. 67. Praeblissus albopictus, dorsal view.

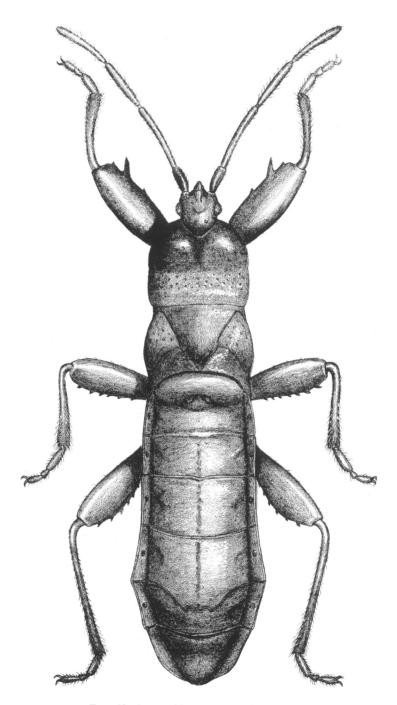


Fig. 68. Praetorblissus gigas, dorsal view.

- 2a. Length of second antennal segment 1-1/2 times as great as interocular width; body color reddish brown; relatively large, more than 12.0 mm. long (fig. 68)gigas Slater

PROCELLADEMUS SLATER AND WILCOX Figure 69

Procellademus Slater and Wilcox, 1966, p. 69.

Type Species: *Procellademus venenatus* Slater and Wilcox. By original designation.

DISTRIBUTION: South America.

BIOLOGY: Unknown.

DIAGNOSIS: Body elongate, robust. Metathoracic scent gland auricle very large, lobate (fig. 12A). Fore femora strongly incrassate, multispinose; middle and hind femora mutic. Head and pronotum shining above, the latter pruinose below on sternal and pleural surfaces. Scutellum completely pruinose. Apical corial margin straight. Membrane thin, well differentiated from much thicker corium. Fore coxal cavities closed. Ocelli small. No wing reduction or sexual dimorphism known. Antennae terete. Ovipositor elongate. Sperm reservoir with relatively small bulb but large platelike wings. Spermathecal bulb small and elliptical, pump slender, moderately elongate.

KEY TO SPECIES OF PROCELLADEMUS

- 1. Second antennal segment subequal to or slightly longer than interocular space (ratio 1.02-1.11); labial segment one as long as, or slightly longer than segment two; larger, more elongate species (7.0-8.0 mm.) (fig. 69)venenatus Slater and Wilcox

PSEUDOBLISSUS, NEW GENUS Figure 70

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

DIAGNOSIS: Head, thorax, and scutellum shining, completely lacking pruinosity ventrally or laterally; antennal segments 2 and 3 clavate, fourth segment fusiform, strongly and abruptly narrowed on proximal one-fifth; prosternum compressed anterior to coxae, prominently produced mesally; fore coxal cavities closed; scutellum lacking a median elevation; fore femora strongly incrassate, armed below with three or four stout spines, distal spines broad, usually bifid, angled toward bases of tibiae, latter somewhat swollen at distal ends with series of short spines present, but not flattened: metathoracic scent gland auricle short, broadly ovoid, earlike; eyes small, set well away from anterior margin of pronotum; antenniferous tubercles simple, truncate, not produced as curved processes; only micropters known, wing pads broad, not meeting at midline; apical corial margins deeply concave. Sperm reservoir reduced to a median plate.

Type Species: Blissus trispinosus (Slater).

DISCUSSION: Slater (1967) tentatively placed trispinosus in the genus Blissus (sensu lato) but pointed out the anomalous nature of the species and its questionable relationship to Blissus and its relatives. The closed fore coxal cavities and lack of prothoractic and scutellar pruinosity indicate that trispinosus is not at all closely related to either Blissus or Dimorphopterus, and that it must be considered to represent a distinct genus. Despite striking differences in habitus, Pseudoblissus appears to be most closely related to Aradademus which is also endemic to Madagascar. The heavy bifid angulated fore femoral spine arrangement, strongly tapered, nearly petiolate proximal fourth antennal segment and the completely non-pruinose bodies are all common to the two genera. Aradademus, with its produced antenniferous tuflattened bercles. greatly body. highly specialized eyes and unique metathoracic scent gland auricle is readily recognizable. Actually Pseudoblissus probably represents a more generalized taxon similar to the common ancestor

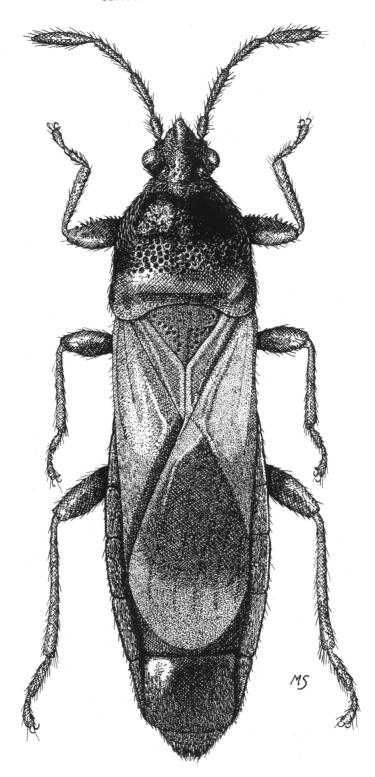


Fig. 69. Procellademus venenatus, dorsal view.

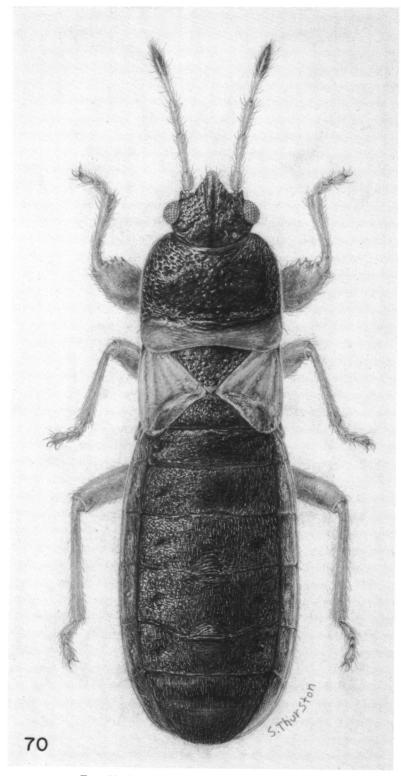


Fig. 70. Pseudoblissus trispinosus, dorsal view.

from which the more specialized *Aradademus* stocks have evolved. The compressed and produced prosternum anterior to the coxae is a unique condition in the Blissinae so far as I am aware.

RAMADADEMUS SLATER Figure 71

Ramadademus Slater, 1967, pp. 23-24.

Type Species: Ramadademus multispinosus Slater. By original designation.

DISTRIBUTION: Madagascar.

BIOLOGY: Unknown.

DIAGNOSIS: Large, very broad, strongly flattened. Metathoracic scent gland auricle elongate, strongly curving anteriorly, its margins rugulose (fig. 12C). All femora multispinose, or fore femora sometimes with a pair of large protruding spines that form a median groove for reception of tibiae. Fore tibiae curved, somewhat flattened; first tarsal segment very elongate, usually longer than segments two and three combined, second segment minute. Body completely shining above and below including corium and clavus, no pruinosity present. Apical corial margins straight. Membrane semitransparent, much thinner and obviously of different texture than corium. Fore coxal cavities closed. Ocelli small. Antennae slender and terete or nearly so. Clasper elongate with a narrow blade strongly curving toward apex, outer lobe short and thick, but tapering, inner lobe not strongly developed. Sperm reservoir broadly elliptical, basally stalked with thin nearly membranous distally broadened, somewhat tear-drop shaped lobes arising from distal end of bulb (may not be homologous with wings) (fig. 1HH).

KEY TO SPECIES OF RAMADADEMUS

few additional scattered setiferous spinules.

2. Anterior lobe of pronotum with a deep median longitudinal groove; fore femora lacking a close set series of setiferous spinules ventrally on posterior margin...sakalava Slater

2a. Anterior lobe of pronotum lacking a median longitudinal groove; fore femora ventrally possessing a closely set series of short acute setiferous spinules anomalous Slater

RETICULATODEMUS SLATER AND WILCOX Figure 72

Reticulatodemus Slater and Wilcox, 1966, pp. 70-71.

Type Species: Reticulatodemus calcar Slater and Wilcox. By original designation.

DISTRIBUTION: South America, Mexico.

BIOLOGY: Unknown.

DIAGNOSIS: Body moderately elongate, linear, non-flattened; metathoracic scent gland auricle sub-rectangular, little elevated above body surface (fig. 11A); fore femora armed below with one large spine; head and thoracic pruinosity variable: often with head pruinose laterally, broadly shining in central area; pronotum with a very broad posterior shining stripe and broadly shining in area of calli even across midline; males completely pruinose below, sometimes head and pronotum completely shining above and shining below on prothorax except mesally between fore coxae; scutellum pruinose laterally; apical corial margin straight; corium and membrane of different texture, latter thin, composed of minute hexagonal and quadrate cells. Fore coxal cavities closed; ocelli small: antennae terete. Antenniferous tubercles "hooked." Sperm reservoir with small bulb and strongly ventrally curved sublinear wings; claspers generalized. Spermatheca variable, usually with short pump.

KEY TO SPECIES OF RETICULATODEMUS

- 2a. Second antennal segment only twice length of segment 1 (31:15), considerably shorter than interocular space (30:38); pronotal calli

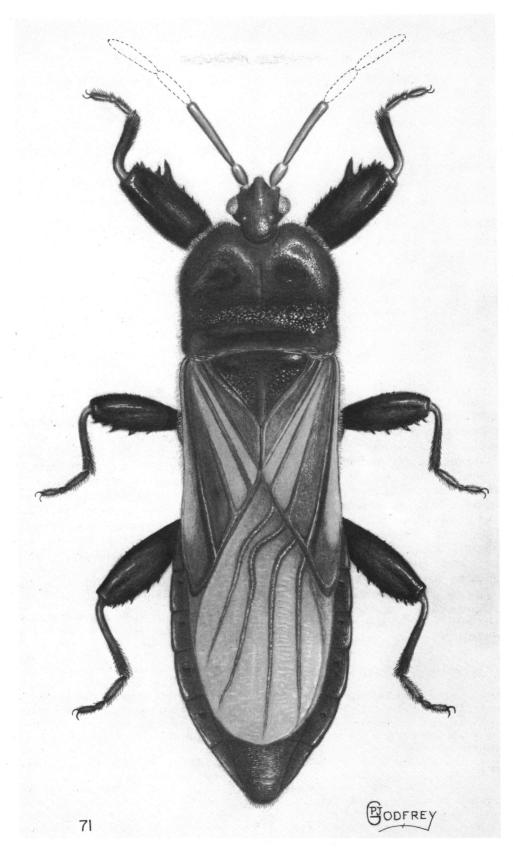


Fig. 71. Ramadademus multispinosus, dorsal view.

smooth, shining; membrane in large part testaceous, infuscated along veins but never

with a large complete discal spot (fig. 72)nitidus Slater and Wilcox

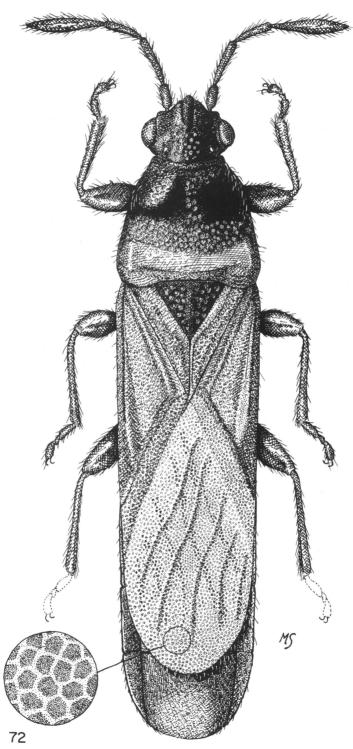


Fig. 72. Reticulatodemus nitidus, dorsal view.

- 3a. Eyes very narrow transversely, only very slightly extended beyond head curvature; length of second antennal segment greater than one-half width of head across eyes . . 4
- Width of head across eyes 1½ times as great as interocular width; with black "points" at caudolateral angles of abdominal connexiva 4-7.....orbiculoides Slater and Wilcox

RIGGIELLA KORMILEV Figure 73

Riggiella Kormilev, 1949, pp. 4, 6.

Type Species: Riggiella vianai Kormilev. Monobasic.

BIOLOGY: Collecting records from bamboo.

DISTRIBUTION: South America.

DIAGNOSIS: Body very broad, flattened. Metathoracic scent gland auricle elongate, not curving forward but enlarged at distal end (fig. 12B). All legs multispinose; no sexual dimorphism. Fore tibiae distally with hooked spines; first tarsal segment large, inflated, with a pad of hair below. Head and pronotum either almost completely shining with only anterior pronotal collar pruinose or with a narrow pruinose strip complete across transverse impression; pruinose below on prothorax except posteriorly and around fore coxae, head pruinose below; scutellum pruinose laterally with a broad shining central area; corium inconspicuously shin-

Ocelli small. Antennae terete. Short stubby genal "tusks" present. Ovipositor elongate. Clasper with elongate shaft and short blade. Sperm reservoir unique, apparently composed of a small distally "opened" bulb and two mi-

nute comma-shaped wings (fig. 1V).

ing from just mesad of raised radial vein to

lateral margin. Corium and membrane unitex-

tured, latter thick and opaque. Apical corial

margins straight. Fore coxal cavities closed.

KEY TO SPECIES OF RIGGIELLA

- 2a. Length of second antennal segment subequal to interocular space; length of hind tibiae considerably greater than that of fore femora; posterior margin of first valvifer nearly flatplanus Slater and Ahmad

SCANSIDEMUS SLATER AND WILCOX Figure 74

Scansidemus Slater and Wilcox, 1969, pp. 2-3.

Type Species: Scansidemus taprobanes Slater and Wilcox. Original designation.

DISTRIBUTION: Burma, Ceylon.

BIOLOGY: Unknown.

DIAGNOSIS: Body very broad, flattened. Metathoracic scent gland auricle elongate and strongly angled anteriorly near distal end. All femora multispinose. Head, pronotum both above and below and scutellum completely shining. Corium pruinose but with radial vein shining narrowly on basal three-fourths, costal margin also narrowly shining, membrane opaque, thickened. Fore coxal cavities closed. No sexual dimorphism. Almost straight apical corial margin. Terete antennae. Elongate ovipositor. Sperm reservoir with bulb reduced to a narrow median projection but with small straplike wings present.

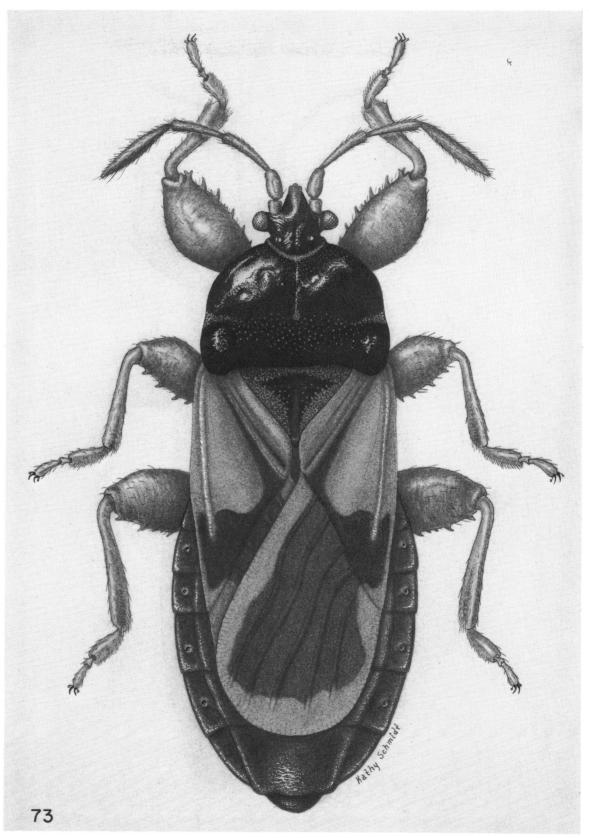


Fig. 73. Riggiella vianai, dorsal view.

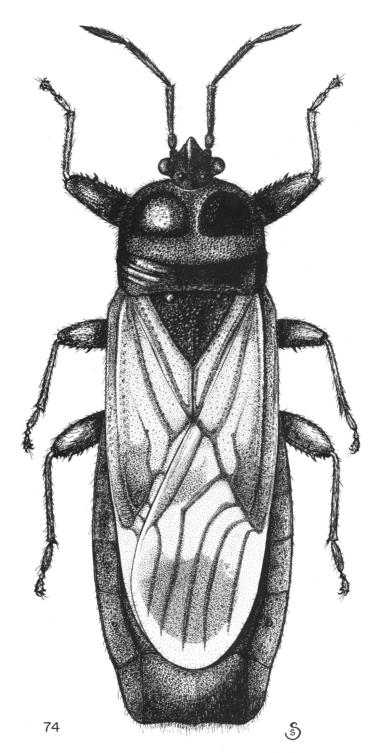


Fig. 74. Scansidemus taprobanes, dorsal view.

KEY TO SPECIES OF SCANSIDEMUS

..... Peregrinus Slater and Wilcox

SCINTILLADEMUS SLATER Figure 75

Scintillademus Slater, 1968, p. 284.

Type Species: Scintillademus gemmatus Slater. Monobasic.

DISTRIBUTION: New Guinea, New Britain. BIOLOGY: Unknown.

DIAGNOSIS: Body moderately broadened, somewhat flattened; metathoracic scent gland auricle elongate with a broad irregularly rounded distal end (fig. 11H). Fore femora with a large bifid ventral spine present and a deep subdistal excavation immediately beyond bifid spine, middle and hind femora mutic. Head and pronotum completely shining above, pruinose below and narrowly above acetabula, but pruinosity not attaining dorsal margin; scutellum pruinose laterally with a broad shining central area; corium largely shining until almost distal end of cubital vein and with an elongate shining stripe on clavus. Corium and membrane nearly unitextured, latter thick and opaque. Apical corial margin very strongly concave. Fore coxal cavities closed. Ocelli small. No marked sexual dimorphism. Antennae terete or moderatly clavate. Sperm reservoir with a small elliptical cup and large thumblike strongly divergent wings (fig. 1X). Ovipositor elongate.

SLATERELLUS DRAKE AND DAVIS Figure 76

Slaterellus Drake and Davis, 1959, pp. 24-25.

Type Species: Slaterellus hackeri Drake and Davis. Monobasic.

DISTRIBUTION: Australia.

BIOLOGY: Reported under bark of wilga and other trees.

DIAGNOSIS: Body short, stout, robust. Metathoracic scent gland auricle rounded, lobate. All femora mutic. Pruinosity absent both above and below, sub-shining, clothed with numerous flattened decumbent scalelike hairs. Apical corial margin somewhat concave. Membrane thin. membranous, much differentiated from thickened corium. Corium short and very stout. membrane with numerous anastomosing veins. Antennae short, stout, sublobate or beadlike. Fore coxal cavities closed. Ocelli small. Clasper rather plesiomorphic but with outer lobe located somewhat farther distad along shaft than in general. Sperm reservoir reduced to a small median scalelike flap (fig. 1A, B). Ovipositor not dividing abdominal sternum six.

DISCUSSION: This monotypic genus was originally described as a new sub-family, the Slaterellinae. Several authors have commented on its close affinities to the Blissinae but it appears to not previously have been formally placed in this subfamily. Despite its numerous apomorphic features *Slaterellus* is a true blissine in all essential features and the subfamily Slaterellinae becomes a junior synonym of the Blissinae.

SPALACOCORIS STÅL Figure 77

Spalacocoris Stål, 1874, pp. 129-130.

Type Species: Spalacocoris sulcifer Stål, 1874=Ischnodemus sulcatus Walker, 1872. Monobasic.

DISTRIBUTION: Southeast Asia, East Indies, Philippines.

BIOLOGY: Record of a specimen sucking the roots of a zingiberaceous plant.

DIAGNOSIS: Robust, elongate, subcylindrical, non-flattened. Metathoracic scent gland auricle elongate, grooved, curved strongly forward (fig. 10D), sometimes angulate. Fore femora enormously incrassate, multispinose; fore tibiae short and thick, enlarged and spined at distal end. Head and pronotum completely shining above and below; scutellum with a median shining elevation and lateral pruinosity. Corium with apical margin shallowly concave; corium

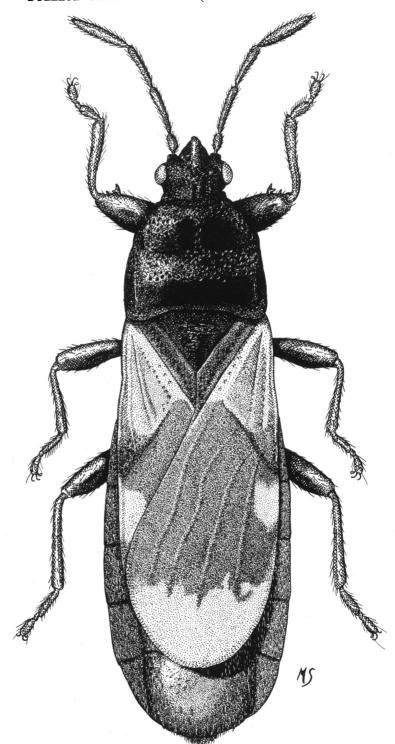


Fig. 75. Scintillademus gemmatus, dorsal view.

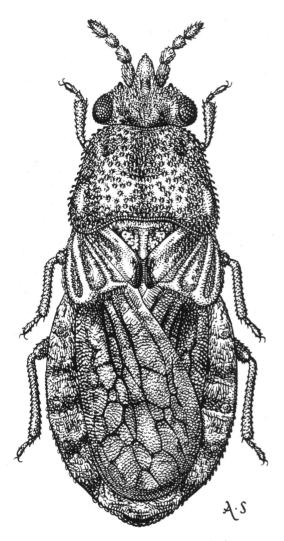


Fig. 76. Slaterellus hackeri, dorsal view.

two textured, area laterad of radial vein strongly shining, area mesad of radial vein pruinose, membrane thickened, opaque, similar in texture to corium. Fore coxal cavities closed. Ocelli very large. Genae somewhat swollen and produced. Pronotum with posterior lobe strongly produced caudad. Labium very short, not attaining fore coxae. Antennae clavate. Parameres blocklike without differentiated basal area and shaft. Sperm reservoir reduced to a distally tapering median projection. Ovipositor not laciniate, reduced to broad blocky plates. Spermatheca with bulb large, bearing a con-

spicuous basal flange; pump short and broad especially at distal end.

KEY TO SPECIES OF SPALACOCORIS

- Scent gland auricle sharply angulate; pronotum lacking a median longitudinal trough; basal width of pronotum greater than median lengthrufusculus Slater and Ahmad
- 3. Larger, generally 12-15 mm. long; claspers strongly curved in the middle; apical portion of sperm reservoir broad and arrowhead shaped (fig. 77)......
- 4. Large species, generally 13.0-16.0 mm. long; all legs uniformly dark reddish brown; claspers pointed at apices; apical portion of sperm reservoir small, knoblike and blunt; anterior inner margin of scent gland auricle truncated nigritus Slater and Ahmad
- 4a. Smaller species (10.83 mm.); fore femora dull, yellowish; claspers rounded at apices; apical portion of sperm reservoir large and broadly rounded at apex; anterior inner margin of scent gland auricle rounded . . . sulcifer Stål

TALPOBLISSUS SLATER AND WILCOX Figure 78

Talpoblissus Slater and Wilcox, 1973, p. 94.

Type Species: *Blissus cydnoides* (Slater). By original designation.

DISTRIBUTION: Africa, India.

BIOLOGY: Presumably live on grasses.

DIAGNOSIS: Body short, stout, thick, robust. Metathoracic scent gland auricle elongately rounded, earlike. Fore femora strongly incrassate, armed below on distal third with a single conspicuous sharp spine. Fore tibiae strongly expanded distally, concave and armed marginally with a series of stout spines to form a large fossorial structure (fig. 7G, H). Head and pro-

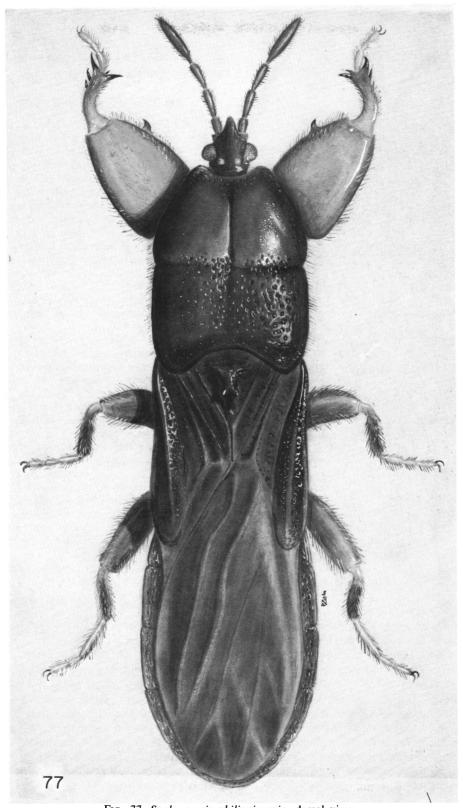


Fig. 77. Spalacocoris philippinensis, dorsal view.

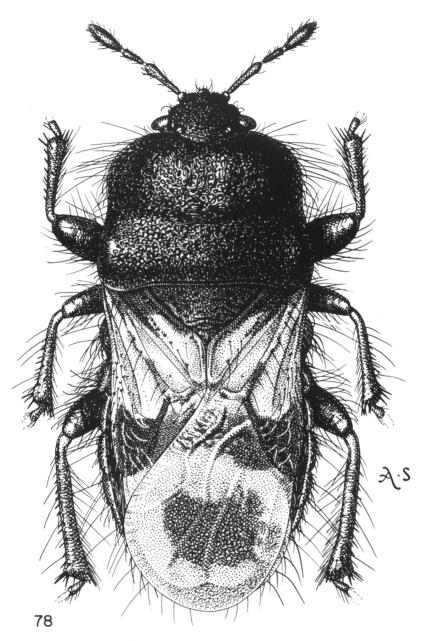


Fig. 78. Talpoblissus cydnoides, dorsal view.

notum above shining, non-pruinose; laterally propleuron lacking pruinosity above acetabulae, latter shining; prosternum with a narrow pruinose area anteriorly mesad of acetabula. Scutellum completely pruinose. Apical corial margins strongly concave. Membrane unitextured, opaque but thinner than adjacent corium.

Fore coxal cavities open. Ocelli small. Wing microptery and brachyptery occur. Second and third antennal segments strongly clavate. Body and appendages thickly clothed with very elongate upstanding hairs. Pronotum subquadrate in shape, nearly as broad across calli as across humeri. Eyes strongly transverse with

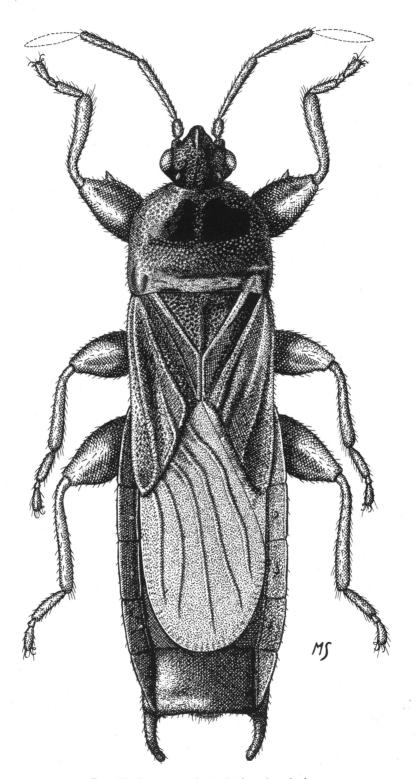


Fig. 79. Toonglasa forficuloides, dorsal view.

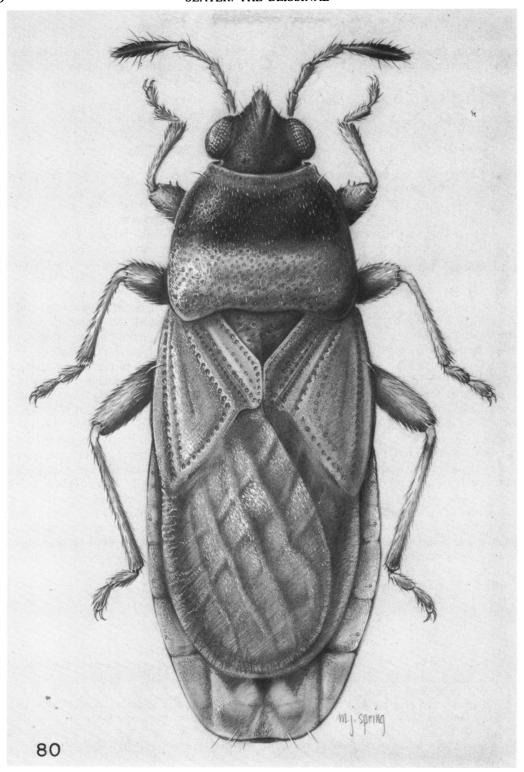


Fig. 80. Xenoblissus lutzi, dorsal view.

posterior margins concave. Claspers generalized; sperm reservoir with large cup and straplike slender wings (fig. 1P).

KEY TO SPECIES OF TALPOBLISSUS

TOONGLASA DISTANT Figure 79

Toonglasa Distant, 1893, pp. 391-392.

Type Species: Toonglasa forficuloides Distant. Monobasic.

DISTRIBUTION: Mexico. BIOLOGY: Unknown.

Diagnosis: Large, broad, robust, somewhat flattened. Fore femora strongly incrassate armed below with a large prominent spine; middle and hind femora mutic. Head pruinose. pronotum above pruinose with shining areas across humeri and a pair of large quadrate shining patches in calli area; completely pruinose below. Scutellum pruinose with a shining median elevation. Apical corial margin straight. Membrane relatively opaque, nearly as thick as adjacent corium. Submacropterous. Fore coxal cavities closed. Ocelli small. Antennae terete or slightly enlarged at distal ends of antennal segments two and three. Seventh abdominal segment with a pair of elongate curving prominent projections extending posteriorly from each lateral angle. Details of genitalia unknown.

Known only from the holotype from "omilteme in Guerrero 8,000 feet." It is obviously a highly apomorphic derivative from an Extarademus-like ancestor.

XENOBLISSUS BARBER Figure 80

Xenoblissus Barber, 1954, p. 223.

Type Species: Xenoblissus lutzi Barber. Monobasic.

DISTRIBUTION: South America.

BIOLOGY: Unknown.

DIAGNOSIS: Body short and stout, non-flattened. Metathoracic scent gland auricle rectangular, little raised above body surface, angled caudolaterad (fig. 11E). Fore femora with a pair of small spines below appearing to almost arise from a single base, mid and hind femora mutic. Head and pronotum subshining above completely non-pruinose; pruinose areas well developed on prosternum and on anterior half of propleuron, pruinosity also extensively present on mesopleuron above acetabulae and on mesosternum and metasternum; scutellum pruinose laterally. Thickened scalelike hairs present on head and pronotum. Apical corial margin moderately sinuate. Corium thicker than membrane; membrane cellular and reticulate (sometimes faintly so). Fore coxal cavities closed. Ocelli small. Antenniferous tubercles hooked. Antennae clavate. Sperm reservoir unique with large elliptical cup, wings straplike moderately elongate lying in a membranous ellipse arising from distal third of bulb (fig. 1Y). Very elongate ovipositor reaching abdominal sternum five.

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